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A REVIEW AND EVALUATION OF THE AMPHIBIAN TOXICOLOGICAL LITERATURE

A. Harfenist
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ABSTRACT

The impacts of environmental contaminants on amphibian populations have not been extensively studied. Amphibians, with the exception of a few specialized species, utilize both the terrestrial and aquatic ecosystems during their life cycles.

This report reviews the published literature on the effects of environmental contaminants on amphibians. Acute toxicity studies, other laboratory studies, field studies and residue data are summarized in Tables 1, 2, 3, and 4 respectively.

Amphibians are particularly sensitive to metals and to acidification. They are considered to be useful indicator species for measuring the effects of local changes in environmental studies. Several species are readily reared in the laboratory so that experimental studies can be undertaken.

RÉSUMÉ

Les répercussions des contaminants sur les populations amphibies n'ont pas été étudiées à fond. Les amphibiens, à l'exception de quelques espèces spécialisées, ont besoin des deux écosystèmes durant les étapes de leur vie.

Ce rapport passe en revue les écrits sur les effets des contaminants environnmentaux sur les amphibiens. Des études de pointe sur la toxicité, différentes recherches de laboratoire, des observations sur le terrain et des données sur les résidus sont relevés sommairement dans les tableaux 1, 2, 3 et 4 respectivement.

Les amphibiens sont particulièrement vulnérables aux métaux et à l'acidification. Ils sont considérés comme des sujets témoins tout indiqué pour mesurer les effets des changements locaux dans les études environnementales. On peut facilement en élever plusieurs espèces en laboratoire à des fins de recherches expérimentales.

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I INTRODUCTION

The impact of environmental contaminants on amphibians is a subject that has received limited attention. This review is an attempt to present the toxicological data that exists. The majority of the toxicological research conducted using amphibians is centered around the anurans and urodeles. The apodans, due to a lack of knowledge concerning the biology and secretive lifestyle have not been studied in relation to toxic chemicals.

Amphibians, with the exception of a few specialized species, require moisture in some form, whether it be a permanent lake or leaf that has collected dew, in order to complete their life cycle. It is this requirement and their subsequent metamorphosis to the combined terrestrial/aquatic adult stage that is the major route of exposure for amphibians to toxic contaminants. Thus, in order to fully comprehend the possible implications of environmental contaminants on amphibians a brief discussion of their life cycle is necessary.

Amphibians fall into three general categories with respect to their breeding habitats. The first category is composed of those that breed in permanent water; the second breed in temporary or seasonal pools; and the third group breeds out of the water (Porter 1972). The latter group still requires a moist location to deposit its eggs in order to ensure the development of the embryos. Thus all amphibians rely upon seasonal and geographical patterns of precipitation which govern their distribution (Romer 1959, Porter 1972). In general, the amphibians of North America breed in spring or early summer (Romer 1959, Porter 1972). The rate of development of the eggs and embryos is species dependent. Eggs deposited in seasonal pools or other moist areas develop more quickly to avoid dessication. Eggs laid in permanent water bodies need not develop as fast due to the unlikelihood of dessication (Porter 1972).

Anurans congregate in leks in a pond where the males call to attract reproductively active females. The female, upon hearing the mating call of her species, will move in the direction of the call. The male mounts the female and clasps her inducing the deposition of the eggs, which are immediately fertilized. The number of eggs deposited by a female during a breeding season, ranges from a single egg (Genus Sminthillus) to 30,000 or more (Genus Bufo) (Porter 1972). Anuran egg masses are of three generalized types and positions enabling them to maximize oxygen availability in different habitats and water temperatures. One type is deposited as a flattened egg mass at the surface of the water such as those of R. catesbeiana; the second type is globular and attached to submerged vegetation as in P. triseriata egg masses; and the third type is characteristic of the true toad species (Genus Bufo), and are deposited in elongated strings (Otto and Towle 1947, Romer 1959, Porter 1972). Depending upon the species, the eggs take between 2 and 30 days to hatch.

Other species exhibit more specialized breeding behaviour. Some female anurans protect their eggs by carrying them on their backs or under the skin. Others build nests while some lay eggs enveloped in a gelatinous material that hardens and reduces moisture loss. For the most part these specialized adaptations are found in amphibians inhabiting the tropics, while the generalized life cycle described here is more characteristic of the species located in North America (Villee et al. 1958).

The embryo hatches out as a larvae equipped with a well developed tail for swimming. This aids it in foraging for submerged vegetation upon which it browses (Villee et al. 1958, Romer 1959, Porter 1972). The anuran larvae respire by gulping water and forcing it over the gills located in the future neck region. Some gaseous exchange is also accomplished through the skin (Romer 1959, Porter 1972). The larvae or tadpoles transform to the adult stage via metamorphosis. This occurs two weeks to three years after hatching, depending upon the species (Porter 1972).

Metamorphosis involves radical structural changes of the body: the gills disappear, lungs and limbs develop rapidly (hind limbs first), the tail is absorbed and the digestive tract shortens. Eventually the adult frog becomes an air breather (Romer 1959). Respiration is accomplished via internal lungs as well as via the skin.

The length of the metamorphic period is also species dependent. In the case of the Spadefoot toad (Scaphiopus holbrookii), metamorphosis into a terrestrial toad takes only twelve to thirteen days from fertilization. Spadefoot toads breed in temporary pools and must develop quickly to avoid dessication (Porter 1972). Some species require a year, (R. catesbeiana) (Cecil and Just 1979), while others require still more time. Several amphibian species are non-metamorphosing and retain both larval characteristics and their aquatic lifestyle (genus Necturus) (Porter 1972).

Adult anurans, unlike the larvae, are carnivorous, consuming almost any moving prey of the right size. This includes worms, insects, other amphibians, reptiles and occasionally mammals. Throughout its life cycle an anuran can occupy several different positions within the food web (Curtis 1968, Porter 1972).

The urodeles also follow a general pattern of breeding behaviour. During courtship the male either clasps or blocks the path of the female and begins lashing with his tail, bumping or rubbing to stimulate the female. The male then deposits a spermatophore (a capsule containing sperm). The female either follows the male or is pulled by him into a position where she can pick up the spermatophore

with the cloaca, where it is stored (Romer 1959). The eggs are fertilized internally and may be deposited in decaying vegetation, logs, pools or other moist environments. The eggs can take up to two months to hatch. Aquatic salamander larvae differ from anuran larvae in that they develop adult appearance quickly, they possess external gills and their diet is primarily insectivorous (Romer 1959). After several months and under the correct environmental conditions the larvae transform into the adult stage and may leave the water. Some species have lungs whereas the Plethontidae, known as the "lungless salamanders", respire via the skin. Another group, the Necturidae, are neotonic, retaining their external gills and aquatic lifestyle even after sexual maturity.

Adult urodeles, like adult anurans, are carnivorous. Their diet consists of arachnids, annelids, insects, amphibians, small fish and their eggs, and in the case of Cryptobranchus, mammals. Adult urodeles inhabit moist locations under logs, rocks, and leaf litter, returning to the water in spring to breed (Porter 1972).

Interruptions of this life cycle by toxic contaminants can have disastrous effects on amphibian populations (Paulov 1977). Amphibians are of significant economic importance to man. Due to their carnivorous adult stage they consume insect pests harmful to crops (Schwabe 1977). Some cultures use amphibian skins for leather goods, amphibian venom for hunting and medicines, while frogs legs (R. catesbeiana) are considered a delicacy by some humans. Amphibians are also the perennial biological specimen in institutions due to their relative abundance and availability (Otto and Towle 1947). Several species have actually benefitted from man's agricultural practices by taking advantage of irrigation ditches and new habitats that have been created, but many more species have been detrimentally affected by habitat destruction and the use of toxic chemicals. Amphibians are not only important to man, but also to other animals within the community. They make major contributions to community biomass (Burton and Likens 1975a, 1975b, Cecil and Just 1979, Debendictis 1974), as well as playing significant roles in competitive and predator-prey relationships (Orser and Shure 1972, Burton and Likens 1975b, Lynch 1979). It is of interest to note that amphibians are the only class of vertebrates that do not include any pests or species harmful to man. Amphibians do not compete for harvests, nor are there any ferocious or destructive species (Porter 1972). Even so amphibians are victimized by man's attempts to control pests with toxic chemicals, primarily insecticides and herbicides (Fashingbauer 1957, Hazelwood 1969, Porter 1972, Cooke 1973a, Curtis 1968).

The toxicity of these contaminants to amphibians in lab bioassays and field experiments as well as their general effects on populations, will be reviewed in order to identify specific substances that pose

potential hazards to amphibians. From this information an evaluation will be made as to whether amphibians can be employed as useful indicators of environmental quality.

II INSECTICIDES

a) ORGANOCHLORINES

ALDRIN AND DIELDRIN

Short-Term Effects

In static bioassays dieldrin was highly toxic to B. woodhousii tadpoles with a 96 hour LC50 of 0.15 mg/L (Sanders 1970) (Table 1).

In static bioassays, complete mortality of adult R. cyanophlyctis (an Asian frog species) occurred within 215 minutes of exposure to 0.006 mg/L aldrin (Rane and Mathur 1978). Although sample sizes were small (2 to 4 frogs per treatment with 6 different treatments) no frogs survived longer than 120 minutes when exposed to 0.125 mg/L (the highest concentration tested) (Table 2). Preceding death, the frogs swam erratically exhibiting a loss of equilibrium. A coagulation of mucous occurred over the body with the belly becoming bloated and the skin turning pale (Ibid 1978). These results indicate that aldrin is very toxic to the adults of R. cyanophlyctis and that the 96 hour LC50 would be much lower than 0.006 mg/L considerably less than the 96 hour LC50 reported for B. woodhousii by Sanders (1970) (Table 1).

Mulla (1962, 1963) found R. catesbeiana tadpoles experienced 100% mortality within 24 hours when maintained in ponds and treated with 0.1 kg/ha dieldrin (Table 3). Since the actual concentrations of dieldrin in the ponds were not measured it is difficult to compare these results with other studies.

Long-Term Effects

Short-term exposures to dieldrin can have long-term effects on developing amphibians. When groups of 150 Limnodynastes tasmaniensis (an Australian frog) embryos were exposed to 0.0, 0.01 and 0.1 mg/L dieldrin for 7 hours, the 0.1 mg/L exposure resulted in accelerated growth and abnormalities later in development (Brooks 1981) (Table 2). No mortality occurred during the exposure, but 21 of the tadpoles treated with 0.1 mg/L dieldrin as eggs, exhibited some degree of deformity later in development. Nineteen days after exposure some

severe cases had cephalic pigmentation disruption resulting in two large transparent spots on the head of the tadpole. These spots were swellings situated over the otolith which was also abnormal. The affected tadpoles also had constricted guts indicating lack of feeding. In general the tadpoles exposed to 0.1 mg/L tended to feed less and frequently rested on the bottom of the aquaria (Ibid 1981).

At a concentration of 0.30 ppm aldrin in solution, R. pipiens suffered 40% mortality after thirty days (Kaplan and Overpeck 1964) (Table 2). In the same study at 0.10 ppm dieldrin in solution the frogs suffered 50% mortality after thirty days (Table 2). Neuromuscular changes were evident in all insecticide concentrations studied, but were more pronounced in aldrin and dieldrin. These changes included coarse tremors, rigidity, abnormal reactions to stimulation, convulsions and a reduction in the ability to orient themselves (Kaplan and Overpeck 1964).

Vinson et al. (1963) examined a number of populations of A. crepitans and A. gryllus to determine if these species had developed cross-resistance to aldrin after being exposed to DDT, toxaphene, methyl parathion and endrin. Six populations of A. crepitans from six localities in Mississippi were chosen, with three localities being free of pesticides and three heavily treated with pesticides in the past fifteen years. Adult frogs were exposed to 1 ml of either 30,000 mg/L or 50,000 mg/L aldrin. The aldrin was pipetted onto filter paper and then the adult frog was placed on the paper for 36 hours. For A. crepitans, mortality varied from 0% at both 30,000 and 50,000 mg/L for individuals from populations that had been treated with other pesticides, to 30% mortality at 30,000 mg/L, and 57% mortality at 50,000 mg/L for individuals from populations that had never been treated with other pesticides. These results indicate a cross-resistance to aldrin which was thought to have occurred by natural selection for a physiological mechanism to confer resistance to pesticides. This resistance was non-specific enough to be effective against aldrin (Ibid 1963) (Table 2).

Residues

Aldrin can be bioaccumulated by amphibians and passed up the food chain (Korschgen 1970). Korschgen (1970) measured aldrin residues in soils, plant and animal tissues from a Missouri corn field that had been treated with aldrin at 1 kg/ha for at least 15 years. Adult B. americanus had 0.03 mg/kg aldrin (wet weight) although young of the year had no measurable aldrin residues. The garter snake (Thamnophis sirtalis), a species that preys upon B. americanus, had very high residue levels ranging from 10.3-14.4 mg/kg indicating that residues bioaccumulate as they are passed up the food web (Ibid 1970) (Table 4).

Niethammer et al. (1984) also found evidence of accumulation up the food chain in an area of Louisiana characterized by oxbow lakes. This area had been subjected to heavy organochlorine pesticide use due to the fertile alluvial soil in the area. Ninety-five percent of all animals tested including R. catesbeiana, R. pipiens and the Bronze frog had residues of one or more organochlorine insecticides (Table 4). Ohlendorf et al. (1981) reported a die-off of herons (Ardea herodias) probably due to insecticide poisoning. Frogs represent a large percentage of a heron's diet and thus, even though frogs appear to be able to mobilize and excrete these insecticides, there is the danger of serious effects occurring through bioaccumulation (Niethammer et al. 1984).

Collections from agricultural areas in Iowa revealed residues of dieldrin and other organochlorine insecticides in the viscera of B. bufo and R. pipiens (Punzo et al. 1979) (Table 4). The values were low or absent due to the method of analysis employed and thus may not represent accurate values contained in the amphibians from this area (Punzo et al. 1979).

Aldrin and dieldrin's moderate water solubility was used by Kaiser and Dunham (1972) to explain the passage of these two insecticides through R. pipiens skin, explaining the rapid excretion of these compounds by amphibians.

Mechanism for Toxicity

The primary site of action of aldrin-transdiol (the active metabolite of dieldrin) is somewhere in the central nervous system (Akkermans et al. 1975b). In preparations of spinal cords of X. laevis, Akkermans et al. (1975b) measured an augmentation of polysynaptic reflex activity and a reduction of orthodromic postsynaptic inhibition in dieldrin poisoned toads. Direct application of aldrin-transdiol to spinal cord preparations resulted in a potentiation of spinal reflex activity and a marked reduction of spinal inhibitory mechanism (Table 2). These excitatory effects were followed by a reduction of spinal excitability. Similar effects have been measured on the motor end-plate of the sartorius nerve with increased synaptic transmission (Akkermans et al. 1974) (Table 2).

Akkermans et al. (1975a) examined the effect of dieldrin and aldrin-transdiol on the sense organs of X. laevis. These two substances failed to induce any sign of repetitive activity (Table 2). Aldrin-transdiol caused an increase in the rate of firing of the lateral-line organ followed by a complete blockage of this firing (Ibid 1975a).

Webb et al. (1979) examined the short circuit current and resistance in isolated intact R. pipiens skin to determine if dieldrin was affecting the sodium balance. Dieldrin caused a significant increase in short circuit current while resistance was decreased. These results were interpreted as indicating that dieldrin was producing an increase in permeability of the outer skin membrane (Ibid 1979) (Table 2).

CHLORDANE

Long-Term Effects

Kaplan and Overpeck (1964) examined the survival of R. pipiens adults in response to varying concentrations of chlordane. At a concentration of 0.50 ppm, 40% of the frogs were dead after thirty days (Table 2). Some of the symptoms of poisoning were neuromuscular changes, excessive thrashing and tremors (Kaplan and Overpeck 1964).

Field studies to examine the potential threat of chlordane to amphibians were conducted by Mulla (1962, 1963). When applied at 0.56 kg/ha a.i. tadpoles in the pond reached maximum mortality within 48 hours (30%) (Mulla 1963) (Table 3). At a lower application (0.11 kg/ha a.i.) no mortality was observed (Ibid 1963).

Residues

Up to 1,036 days after a lake in British Columbia was sprayed with technical grade chlordane at 0.010 mg/L, residue levels were measured in T. torosa (Albright et al. 1980). Residues were high within the first 14 days (8.9 ppm mean whole body without liver and stomach, 26.3 ppm liver, 2.2 ppm stomach) (Table 4). After 1,036 days residue levels declined to 0.2 ppm body without liver and stomach, 0.3 ppm liver, and only a trace in the stomach. Up to 93 days after treatment residue levels in T. torosa were much greater than in the water (Table 4). Of the chlordane constituents analyzed trans-nonachlor was most persistent. Although trans-nonachlor was present in the technical grade chlordane at 5.26% it accounted for 10-17% of the residues in newts after 14 days. Residue levels in T. torosa were compared with Salmo clarki and it appeared that the newts were more capable of metabolizing and eliminating chlordane residues (Ibid 1980).

Mechanism for Toxicity

Several chlorinated hydrocarbon insecticides have been shown to inhibit ATPase enzymes including those activated by sodium and potasium (Webb et al. 1979). Webb et al. (1979) measured sodium (Na^+) transport by measuring the short-circuit current across R. pipiens skin. At 2 X

10^{-4} M a- chlordane and y-chlordane produced no significant change in the short-circuit current and thus it was concluded that sodium transport is not severely affected by chlorinated hydrocarbons based on suggested application rates (Webb et al. 1979) (Table 2).

CHLORDIMEFORM

Short-Term Effects

Chlordimeform was used extensively in Japan against rice stem borer larvae. Most of the research conducted on amphibians has been in physiological or biochemical studies to elucidate the mechanism of toxicity (Martin 1968, Wang et al. 1975).

Watanabe et al. (1975) conducted toxicity experiments by injecting adult R. nigromaculata with chlordimeform. At 300 mg/kg, 80% mortality was observed, and at 100 mg/kg no mortality was recorded (Table 2). Watanabe et al. (1975, 1976) also showed that chlordimeform at concentrations of 10^{-4} to 10^{-3} M causes the contraction of the rectus abdominus muscle (Table 2).

Mechanism for Toxicity

The effect of chlordimeform on the muscle of the frog is the suppression of contraction induced by acetylcholine and potassium. This occurs due to a blockage of neuromuscular transmission by the depression of motor end-plate sensitivity to the transmitter (Wang et al. 1975, Watanabe et al. 1975, 1976) (Table 2).

DDT

Short-Term Effects

In static bioassays with B. woodhousii tadpoles and P. triseriata tadpoles, DDT was one of the least toxic insecticides tested, resulting in 24 hour LC50s of 2.4 mg/L and 1.4 mg/L respectively (Sanders 1970) (Table 1). Examination of DDT toxicity to B. woodhousii at different lifestages indicated an increasing sensitivity with age with the most sensitive stages being tadpoles at 3 to 7 weeks of age. The 24 hour LC50 for two-three week old tadpoles was 5.3 mg/L but by seven weeks this had dropped to 1.4 mg/L (Table 1). The increased sensitivity to DDT was thought to be related to developmental changes associated with metamorphosis. These changes include the disappearance of external gills as the internal lungs begin to function; an increase in thyroid activity leading to an increased rate of metabolism, the alteration in the permeability and ion transport of the integument and an increase in weight (Ibid 1970).

In contrast Marchal-Segault (1976) reported DDT to be slightly more toxic to B. bufo tadpoles with the 95% confidence limit of the 24 hour LC50 lying between 0.7 and 2.3 mg/L, although some mortality occurred at concentrations as low as 0.03 mg/L (Table 1). This worker confirmed the finding of Sanders (1970) that the sensitivity of tadpoles diminished after metamorphosis.

R. temporaria tadpoles exposed to 0.1 to 10.0 mg/L DDT exhibited a lack of coordination and hyperactivity. This behaviour returned to normal when the tadpoles were removed from the test medium, older tadpoles recovering more rapidly. At concentrations of 1.0 and 10.0 mg/L the tadpoles suffered mortality of 20 and 60-70 percent respectively (Table 2). In all tadpoles, growth was reduced and large tadpoles frequently surfaced to breathe, sometimes developing a hole in the snout between the nostrils and upper mandible (Cooke 1970, 1979). These abnormal behaviours after exposure to DDT were in part dependent on tadpole size and density. Small tadpoles at high density (50 tadpoles/L) exhibited more severe effects than larger tadpoles reared at a density of 10 tadpoles/L (Cooke 1979).

R. temporaria adults force-fed DDT dissolved in olive oil had a 96 hour LD50 of 7.6 mg/kg (Harri et al. 1979) (Table 1).

Ellis et al. (1944) reported one hundred percent mortality after injections of 150 mg/kg of DDT. Some mortality was observed following injections of as little as 10 mg/kg (Table 2).

Treated frogs displayed hyperexcitability, tremors, and a lack of locomotion and co-ordination. Frogs that survived for at least 10 days, lost all observable signs of poisoning and their behaviour returned to normal (Harri et al. 1979).

Issacson (1968) conducted experiments to examine peripheral nervous system involvement in the behaviour associated with DDT intoxication (Table 2) and found R. pipiens exhibited increased activity until the time of death. He theorized that an increase in activity of this type would place the frogs at a selective disadvantage. In studies exposing DDT intoxicated R. temporaria tadpoles to a predator Triturus cristatus, the newt made significantly more lunges at hyperactive tadpoles (Cooke 1971) (Table 2).

Mortality of amphibians following field applications of DDT varies and is dependent upon the amount of chemical, the weather at the time of application, the depth of pond, mode of application and the amount of canopy cover. These parameters render field results difficult to compare (Herald 1949, Cooke 1973b, Pearce and Price 1977). In field applications of DDT to ponds, 0.11 kg/ha caused no mortality of R. catesbeiana tadpoles (of varying sizes and stages) whereas 1.0 kg/ha

caused 80% mortality within 48 hours (Mulla 1963) (Table 3). In Minnesota, Fashingbauer (1957) reported a serious reduction in a local population of R. sylvatica following an aerial spray operation aimed at the reduction of forest tent caterpillars (Table 3). He noted no immediate adverse effects, but 60 hours after the application discovered large numbers of dead frogs. It was not known if the accumulation of DDT via the caterpillars or contact with the surface film caused the mortality but 94% of frogs had ingested caterpillars. Routine spraying of a forest in Georgia with 0.11 and 0.56 kg/ha DDT resulted in some tadpole and frog mortality (Tarzwell 1950) (Table 3).

Long-Term Effects

R. temporaria tadpoles exposed to 0.001 mg/L DDT exhibited uncoordinated hyperactive behaviour during the period day 5 to day 8 post-exposure (Cooke 1970, 1971, 1972a). The onset of the behaviour coincides with DDT levels having reached 2 ppm in the tissues. These hyperactive tadpoles developed at a slower rate than the controls, presumably due to the elevated pesticide residues in their tissues (Cooke 1973a) (Table 4).

When R. temporaria tadpoles were exposed to 0.0001 mg/L they behaved normally even though, from day 15 onward, the tadpoles and small frogs contained between 2 and 5 ppm DDT (Table 4). This is in contrast to tadpoles receiving an acute dose of DDT, which resulted in hyperactivity when tissue levels reached 2 ppm (Cooke 1973a). An acute dose can result in high residue levels in the blood and thus a build up in the nervous system, causing hyperactive behaviour. The slower rate of intake induced by chronic exposure, results in lower residue levels in the blood and does not cause an overload (Cooke 1973a). Differences in response to acute and chronic dosing were considered to be related to the storage of residues in the fatty tissues (eg. tail) (Cooke 1973a).

Long term effects of DDT in the field were examined by Pearce and Price (1977) in New Brunswick forests (Table 3). Substantial residues were discovered in four species of amphibians while other species had lower levels. No long-term effects on the populations of these amphibians were reported.

The effect of DDT on tail regeneration of R. pipiens and R. catesbeiana was examined by Weiss (1975). The two species exhibited both different rates of regeneration and vulnerability to DDT in terms of direct mortality. R. pipiens experienced slower and retarded tail regeneration at 0.005 ppm with 30% mortality after 12 days, while regeneration in R. catesbeiana at this concentration, was not different from controls (Table 2). At 0.025 ppm, R. pipiens suffered 95% mortality and retarded tail regeneration after 12 days. R. catesbeiana

experienced 60% mortality and a reduced regenerative capability at this concentration. Weiss (1975) suggested that a retardation of regeneration may increase the susceptibility of the organism to predation, causing an increase in the concentration of a contaminant at higher levels in the food chain.

Residues

A large portion of the research into residues in amphibians has centered around DDT. The level of residues accumulated in amphibians is dependent upon a number of factors. One factor is the stage of development during exposure. Cooke (1972a) was unable to detect DDT residues in R. temporaria, B. bufo, and Triturus vulgaris spawn after 24 to 48 hour exposures to DDT (Cooke 1972a). R. temporaria and B. bufo tadpoles exposed to DDT prior to or during hind limb bud development, had tissue concentrations of 2-3 and 3-4 mg/kg respectively. During tail resorption the frog was susceptible to levels that had been accumulated and stored during exposure at the larval stage. In general R. temporaria was more sensitive to DDT than B. bufo, indicating species differences in the ability to accumulate residues (Cooke 1972a) (Table 2).

The absolute amount of DDT taken in by R. temporaria tadpoles exposed to 0.1, 1.0 or 10.0 mg/L DDT was independent of size. Smaller tadpoles had higher tissue content of DDT, and higher mortality than larger individuals (Cooke 1970).

Licht (1976a) studied the effect of length of exposure and temperature on accumulation. Subacute exposure of R. sylvatica tadpoles for 7 days to 0.001 to 0.002 mg/L DDT reached maximum tissue levels in 24 hours (Licht 1976a) (Tables 2 and 4). After 24 hours exposure to 0.003 mg/L DDT at 15°C the liver had the highest level with 27.04 mg/kg (Table 4) while whole body residues were 0.7 mg/kg. These levels declined to less than 18 mg/kg in the liver and 0.6 mg/kg whole body after 80 hours. When treated with 0.003 mg/L DDT for 7 days R. sylvatica maintained higher DDT levels for longer at 15°C than tadpoles at 21°C (Licht 1976a) (Table 4).

Feeding also influences DDT accumulation. Fed R. temporaria adults had lower DDT residues than starved frogs in tissues other than fat (Harri et al. 1979). Loss of DDT residues from body tissues can vary depending upon previous exposure to DDT and the physical environment (Harri et al. 1979, Licht 1979). Previous exposure to DDT can increase the rate of loss of DDT after re-exposure in R. temporaria adults (Harri et al. 1979). It was speculated that previous DDT exposure leads to increased activity of drug-metabolizing enzymes that increases DDT excretion (Ibid 1979). R. sylvatica adults treated as tadpoles for 1 week with 0.003 mg/L DDT lost DDT much more slowly when maintained in

an aquatic environment for 3 to 4 weeks than adults maintained in a dry terrestrial environment (Licht 1976a, b, Licht 1979). DDT in the fat of the terrestrial adults was 37.4 mg/kg compared to 4.93 mg/kg in the aquatic tadpoles (Ibid 1976b). Adult R. temporaria, however, were found to eliminate DDT at the same rate in both aquatic and terrestrial environments (Harri et al. 1979).

Niethammer et al. (1984) found frogs from a contaminated area in Louisiana had low levels of DDT and suggested that this was due to efficient excretion and their mobility, which allows them to flee unsuitable areas (Table 4).

Finley and Pillmore (1963) found traces of DDD in R. pipiens in an area where only DDT had been sprayed. This suggests that DDT is converted to DDD by animal tissues and thus possibly establishes another route of excretion.

Residues vary greatly in different parts of the body (Harri et al. 1979, Licht 1976a, b). Fat had the highest residue concentration in R. sylvatica tadpoles exposed to 0.001 to 0.003 mg/L DDT for 7 days with up to 60 mg/kg, the liver had up to 20 mg/kg while the heart, head, gut and lungs contained only a trace (Licht 1976b). R. temporaria also had maximum DDT residues in fat, tissues with high concentrations of fat and the liver (Harri et al. 1979).

DDT residues have been correlated with mortality and behavioural and developmental abnormalities (Cooke 1970, 1972b, 1973a, Harri et al. 1979). The LD50 for adult R. temporaria was 7.6 mg/kg (Harri et al. 1979) (Table 1). Chronic dosing of R. temporaria tadpoles in the field and lab both showed that residues up to 5 mg/kg DDT resulted in mild hyperactivity (Cooke 1973a) (Table 2). Acute doses of 2 to 5 mg/kg caused severe hyperactivity, weight loss and delay in development (Cooke 1970, 1972b, 1973a).

Size may also influence the severity of effects from residues. Cooke (1979) found R. temporaria tadpoles of the same age, but larger weight ($x = 689 \pm 44$ mg) had no behavioural abnormalities at 2.5 mg/kg DDT while much smaller tadpoles ($x = 363 \pm 11$ mg) exhibited hyperactivity with only 2.2 mg/kg DDT (Table 2).

DDT residues can be retained for a long period of time after applications are stopped (Dimond et al. 1968). Sampling of P. cinereus in sprayed forests in Maine showed that it took 8 to 9 years before DDT residues in adult salamander returned to pre-spray levels (Ibid 1968) (Table 4).

Mechanism for Toxicity

DDT enters the organism at different life stages via several routes; at the early spawn stage (Cooke 1972a) (Table 2), via prey species and water intake both orally and via absorption through the skin (Kaiser and Dunham 1972-73) (Table 2). Having entered the body, the mechanism for toxicity of DDT, although mainly due to a physiological response, can in part be due to behavioural aberrations. The morphological abnormalities observed in R. temporaria tadpoles after exposure to DDT were due to a disruption of glandular development in the external skin of the snout (Cooke 1970, Osborn et al. 1981). The disruption of glandular development coupled with the hyperactive behaviour, causes the lower mandible to strike the inner surface of the upper mandible. This damage, in conjunction with the disrupted external skin gland, can cause the loss of the upper mandible in some individuals. These deformities were sometimes permanent, resulting in a blunt snout and brain deformities (Ibid 1981) (Table 2).

DDT has been shown to significantly decrease the shortening time, twitch duration, contraction and relaxation time of the skeletal muscles of Rana hexadactyla (Rajendra et al. 1980). DDT may also increase nerve acetylcholine by enhancing liberation from bound reserves which play an important role in the dynamics of contractile tissues (Rajendra et al. 1980, Craciun et al. 1981) (Table 2).

Akkermans et al. (1975a) studied another physiological response to DDT. They reported DDT caused repetitive activity in the cutaneous touch receptors of X. laevis (Table 2) which may affect the toad's reactions and behaviour to external stimuli.

ENDOSULFAN

Short-Term Effects

In static bioassays with endosulfan, R. tigrina tadpoles had LC50 an order of magnitude lower than juvenile catfish (Clarias batrachus) and damselfly nymphs (Enallagma spp.) (Gopal et al. 1981) (Table 1). A reduced swimming capacity, lack of physical stamina and a thinning and transparency of the gills were the observed symptoms of endosulfan poisoning (Ibid 1981).

Long-Term Effects

In an aerial application of 0.014 kg/ha a.i. endosulfan on a savannah woodland in Zimbabwe-Rhodesia, no mortality was recorded for adult Chiromantis xeampelina adults maintained in cages for 19 days after treatment (Cockbill 1979) (Table 3), although mortality was recorded to some shallow water fish (Tilapia spp.). There are no data for residue accumulations of endosulfan in amphibians.

ENDRIN

Short-Term Effects

Endrin toxicity to amphibians varies depending upon the species and the lifestage. R. sphenocephala eggs in continuous flow bioassays were sensitivity of R. temporaria tadpoles to endrin in static bioassays gradually increased during development with two-day old tadpoles having a 72 hour LC50 of 0.425 mg/L while for 107-day old tadpoles (with fore and hind limbs developed) the 72 hour LC50 was 0.015 mg/L (Wohlgemuth 1977) (Table 1). The increase in sensitivity to endrin during development was thought to be due to changes in respiration and metabolism (Ibid 1977).

Wohlgemuth (1977) conducted a comparison of the toxicity of endrin to seven species of amphibians. The results are presented on Table 1. All species were in the tadpole stage except Ambystoma opacum. R. sylvatica and A. opacum were most resistant to endrin but the high LC50 for A. opacum was thought to be due to the older age of these individuals. R. catesbeiana was the most sensitive to endrin toxicity. Behavioural abnormalities during the exposure were all associated with mortality (Ibid 1977).

In static bioassays with 4-5 week old B. woodhousii tadpoles, endrin had a 96 hour LC50 of 0.12 mg/L and was the second most toxic pesticide tested (Sanders 1970) (Table 1). One week old P. triseriata tadpoles had a similar 96 hour LC50 of 0.18 mg/L endrin (Table 1). This 96 hour LC50 cannot be compared directly with those for the species used by Hall and Swineford (1980, 1981) because the former used continuous flow dosing, while the latter used static dosing (Table 1).

Field applications of endrin have also been shown to be highly toxic to amphibians. Applications of 0.11 and 0.56 kg/ha endrin caused 100% mortality to R. catesbeiana, B. boreas and S. hammondi tadpoles within 5 days of treatment (Mulla 1962, 1963) (Table 3). Endrin was one of the most toxic of the organochlorine insecticides tested (Ibid 1963).

In an experiment designed to study the effects of endrin on an artificially evacuated pool with no inlet, 0.5L of 20% endrin was poured in at one end. Immediately a white turbid cloud was formed which moved off due to the wind. Concentrations of endrin were found to be 0.047 mg/L 10 metres from the point of entry (Wohlgemuth and Trnkova 1979) (Table 3). Aquatic invertebrates died within five minutes of contact with the endrin. The indigenous population of R. esculenta adults began to emit high shrieks and experience severe cramping and paralysis. This was followed by death. Caged R. temporaria tadpoles died within thirteen minutes of contact with the endrin. The concentration of endrin over the volume of the pool was calculated to be 2.5 mg/L (Ibid 1979) (Table 3).

Long-Term Effects

Kaplan and Overpeck (1964) placed R. pipiens adults in varying concentrations of endrin and recorded their survival after 30 days (Table 2). Frogs exposed to 0.03 ppm endrin in solution suffered 30% mortality after 30 days. Endrin caused neuromuscular changes to occur including excessive thrashing and tremors (Ibid 1964).

Webb et al. (1979) examined the effect of endrin of the short-circuit current and active sodium transport across the skin of R. pipiens adults (Table 2). They found endrin did not produce a significant effect on the short-circuit current or resistance of the frog skin at a concentration of 2×10^{-4} M.

Residues

R. sphenocephala accumulated endrin residues up to 2.8 mg/kg which was 94 times water levels (Hall and Swineford 1980). Frogs which survived endrin exposures however never accumulated more than 0.06 mg/kg which was a concentration factor of only six (Ibid 1980).

Accumulation of endrin residues in amphibian foods can cause mortality when ingested. An endrin resistant population of mosquito fish (Gambusia affinis) exposed to 2 mg/L endrin for 7 days and then force fed to R. catesbeiana adults resulted in 100% frog mortality within 24 hours (Rosato and Ferguson 1968) (Table 2). A pooled sample (N=4) of field collected mosquito fish contained 9.65 mg/kg DDT and a pooled sample (N=4) of endrin treated mosquito fish exhibited residues of 890 mg/kg of endrin.

Niethammer et al. (1984) analyzed three species of frog from alluvial cropland in Louisiana that had been subjected to heavy pesticide use (Table 2). The low levels reported for amphibians were considered to be due to an efficient ability to excrete these contaminants.

LINDANE

Short-Term Effects

In static bioassays lindane was the least toxic of 14 insecticides to 1 week old P. triseriata and 4 week old B. woodhousii tadpoles (Sanders 1970) (Table 1). Mortality was preceded by irritability and loss of equilibrium. These effects were noted at concentrations lower than the 96 hour LC50 value and were irreversible (Ibid 1970).

Long-Term Effects

In chronic exposures to lindane, the egg stage of X. laevis had lower hatching success at all the lindane concentrations tested (Marchal-Segault and Ramade 1981) (Table 2). When the tadpole stage was exposed to the same lindane concentrations, mortality increased with time and after 6 weeks' exposure was 100% (Table 2). Tadpoles exposed to lindane at lower weights, showed morphological deformities in comparison with the controls. The morphological deformities included lateral deviation of the body axis and a dorsal lump on the cranial area. A darkening of pigmentation also occurred in treated tadpoles and persisted in the surviving adults (Ibid 1981).

Mulla (1962, 1963) found lindane caused the lowest mortality of R. catesbeiana tadpoles in comparison with ten other organochlorine insecticides. Lindane caused only 10% mortality after 7 days at an application rate of 0.56 kg/ha (Table 3).

Residues

Whitacre and Ware (1967) examined the chronic effects of vaporized lindane on R. pipiens adults. Lindane residues in the brain, liver and fat were measured during 38 day exposures. Residue accumulation increased with increased exposure to lindane (Table 4). Residues contained in fat actually declined over the exposure period and were not detectable after day 38 (Table 4). No frog mortality occurred during the exposure but residues were much greater than in carp, guppies, mice or rats. The high residue levels in the frogs were thought to have been caused by the frogs not eating during exposure. This could have lowered the rate of metabolism and excretion resulting in accumulation of high levels (Ibid 1967).

Lindane has also been shown to readily pass through the skin of R. pipiens (Kaiser and Dunham 1972) (Table 2). The high water solubility of lindane and simple diffusion through the skin were speculated to be the cause of lindane being readily accumulated (Ibid 1972).

Mechanism for Toxicity

Subacute chronic exposures to lindane during growth and development have effects analogous to those reported for eggs and tadpoles intoxicated with fungicides (Bancroft and Prahlad 1973), herbicides (Cooke 1977), and fenthion (an organophosphorous insecticide) and so may be caused by similar mechanisms (Marchal-Segault and Ramade 1981). Marchal-Segault and Ramade (1981) suggested that the delays in growth and development they observed due to chronic exposures of lindane, indicate some endocrine dysfunction of the hypothalamo-hypophyseal axis. This dysfunction did not appear to be complete, since some surviving animals attained the final stage of metamorphosis. An increase in the pigmentation, which was also noted during the tadpole stage, indicated a dysfunction of the intermediate lobe of the hypophysis that controls pigmentation (Ibid 1981).

In adult frogs lindane has been shown to cause damage to skeletal muscle (Publicover et al. 1979). The damage was caused by a rise in calcium concentration resulting in the dissolution of muscle myofilaments. In vivo, the increased calcium also caused a marked increase in the miniature endplate potential frequency at the neuromuscular junction with the threshold for an observable response at 0.0003 M (Ibid 1979) (Table 2). Publicover and Duncan (1979) speculated that if lindane has similar effects on other synapses of the central nervous system then hyperexcitability, tremors and convulsions could be produced. These effects, however, have not been reported for amphibians exposed to lindane.

The increase in calcium concentration appears to be caused by lindane decreasing the permeability of the outer skin membrane to all ions except sodium. Sodium permeability is actually increased, promoting calcium release from storage sites such as the mitochondria (Publicover et al. 1979, Publicover and Duncan 1979, Webb et al. 1979).

METHOXYCHLOR

Short-Term Effects

Methoxychlor was moderately toxic to P. triseriatata tadpoles in static bioassays with a 48 hour LC50 of 0.42 mg/L while B. woodhousii tadpoles had a 48 hour LC50 of 0.11 mg/L (Sanders 1970) (Table 2).

Residues

Methoxychlor was much more readily accumulated in B. americanus adults after exposure in water than through ingestion of mealworms containing methoxychlor residues (Hall and Swineford 1979). Feeding tests in which adult toads were fed mealworms containing 0.024 mg/kg of methoxychlor for 6 days revealed an accumulation of 0.008 mg/kg in the whole body of the toads. Conversely, toads that were exposed to water containing 0.069±0.036 mg/L methoxychlor for 6 days accumulated 0.244 mg/kg in the body (Table 4). Residue levels were not correlated with duration of exposure suggesting equilibrium is reached rapidly. There were no changes in organ weights, feeding, behaviour or survival due to the exposure to methoxychlor (Hall and Swineford 1979).

MIREX

Residues

Mirex is a chlorinated hydrocarbon insecticide used specifically for control of the fire ants (Salenopsis saevissima) in the southeastern United States. In separate studies in Florida (Wheeler et al. 1977), Georgia (Wojcik et al. 1975), Mississippi (Naqvi and de la Cruz 1973) and Louisiana (Niethammer et al. 1984) reported the levels

of mirex in amphibians. Wojcik et al. (1975) reported that the highest levels occurred one to three months after treatment and Wheeler et al. (1977) found that amphibians still showed low levels a year after a single application of mirex. Naqvi and de la Cruz (1973) found residues in some organisms in an area not treated with mirex, suggesting widespread movement in the environment and evidence of some biological magnification. Niethammer et al. (1984) considered that the low residue levels in frogs might be due to their high turn-over of fatty acids (Brown 1964) which could increase their ability to mobilize and excrete chlorinated hydrocarbon compounds.

After an application of mirex to a fish pond in Mississippi the residues found in A. gryllus reached a maximum after 6 months and declined to 0.05 ppm within 16 months of treatment. The initial increase in residues was partly attributed to an accumulation through the food chain (Collins et al. 1973) (Table 4).

TOXAPHENE

Short-Term Effects

Differences in toxicity of toxaphene to amphibians have been shown to vary with life stage and species. In continuous flow bioassays, R. sphenocephala eggs were resistant to toxaphene in comparison with the tadpole stage, which in turn was more sensitive than the two sub-adults tested (Hall and Swineford 1980) (Table 1). Behavioural aberrations (mainly hyperirritability and prolonged stupor) and growth retardation were observed for toxaphene treated tadpoles with effects on growth in 96 hour exposures, occurring at 0.013-0.018 mg/L (Ibid 1980).

Species differences in sensitivity to toxaphene were observed in continuous flow bioassays for seven species, although the species were at varying lifestages (Hall and Swineford 1981). The species and 96 hour LC50s are presented on Table 1. A. opacum and R. sylvatica were most tolerant while A. crepitans and A. maculatum were most sensitive to toxaphene. The high tolerance of A. opacum was thought to be partly due to age as they were field collected as adults as all other species were in the larval stages (Ibid 1981).

In static bioassays Sanders (1970) measured the 96 hour LC50 of toxaphene for B. woodhousii and P. triseriata (Table 1). The variation in sensitivity may have been a result of species differences and/or differences in age (Sanders 1970).

The seven species exhibited a range in severity of behavioural abnormalities which were caused by exposure to different concentrations of toxaphene (studied by Hall and Swineford 1981). R. catesbeiana exhibited only subtle behavioural abnormalities with mild loss of equilibrium and disorientation where A. opacum displayed debilitating contortions and immobilization. Some species exhibited behavioural abnormalities at concentrations much lower than their LC50 values. The independence of the behavioural abnormalities and mortality led Hall and Swineford (1981) to conclude that the mechanisms for these two effects were acting separately.

In field trials, using R. catesbeiana, Mulla (1962, 1963) found toxaphene to be among the most toxic substances that were tested. At an application rate of 0.5 kg/ha the tadpoles suffered 100% mortality within 24 hours (Table 3). No mortality was observed when toxaphene was applied at 0.1 kg/ha (Mulla 1963) (Table 3).

Long-Term Effects

Kaplan and Overpeck (1964) examined the survival of R. pipiens at different concentrations of toxaphene (Table 2). After 30 days the frogs exposed to the highest level (0.60 ppm) suffered 25% mortality. Neuromuscular changes were observed in treated frogs in conjunction with thrashing and tremors (Kaplan and Overpeck 1964).

Residues

Organochlorine residues were measured in the fauna of oxbow lakes in Louisiana that have acted as sumps in accumulating residues from nearby agricultural land (Niethammer et al. 1984) (Table 4). Toxaphene was found to be one of the principal residues detected. The levels in R. catesbeiana, R. sphenocephala and the Bronze frog were low (Table 4) when compared to their trophic level. These workers related this to the high turnover of fatty acids in these species (Brown 1964).

Hall and Swineford (1980) reported that frogs (R. sphenocephala) were able to concentrate toxaphene residues by a factor of one hundred. These results are based on animals under constant exposure to the toxicant and for comparison, it was also noted that some fish concentrate residues by factors of one hundred thousand under these conditions (Hall and Swineford 1980).

OTHERS

This section deals with several organochlorine insecticides for which there is little information available concerning their toxicity to amphibians.

Kepone EC2 is utilized primarily for agricultural pest control. When applied to ponds at 0.11 and 0.56 kg/ha it produced no mortality among R. catesbeiana tadpoles (Mulla 1963) (Table 3).

Thiodan I is also an agricultural pest control insecticide. An application of 0.56 kg/ha to a pond resulted in 100 percent mortality of tadpoles within 24 hours (Mulla 1962, 1963) (Table 3). When Thiodan II was applied at the identical rate the result was the same to tadpoles of R. catesbeiana (Mulla 1963).

Trichlorphon injected into R. temporaria adults at a dose of 50-300 mg/kg caused a reduced red blood cell count and shifts in the composition of white blood cells (Szubartowska 1979) (Table 2).

Benzene hexachloride, an insecticide closely related to lindane, was reported as being moderately toxic to Bufo woodhousii tadpoles (Sanders 1970) (Table 1). Kaplan and Overpeck (1964) observed the survival of R. pipiens adults in varying concentrations of BHC and reported that they suffered 40% mortality after exposure to the highest concentration (Table 2).

Residues of BHC were not detected in any amphibian sample from contaminated oxbow lakes in Louisiana (Niethammer et al. 1984) (Table 4).

Heptachlor, another insecticide, was also found to be moderately toxic to B. woodhousii tadpoles (Sanders 1970) (Table 1). At an application rate of .56 kg/ha heptachlor caused 80% mortality two days after treatment (Mulla 1962, 1963) (Table 3). No further mortality was observed 5 days after treatment. Albright et al. (1980) analyzed Tarichia torosa in British Columbia and found heptachlor levels in the newts 4 days after treatment with chlordane. Subsequent samples taken from the same location had non-detectable amounts of heptachlor epoxide (Albright et al. 1980) (Table 4).

b) CARBAMATES

Short-Term Effects

In static bioassays using Rana clamitans tadpoles, aminocarb had a 24 hour LC50 of 247 mg/L for small tadpoles and 234 mg/L for larger tadpoles. The 96 hour LC50 for small tadpoles was 118 mg/L (Lyons et al. 1976) (Table 2). Treated tadpoles had extensive hemorrhagic regions and exhibited jaw-twitching, ecdysis, difficulty in swimming, buoyancy problems, colour change, mouth deformities, edema and bent tails. The mouth deformities were thought to have occurred through direct contact of the snout with aminocarb that had adhered to the sides of the treatment container (Ibid 1976).

In other toxicity tests involving carbamates, Marking and Chandler (1981) reported a 96 hour LC50 of 8.7 mg/L for methiocarb, employing R. sphenocephala larvae (Table 1). Marchal-Segault (1976), exposing B. bufo tadpoles to various concentrations of sevin, found a 72 hour LC50 of between 16.8 and 20.6 mg/L (Table 1). Tucker and Crabtree (1969) estimated an LD50 of 283-800 mg/kg in a single oral dose of zectran to adult R. catesbeiana (Table 1). They considered that zectran would not be toxic to adult amphibians at the current field application rates.

Marian et al. (1983) obtained a 96 hour LC50 value of 6.2 mg/L of carbaryl for tadpoles of R. tigrina. Rzehak et al. (1977) conducted static bioassays using different concentrations of carbaryl and both R. temporaria and X. laevis tadpoles. Exposure to a concentration of 0.1% caused an increase in mobility of X. laevis tadpoles. When placed in tap water after exposures of 30, 60 and 120 minutes the tadpoles gradually returned to normal. Histological examination of the group exposed for two hours revealed contraction of miomers; partial separation of muscle fibers and oedema of the intestinal epithelium (Rzehak et al. 1977). Tadpoles of R. temporaria when exposed to a concentration of 0.1% and then returned to tap water after 15 minutes, reacted in the same manner as X. laevis. None of these exposures seem to affect the development of the organism. Marian et al. (1983) found R. tigrina tadpoles to be moderately resistant to carbaryl in 96 hour toxicity tests (Table 1). However, prolonged exposures using 0.01% and 0.001% carbaryl produced different results (Table 2).

After an exposure of 24 hours to 0.01% carbaryl tadpoles of R. temporaria were smaller than the corresponding controls and exhibited bent tails, causing the tadpoles to swim in tight circles (Rzehak et al. 1977) (Table 2). When transferred into tap water, 90% of the tadpoles survived, though some with severe abnormalities that restricted their movements died later. Under the same conditions, tadpoles of X. laevis exhibited a reduced heartbeat, bent tails, partial separation and contraction of muscle fibres and appeared moribund. Eighty percent of these tadpoles died within a week after exposure. Exposures of 10 days for R. temporaria and 8 days for X. laevis to 0.001% carbaryl led to permanent tail malformation in both species. R. temporaria also exhibited growth inhibition, with 90% of the tadpoles dying between day 7 and day 10 of the treatment and the remainder dying within 2 days following transferral to tap water. After 8 days of exposure to 0.001% carbaryl, further growth and development of X. laevis tadpoles was inhibited. Upon transferral to tap water many individuals died and mortality for the first week after treatment was 20% (Ibid 1977) (Table 2).

In general both species exhibited behavioural changes immediately after exposure to carbaryl. Their activity increased and then, after a period of time, their movement slowed until they lay motionless on the bottom of the test vessel. The length of time for this behaviour varied with the concentration of carbaryl (Rzehak et al. 1977).

Application of carbofuran to rice fields in Texas had severe effects on A. crepitans blanchardi adults (Flickinger et al. 1980). Frogs were found paralyzed or exhibiting abnormal behaviour as early as 15 minutes after application. Four out of 19 frogs observed one half hour after treatment were paralyzed. Only one frog was found dead (Ibid 1980) (Table 3).

R. temporaria tadpoles maintained in cages and suspended in seasonal ponds in ditches near farm fields sprayed with oxamyl exhibited a significantly higher incidence of vertical curvature deformities and tail tip defects than tadpoles maintained in ditches near fields not sprayed with oxamyl (Cooke 1981). In the lab after exposure for 1 hour to 100 mg/L oxamyl, 90% of the tadpoles exhibited vertical curvature deformities similar to those observed in the field. The deformities appeared not to be spinal, but rather of muscular origin (Ibid 1981) (Tables 2 and 3).

Long Term Effects

Field applications of zectran in New Brunswick over several years produced no adverse effects on the population of R. clamitans and posed no serious hazard to either larval or adult frogs (Pearce and Rick 1969, Pearce and Price 1977, Rick and Price 1974) (Tables 2 and 3). Matacil (aminocarb) produced no adverse effects on individual amphibians or their populations. Bracher and Bider (1982) found no significant change in populations of R. sylvatica and R. pipiens following aerial application of matacil in Quebec (Table 3). Similarly, following spraying in New Brunswick Rick and Price (1974) observed no mortality of frogs, toads or tadpoles and Pearce and Price (1977) concluded adult and larval amphibians were unaffected by matacil (Table 3).

In experiments to determine the effects of carbaryl on growth and metamorphosis of R. tigrina, Marian et al. (1983) raised tadpoles from hatching to metamorphosis in sublethal concentrations. It was found that carbaryl does not affect the larval or metamorphic period. During metamorphosis mortality was significant at the 2 mg/L level (Table 2). It was concluded that any level below 5 ppm may be considered as sublethal (Marian et al. 1983).

Residues

Bullfrog tadpoles were analyzed for residues of carbofuran following an application of 0.025 ppm in conjunction with an application of atrazine at 0.3 ppm. Residues of carbofuran were not detectable (less than 0.4 ppb) 77 days post-treatment (Klaassen and Kadoum 1979).

Mechanism for Toxicity

Carbamates are systemic insecticides that act by inhibiting cholinesterase activity (Corbett 1974, Przĕzdziecki 1976). The cholinesterase inhibition caused by carbaryl led to contraction and partial separation of muscle fibres and the eventual immobilization of tadpoles (Rzehak et al. 1977). Carbaryl also had an effect on frog skin where it increased the sodium (Na^+) uptake suggesting that the outer border of transporting cells is the area of action of externally applied insecticides (Webb et al. 1979) (Table 2). Takeno et al. (1977) found the response of R. pipiens nerve muscle to indirect stimulation was suppressed more effectively than the response to direct stimulation in the presence of 2×10^{-4} M carbofuran. Various concentrations of carbofuran also suppressed muscle contractions in frog nerve-muscle (Table 2).

Prolonged exposures to carbaryl leading to long-term contraction of trunk and tail muscles may lead to secondary changes in other organs such as the tail bones (Rzehak et al. 1977). Carbaryl derivatives can be detoxified, which explains why tadpoles recovered from short exposures in the experiment conducted by Rzehak et al. (1977) (Table 2).

c) ORGANOPHOSPHATES

FENITROTHION

Short-Term Effects

Lyons et al. (1976) using R. clamitans tadpoles, established a 96 hour LC50 of 4.9 ppm (Table 1). Fenitrothion was more toxic to amphibians than two other compounds tested simultaneously (Matacil and Orthene). Symptoms of fenitrothion poisoning were characterized by swimming difficulties, bent tails, jaw twitching, ecdysis and gulping, apparently in an effort to move more water over the gills (Lyons et al. 1976). The authors speculated that altered physical behaviour stimulated by reduced cholinesteratic activity induced by organophosphates would place the tadpoles at a selective disadvantage.

Long-Term Effects

Folithion (fenitrothion) drastically altered the length of the larval period of R. tigrina and reduced the number that metamorphosed (Mohanty-Hejmadi and Dutta 1981) (See Folithion, Table 2). Concentra-

tions of 4 ppm reduced the number of larvae that metamorphosed and extended the larval period by approximately 40 days compared to the control group (Table 2).

In field studies on the effect of fenitrothion on amphibian populations two distinct results have been reported. In two separate studies conducted in New Brunswick no mortality or unusual behaviour was observed in frogs, toads and tadpoles (Pearce and Price 1977, Rick and Price 1974) (Table 3). However, Thirumurthi et al. (1973) observed 100% mortality of frogs in Indian rice fields (Table 3). The application rate on the rice fields was 0.5 kg/ha and in New Brunswick it was 0.14 kg/ha applied twice, in one study, and 0.057 kg/ha, applied twice, in the second study.

FENTHION

Short-Term Effects

Using 26 to 37 day old B. bufo tadpoles, Marchal-Segault (1976) reported a 48 hour LC50 of 2.0-2.2 ppm for fenthion (Table 1). It was also reported that development of the larvae was retarded by exposure to fenthion (Ibid 1976). Hall and Kolbe (1980) calculated a 96 hour LD50 of 4.9 ppm for R. catesbeiana tadpoles (Table 1).

Long-Term Effects

A single aerial application of fenthion at 0.5 kg/ha on Indian rice fields produced 100% mortality of frogs in the field (Thirumurthi et al. 1973) (Table 3).

Johnson and Prine (1976) reported that fenthion, when used at one-half of the usual field concentrations, produced a pronounced reduction in the temperature tolerance range of B. boreas (Table 2). Sublethal effects of this type may render the organism more susceptible to environmental temperature shifts causing death within a much narrower range than non contaminated healthy toads. Fenthion produced the largest reduction of the six insecticides tested (Johnson and Prine 1976).

Residues

In two experiments to test whether Rana catesbeiana tadpoles could accumulate fenthion to levels that would be lethal to Mallard (Anas platyrhynchos) ducklings, Hall and Kolbe (1980) found that 8 of 10 ducklings died after being fed exposed tadpoles (Table 2). In the first experiment, tadpoles placed in water containing 2.2 ppm of fenthion and then fed to ducklings killed 4 of 6 ducklings. Mortality was 100% within 5 hours in ducklings fed tadpoles dosed at 5 ppm fenthion (Table 2). Significant reductions in cholinesteratic activity were reported for the ducklings (Hall and Kolbe 1980).

GUTHION

Short-Term Effects

In static bioassays using B. woodhousii tadpoles Sanders (1970) reported a 96 hour LC50 of 0.13 ppm for guthion (Table 1), the organisms went through a period of irritability followed by a loss of equilibrium and finally death.

Long-Term Effects

Mulla (1962) reported that guthion at an application rate of 1.8 kg/ha, resulted in 100% mortality of R. catesbeiana. Mulla et al. (1963) also reported that at rates of .11 and .45 kg/ha a.i. no mortality was observed within cages containing B. boreas and S. hammondi (Table 3).

MALATHION

Short-Term Effects

In static toxicity tests Sanders (1970) found 96 hour LC50's of 0.42 and 0.20 ppm to B. woodhousii and P. triseriata respectively. When exposed to a concentration of approximately 5 ppm malathion for 96 hours, forty percent of R. catesbeiana tadpoles died (Hall and Kolbe 1980) (Table 1).

Malathion also caused abnormal development in Microhyla ornata at the yolk plug stage when exposed to concentrations of between 1 and 20 ppm (Pawar et al. 1983) (Table 2). Observed abnormalities included head, trunk and tail defects, behavioural aberrations, loss of balance, poor pigmentation and retarded growth. Mortality increased with increasing concentrations of malathion (Ibid 1983).

Long-Term Effects

Mohanty-Hejmadi and Dutta (1981) also reported that malathion affects growth of amphibians. In tests using R. tigrina it was found that malathion caused a prolongation of life history and smaller animals after metamorphosis (Table 2). The concentrations used in this experiment, though introduced via chronic exposure, were much lower than levels recommended for field applications of malathion.

The effects of malathion at different concentrations ranged from a decrease in white cell counts, and depression of red blood cells to physical lethargy causing reduced avoidance response (Kaplan and Glaczenski 1965) (Table 2).

Aerial applications using Ambithion (malathion and fenitrothion) effectively eliminated frog populations in sprayed areas (Thirumurthi et al. 1973) (See Ambithion, Table 3).

Residues

Hall and Kolbe (1980) studied the accumulation of several organophosphate insecticides by amphibians. R. catesbeiana tadpoles were dosed with malathion at 5 mg/L and then force fed to Mallard ducklings. No mortality among the ducklings was observed and it was concluded that symptoms of malathion intoxication would only be observed if the insecticide was massively accumulated (Ibid 1980) (Table 2).

PARATHION

Short-Term Effects

In static toxicity bioassays using P. triseriata the 96 hour LC50 for parathion was 1.0 ppm (Sanders 1970) (Table 1). Hall and Kolbe (1980) calculated the LD50 for R. catesbeiana tadpoles to be 1.32 ppm and the LD50 reported by Edery and Schatzberg-Porath (1960) following injection into the dorsal lymphatic sac, was 967 mg/kg (Table 1). In the latter study B. vulgaris was found to be more resistant to parathion than either the frog or mouse used as comparative species.

Guzman and Guardia (1978) conducted experiments on the short-term and long-term effects of exposure to parathion. B. arenarum were exposed to parathion in two groups. Group 1 received constant exposure and Group 2 was exposed for 2 hours. Both groups exhibited an initial loss of cholinesteratic activity and subsequently recovered when removed from the contaminated medium (Ibid 1978) (Table 2).

Long-Term Effects

Kaplan and Glaczenski (1965) noted the same results in hematological tests with parathion as those exhibited by frogs exposed to malathion, though to a slightly more severe degree (Table 2).

Field studies examining the effect of parathion on populations of amphibians revealed that aerial applications of 0.11 to 1.12 kg/ha did not adversely affect amphibian populations in several different habitats (Mulla 1962, Mulla et al. 1963, 1966) (Table 3).

Residues

Amphibians are generally resistant to cholinesterase inhibitors (Potter and O'Brien 1964). This coupled with the fact that organophosphates have a low persistence in the environment may indicate

that adverse effects from utilizing these insecticides may be a result of the ability of certain organisms to accumulate the contaminants and move large concentrations up through the food chain (Hall and Kolbe 1980). Several studies have been undertaken to test this idea using amphibians and organisms located at higher trophic levels.

Hall and Kolbe (1980) fed Mallard ducklings tadpoles that had been exposed to 1 ppm parathion. Accumulation in the tadpoles was estimated to be 64 times the level at which they were exposed. The ducklings fed these tadpoles all died. Similarly all ducklings died within one half hour after ingesting tadpoles dosed to 5 ppm of parathion (Ibid 1980) (Table 3).

Fleming et al. (1982), fed American kestrels (Falco sparverius) frogs (A. crepitans) that had been exposed to concentrations of 0.1-10 ppm parathion for 96 hours. One bird of four died 3 hours after eating 5 frogs exposed to 10 ppm. No mortality of kestrels was recorded from any other group (Table 2). The authors noted that 10 ppm was an unrealistic exposure rate for field applications (Fleming et al. 1982). However, frogs can accumulate parathion from water as Fleming et al. (1982) found that after 96 hours parathion was not detectable in the water but was present in frogs (Ibid 1982) (Table 2). This may be due to the constant availability of parathion to the frogs which is a situation that is unlikely to be duplicated in the field.

ORTHENE (Acephate)

Short-term Effects

In tests using R. clamitans tadpoles Lyons et al. (1976), from the maximum likelihood regression analysis, established a 24 hour LC50 of 6433 ppm (Table 1). Similarly, Geen et al. (1984) reported an equally low toxicity for the salamander Ambystoma gracile. Sixty-nine day old larvae had a 96 hour LC50 of 8816 mg/L (Table 1). Amphibians would not be affected by control programs using Orthene as the maximum surface residue from current field application rates would be less than 1 ppm (Ibid 1984).

Long-Term Effects

Geen et al. (1984) found that 798 mg/L of Orthene did not produce significant differences in mortality to Ambystoma gracile (Table 2). There was some decrease in the growth rate and the appearance of abnormalities in salamanders exposed to 382 and 798 mg/L (Table 2), but these are well above the levels likely to be encountered in the field (Geen et al. 1984).

Buckner and McLeod (1975) monitored R. sylvatica, Hyla crucifer and Ambystoma laterale for 7 days after an aerial application of Orthene in two mixed wood stands in Ontario. It was concluded that the amphibians "were not affected by the insecticide treatment" (Ibid 1975) (Table 3).

Residues

When ducklings were force fed tadpoles exposed to 5 ppm Orthene no adverse effects or mortality were observed (Hall and Kolbe 1980) (Table 1).

OTHERS

Short-Term Effects

Using P. triseriata in static toxicity bioassays Sanders (1970) determined the 96 hour LC50's of a number of organophosphate insecticides. Carbophenthion was found to be the most toxic of the sixteen substances tested with a 96 hour LC50 of 0.028 mg/L (Table 1). Naled, another insecticide, was moderately toxic with a 96 hour LC50 of 1.7 mg/L (Sanders 1970) (Table 1).

In 96 hour flow through experiments exposing R. catesbeiana tadpoles to 5 mg/L of dicrotophos no mortality was observed though there was evidence of adverse cholinesterase effects (Hall and Kolbe 1980).

Injection into the dorsal lymphatic sac of amphibians yielded the following LD50's for B. viridis; 188 mg/kg paraoxon, 540 mg/kg TEPP, 1,410 mg/kg Dimefox and 1,450 mg/kg DFP. LD50's for R. ridibunda were only conducted with paraoxon and TEPP and were 91 mg/kg and 34 mg/kg respectively (Edery and Schatzberg-Porath 1960) (Table 1). Symptoms of poisoning were reported as a generalized paralysis followed by a loss of ability to right themselves and finally coma. Recovery was observed after removal from the contaminated medium. Edery and Schatzberg-Porath (1960) stated that amphibians show an unusually strong resistance to organophosphorus intoxication.

Long-Term Effects

Chronic exposures to Abate, chlorpyrifos-ethyl, chlorpyrifos-methyl (0.03 or 0.06 ppm) and methyl-parathion (0.025 or 0.05 ppm) caused a reduction in the thermal tolerance of B. boreas (Johnson and Prine 1976) (Table 2). The levels employed only represented fifty percent of field concentrations (Ibid 1976). Johnson (1980) also tested temperature tolerance in the presence of temephos and chlorpyrifos and found it was significantly reduced even at concentrations less than recommended field application rates (Table 2).

Development of R. tigrina affected by DDT and methyl-parathion was studied by Mohanty-Hejmadi and Dutta (1981) (Table 2). The toxic effects caused by exposures of 1-7 mg/L varied depending upon the tadpole's developmental stage at the time of treatment. The limb-bud and well developed hind limb stages were the most sensitive to these insecticides. Complete developmental arrest occurred at higher concentrations and all treated frogs were smaller than controls at metamorphosis. In general, all the insecticides caused an extension of the life histories, even at concentrations lower than recommended field application rates (Ibid 1981). These effects were also observed by Dutta and Mohanty-Hejmadi (1978) in studies using different concentrations of Rogor (dimethoate) and R. tigrina (Table 2).

In field studies conducted by Mulla et al. (1963) insecticides were applied to ponds and the resulting mortality of amphibians recorded. Applications of 0.11 and 0.45 kg/ha of methyl-parathion resulted in no mortality to B. boreas and S. hammondii populations (Table 3). No mortality was observed for R. catesbeiana tadpoles following an application 0.56 kg/ha of naled, however applications of 1.8 and 0.45 kg/ha carbophenthion resulted in 100 percent mortality within twenty-four hours (Mulla et al. 1963) (Table 3).

Several other insecticides were tested (See Table 3 for data on Bayer 38920, G-27365, Bayer 34042, Ethyl Guthion, G.C.-3582, Bayer 44831, Bayer 29952, Bayer 22408, G-28029, Bayer 37289, Methyl-parathion, G-30494, G-30493 and Ronnel) and only G.C.-3582 was toxic to R. catesbeiana with 100 percent mortality when applied at 1.8 kg/ha (Mulla et al. 1963) (Table 3).

Field studies involving leptophos and phosphamidon concluded that these insecticides were highly toxic to frogs, causing 100 and 75 percent mortality respectively (Thirumurthi et al. 1973) (Table 3). In contrast Pearce and Price (1977) concluded that phosphamidon, due to relatively low toxicity observed in toxicity tests, would not present a major direct hazard to amphibians. Pearce and Teeple (1969) also examined the effects of sumithion spraying on amphibians in New Brunswick and concluded that at 0.14 kg/ha it had no effect on frogs and toads (Table 3).

Residues

Hall and Kolbe (1980) studied the ability of R. catesbeiana to accumulate dicrotophos (Bidrin) to levels lethal to Mallard ducklings. Tadpoles dosed with 5 ppm and fed to ducklings caused some reduction in cholinesterase activity but resulted in no mortality of ducklings suggesting the tadpoles could not accumulate dicrotophos to levels lethal to duckling consuming one meal (Table 2).

Mechanism of Toxicity

Organophosphate insecticides are toxic to most organisms due to the inhibition of acetylcholinesterase enzymes that are essential for nerve impulse conduction and transmission (Corbett 1974). This inhibition can lead to erratic altered behaviour (Lyons et al. 1976, Pawar et al. 1983, Sanders 1970). Erratic behaviour may place the organism at a selective disadvantage and lead to increased predation (Cooke 1971, Hall and Kolbe 1980, Lyons et al. 1976, Mohanty-Hejmadi and Dutta 1981). Direct effects of this reduction of cholinesterase activity have been studied by Dekin et al. (1978) who observed inhibition of muscle response caused by exposure to organophosphates (Table 2). Guy et al. (1977) showed a reduced amplitude of the response to stimulation of the sartorius muscle in the presence of OMPA (Table 2). These studies suggest that organophosphates may exert their toxicity by affecting the cholinergic receptor-channel complex.

Several studies have shown that amphibians can recover from this reduction of cholinesterase activity if returned to contaminant free media (Dekin et al. 1978, Guy et al. 1977, Guzman and Guardia 1978).

Potter and O'Brien (1964) attempting to explain the relative low toxicity of organophosphates to frogs in contrast with some insects and mammals concluded that the insensitivity of frogs to paraoxon was due to its insensitive cholinesterase which may account for the low relative toxicities to many organophosphate insecticides (Ibid 1963).

d) PYRETHROIDS

Short-Term Effects

Pyrethroids are a group of insecticides developed from a natural substance (Pyrethrum) and are of particular interest due to a short half life in the environment and their high toxicity to insects.

In static bioassays, permethrin had a 96 hour LC50 of 7.03 mg/L for two week old R. catesbeiana tadpoles (Jolly et al. 1978) (Table 1). Cole and Casida (1983) found frogs highly sensitive to eleven different pyrethroids. In general the trans-isomers were less toxic than the cis-isomers. The cis-isomers of permethrin (IR S)-Cypermethrin and Deltametrin were the most toxic to adult R. pipiens (Ibid 1983) (Table 1). (S)-Fenpropathin and (SS)-Fenvalerate were also highly toxic to adult frogs (Table 1). Tadpoles of R. catesbeiana were especially sensitive to (1R, S)-Cypermethrin with an LD50 of 0.04 mg/kg (Ibid 1983) (Table 1), (See Table 1 for data on Pyrethrins, S-Bioallethrin, (IR)-Tetramethrin, Kadethrin, (1RS)-Resmethrin and (IR)-Phenothrin).

Mechanism for Toxicity

Pyrethroid insecticides are recognized as typical neurotoxicants (Vijverberg et al. 1982).

Allethrin, decamethrin and 5-benzyl-3-furylmethyl(+)-cischrysanthemate (NRDC 119) acts on the peripheral nervous system inducing repetitive activity in sensory nerve fibres, sense organs and the distal portion of motor fibres resulting in repetition in the motor end-plate (Evans 1976, van den Bercken 1977, Wouters et al. 1977, Vijverberg and van den Bercken 1979) (Table 2). The repetitive activity of nerves occurs due to a transient increase in the prolongation of sodium permeability. The effect of pyrethroids on nerve fibres is temperature dependent with more pronounced effects at lower temperatures (Wouters et al. 1977, Vijverberg and Ruigt 1981) (Table 2). The effect of pyrethroids on nerve fibres can account for the symptoms of hyperexcitability, convulsive tremors and paralysis prior to death, noted in pyrethroid treated frogs (van den Bercken 1977) (Table 2). Pyrethroids have no direct effect on muscle fibres (Wouters et al. 1977) (Table 2).

Akkermans et al. (1975a) compared the effects of allethrin and several organochlorines and found DDT produced effects similar to those of allethrin (Table 2). DDT and allethrin produced repetitive firing in both the lateral-line organ and cutaneous touch receptors (Akkermans et al. 1975a, van den Bercken et al. 1973).

The frog nerve is not as sensitive to pyrethroids as crayfish nerves (Takeno et al. 1977), which could explain the lower sensitivity of R. catesbeiana tadpoles to permethrin in acute bioassays reported by Jolly et al. (1978) (Table 1).

III HERBICIDES AND FUNGICIDES

a) CARBAMIC ACIDS

MANEB

Short-Term Effects

Maneb (Manganese Ethylene bis dithiocarbamate) is a widely used fungicide in several parts of the world. Dithiocarbamates are known to contaminate plants, soil and water to certain degrees and thus present a hazard to public health (Arias and Zavanella 1979). Little is known of their toxicity and toxic effects to aquatic vertebrates. In tests to examine the survival of T. cristatus in maneb, Zaffaroni et al. (1978) found the fungicide caused 100% mortality within 15 days at 50 ppm (Table 2). It was also noted that males were slightly less susceptible than females (Ibid 1978).

Long-Term Effects

The effects of maneb on the forelimb regenerative ability of newts T. cristatus was examined by Arias and Zavanella (1979) and Zavanella et al. (1979). Both studies found that limbs regenerated in maneb exhibited delayed growth, reduced melanogenesis and malformations (Table 2). An overall delay in growth was also observed in newts exposed to maneb (Zavanella et al. 1979) (Table 2).

Delay in development was also noted by Bancroft and Prahlad (1973) for X. laevis embryos placed in maneb solutions (Table 2). At levels of 1-5 ppm, maneb caused reduced melanogenesis, shortened tail and notochord waviness in developing embryos (Ibid 1973). At these levels no tumors were found in any test animals (T. cristatus) though the spleen weight increased and there was some edema of the soft tissues (Zavanella et al. 1979) (Table 2).

Mechanism for Toxicity

Though little is known about the precise mechanism of lethal effects, Zaffaroni et al. (1978) hypothesized that the toxic effects of maneb are caused by the impairment of cutaneous respiration and osmoregulatory function of the skin.

NABAM

Long-Term Effects

In experiments to determine nabam's effect upon amphibians, it was found that X. laevis embryos exhibited reduced melanogenesis, growth delays, shortened tail and notochord and a difficulty in swimming at various concentrations (Bancroft and Prahlad 1973) (Table 2). Ultrastructural changes were also induced by exposure to nabam (Prahlad et al. 1974) (Table 2). Anderson and Prahlad (1976) examined the synergistic effect of diquat (a herbicide) and nabam and noted many ultrastructural changes and a high mortality rate (See Diquat, Table 2). Thus, nabam, when applied at low concentrations may represent a serious embryotoxic and teratogenic hazard to developing organisms (Bancroft and Prahlad 1973, Anderson and Prahlad 1976).

OTHER CARBAMIC ACID FUNGICIDES AND HERBICIDES

It was reported that preparations of carbamic acids were the most toxic substances in comparison with nine groups of herbicides and insecticides (Zazhivilov 1972).

Sodium diethyldithiocarbamate (NaDEDC) at concentrations of 1-3 mg/L caused severe embryo malformation, growth delay, notochord abnormalities and curvature of the body axis. At levels higher than 3 mg/L, NaDEDC was lethal within 24 hours (Ghate and Mulherkar 1980a). Yalan and eptam are two herbicides used to control weeds in agricultural fields. Though they both exhibit relatively low toxicities to tadpoles (Table 1), it was noted that they produced a decrease in acetylcholinesterase activity in fish similar to the effects induced by sevin (Perevozchenko 1977).

b) DIQUAT AND PARAQUAT

Short-Term Effects

Paraquat was the least toxic of a number of pesticides and herbicides tested using B. woodhousii and P. triseriata tadpoles (Sanders 1970) (Table 1). Johnson (1976) also reported that paraquat had a low toxicity in assays employing two Australian frog species (Adelotus brevis and Limnodynastes peroni) (Table 1). The large difference between the two species demonstrates the wide range in species sensitivity to paraquat.

Exposure to diquat during the egg stage can reduce survival later in development. R. pipiens eggs exposed to 100 mg/L diquat during fertilization or later in development had higher mortality post-hatching than controls (Bimber and Mitchell 1978) (Table 2).

Anderson and Prahlad (1976) also reported diquat to be highly embryotoxic to X. laevis embryos (Table 2). Concentrations higher than 2 ppm caused distortion in body shape and 78% mortality while levels lower than 2 ppm were embryotoxic (Ibid 1976).

Long-Term Effects

No mortality, behavioural or developmental abnormalities were observed in R. temporaria or B. bufo tadpoles or T. vulgaris adults exposed in ponds treated with 1 mg/L diquat (Cooke 1977) (Table 3). Thirty-two days after the pond was treated with diquat, R. temporaria and B. bufo tadpoles were heavier than control tadpoles (Cooke 1977). The increase in tadpole weight was attributed to an algal bloom which had occurred due to the diquat. The algae appeared to be an acceptable food source for the tadpoles. The algae was, however, thought to have reduced the suitability of the pond as a newt breeding site (Ibid 1977).

Dial and Bauer (1984) speculated that paraquat could represent a serious hazard to natural populations of R. pipiens. Field application rates vary from 0.1 to 2.0 ppm for aquatic weed control and it was observed that after 12 days no frogs survived when exposed to 2.0 ppm paraquat (Table 2). Paraquat also caused growth delays, tail abnormalities and poor head development.

c) OTHER HERBICIDES

Toxicity data for eleven herbicides have been reported by Sanders (1970) and Johnson (1976) for North American and Australian amphibians respectively (Table 1). The herbicides tested were: 1) 2,2 DPA, 2) Amitrole-T, 3) DEF, 4) DSMA, 5) Dicamba, 6) Fenoprop, 7) Hydrothol 191, 8) Molinate, 9) Picloram, 10) Sodium arsenite, 11) Trifluralin.

Johnson (1976) noted that fenoprop was the most toxic of the substances he tested against anurans. The thermal tolerance of some species was reduced when exposed to all of the herbicides. Both Sanders (1970) and Johnson (1976) found adult frogs to be more resistant than tadpoles due to physiological and morphological differences.

Monosodium methanearsonate (MSMA) was used in bioassays with Scaphiopus couchii juveniles and adults (Judd 1977) (Table 2). At 477 mg/L MSMA mortality of juveniles was 87% in 1 week, although at 4770 mg/L 100% of the juveniles died within 72 hours, while adults experienced only 20% mortality. High juvenile toad mortality was expected to occur when MSMA is used as a herbicide since the recommended application rate is 8.3 mg/L (Ibid 1977).

A triazine compound, cyanatryn had a 96 hour LC50 of 30 mg/L for R. temporaria tadpoles in static bioassays (Scorgie and Cooke 1979) (Table 1). At 20 mg/L tadpoles were lethargic with occasional spasmodic twitching and did not feed. Body weight declined in increasing concentrations of triazine. In 20 day exposures to cyanatryn within the range of recommended treatment rates, tadpole body weight declined by half from 240 mg to 114 mg (Ibid 1979) (Table 2).

Long-Term Effects

Long-term effects of dichlobenil were found to be similar to those for diquat (Cooke 1977). In field applications, dichlobenil caused no mortality of B. bufo or R. temporaria tadpoles, but weights were increased as a result of elevated food resources due to an algal bloom caused by the dichlobenil (Table 3).

Two compounds with both herbicidal and fungicidal properties, Gesagard 50 and Miedzian 50, were introduced to X. laevis tadpoles to examine the effects on tadpole development (Jordan et al. 1977). At both concentrations tested, all tadpoles in Gesagard 50 died within 72 hours. Those exposed to Miedzian 50 suffered high mortality at the two concentrations. Transferal to tap water did not result in recovery

from the induced effects (Table 2). Application rates for these two compounds are up to 300 times greater than the concentrations used in the tests (Ibid 1977).

Chloranil is a herbicide that causes embryotoxic and other detrimental effects on developing X. laevis embryos (Anderson and Prahlad 1976) (Table 2). At 2.0 ppm chloranil, only 5 percent of the embryos used survived more than 24 hours and these exhibited alterations in development (Anderson and Prahlad 1976).

Defenuron (N-phenyl-N'-methyl urea) is a wide spectrum herbicide that has low toxicity for vertebrates but has been shown to inhibit development of mosquitoes (Paulov 1977). When R. temporaria tadpoles were exposed to 100 ppm defenuron development and metamorphosis were delayed (Table 2). One observation made by Paulov (1977) was the development of "giant" tadpoles. This was due to a multiplication of cells coupled with the inhibition of metamorphosis (Paulov 1977) (Table 2).

d) OTHER FUNGICIDES

Malachite green, a fungicide frequently used in fish culture, was tested in flow-through bioassays with adult Notophtalmus viridescens, R. pipiens and Bufo sp. tadpoles (Bills et al. 1977) (Table 1). Adult newts were more resistant to malachite green than either frog tadpoles or toad tadpoles (Table 1). The tadpoles had a sensitivity to malachite green similar to most fish species tested (Ibid 1977).

Afugan (a fungicide containing pyrasophos) was lethal at 1.0 mg/L to R. temporaria tadpoles (Paulov 1981). Exposures to afugan during larval development resulted in stimulation of growth but inhibition of metamorphosis. Tadpoles exposed to sublethal doses exhibited alterations in enzymatic activity of transaminase (Ibid 1981) (Table 2).

The effects of a number of chlorinated bisphenols were tested on grass frog blood (species not stated) (Flores and Buhler 1974). Most bisphenols were highly toxic to frog blood in vitro, resulting in hemolysis with the degree of hemolysis being directly related to incubation. In vivo exposures to lethal doses of bisphenols did not, however, cause hemolysis (Ibid 1974) (see Hexachlorophene, Table 2).

Anderson and Prahlad (1976) found dichlone to be extremely toxic to X. laevis embryos (Table 2). Concentrations above 0.2 ppm caused 100% mortality within two to four days. Additionally there was inhibition of growth and disruptions of the cranial end resulting in headless individuals.

e) PHENOXY ACID HERBICIDES

The Phenoxy acid herbicides are related to natural plant growth regulators and are widely used in agriculture and park maintenance to control weeds. The LC50 toxicity data for 2,4-D, 2,4,5-T, silvex and Weedar 64 are found on Table 1 (Sanders 1970, Johnson 1976).

Studies examining the effects of 2,4-dichloro-phenoxyactic acid on developing R. temporaria tadpoles revealed no visible effects or tissue residues even after treatment with 50 ppm (Cooke 1972a). In contrast Buslovich and Borushko (1976) found that 1-2 mg/L of 2,4-dichloro-phenoxyactic acid significantly inhibits metamorphosis when in solution with its sodium and diethylamine salts (Table 2).

TCDD (2,3,7,8-tetrachlorodibenzo-p-dioxin) is a highly toxic contaminant of commercial supplies of 2,4,5-T and silvex. TCDD causes teratogenic effects in mammals and fish, yet Beatty et al. (1976) and Neal et al. (1979) reported little if any effect on R. catesbeiana development and survival (Table 2). Following an accident in Italy which released TCDD, Fanelli et al. (1980) found the TCDD level in a toad to be the lowest of the five animal species examined (Table 4).

IV BACTERICIDES

FURANACE

Furanace, a nitrofuran used as a fish bactericide, was highly toxic to R. pipiens tadpoles in static bioassays, compared to the fish and invertebrate species that were tested (Marking et al. 1977) (Table 1). The egg stage was not as sensitive as treatments up to 4 ppm failed to produce any effect on development. Above 4.0 mg/L few embryos survived, either failing to develop or not escaping from the egg mass during hatching (Ibid 1977).

V LAMPRICIDES

TFM

The lampricide TFM (3-trifluoromethyl-4-nitrophenol) has been used to control the sea lamprey in the Great Lakes. In toxicity tests using Hyla versicolor, R. pipiens and R. catesbeiana tadpoles Chandler and Marking (1975) found little difference in species sensitivity (Table 1). TFM is applied in twelve hour exposures and it is doubtful that this would have any effect on amphibians (Chandler and Marking 1975).

VI ORGANICS

a) AROMATIC AMINES

Short-Term Effects

Toxicity studies revealed that of four aromatic amines and two methyl derivatives, acridine was the most toxic followed by quinoline, and its two methyl derivatives, (2-methyl-quinoline,2,6-dimethyl -quinoline), aniline and pyridine (Dumont et al. 1979, Sloof and Baerselman 1980, Davis et al. 1981) (Table 1). All of the compounds caused embryos to become malformed in various ways. These included exogastrulation, edema, formation of blisters and at concentrations

near the 96 hour LC50's, axial lengthening resulting in short, fat larvae with underdeveloped digestive tracts. These effects were more severe in exposures to acridine than the three other amines (Davis et al. 1981). Electron micrographs of the exposed larvae revealed damage to the spinal chord and muscle alterations. In general, larvae, when exposed to sublethal concentrations, were immobile, slower to develop and had abnormal pigmentation (Dumont et al. 1979, Davis et al. 1981) (Table 1).

Greenhouse (1976a,b,c, 1977) assessed the toxicity of a group of organic amines found in jet fuels employed by the United States Air Force. Only N-phenyl-a-napthylamine was found to be toxic to developing frogs (Greenhouse 1976a). Greenhouse (1976c) also found this compound to be teratogenic, whereas octyl-phenyl-a-napthylamine and dioctyldiphenylamine were neither teratogenic nor otherwise toxic to frog embryos probably due to their insolubility (Table 2).

In studies to examine the effect of hydrazine and its methylated derivatives on development of X. laevis Greenhouse (1976b, 1977) determined that these compounds were teratogenic (Table 2). Two types of malformation were observed: i) malformations of the axial skeleton and tail and ii) those of the head and trunk.

This group of compounds is readily soluble in water and were found to be toxic to X. laevis embryos. At a concentration of 1 mg/L hydrazine, methylhydrazine, and symmetrical dimethylhydrazine caused irreversible toxic effects and 100 percent mortality to Xenopus larvae (Greenhouse 1976a) (Table 2). Greenhouse (1976a,b,c, 1977) noted that embryos were able to withstand much higher concentrations of these compounds than were the larval stages.

Mechanism for Toxicity

The mechanism for toxicity of acridine, aniline, pyridine and quinoline was speculated by Davis et al. (1981) to be through the association of the amines with hydrophobic components of cells and tissue (Ibid 1981).

b) AROMATIC HYDROCARBONS

Long-Term Effects

Triturus cristatus adults when exposed to benzene in air at varying frequencies during a 43 day period exhibited a decrease in leukocyte number, while the number of white blood cells increased (Garavini and Seren 1978) (Table 2). Preceding a decrease in red blood cell numbers the red blood cells became enlarged and immature red blood cell numbers increased. These results were interpreted to indicate that benzene was inhibiting blood cell maturation with the degree of inhibition dependent on frequency of exposure (Ibid 1978).

Toluene had some of the same effects on the blood as benzene (Ibid 1978) (Table 2). T. cristatus, after inhaling varying doses of toluene, exhibited initially an increase in red blood cells followed by a reduction in number. There was no change in white blood cell count, but there was a reduction in the phagocytic activity of the neutrophils (Ibid 1978).

Aromatic hydrocarbons can lead to high incidences of cancer (Busbee et al. 1978). A. tigrinum collected from a sewage polluted pond with high concentration of polycyclic aromatic hydrocarbons were found to have abnormal metabolite ratios and a high rate of cancer (Ibid 1978).

c) PHENOLS

Residues

Residues in tissues of R. temporaria adults after injections of 3 mg/kg of either phenol, 3-nitrophenol, 3,5-diethylphenol or 4-aminophenol were similar for all four phenol compounds (Nagel and Urich 1981). Residues declined with time except in the gallbladder and gut, indicating that bilary excretion was minor in comparison to renal elimination (Ibid 1981).

Mechanism for Toxicity

One mechanism of phenol toxicity has been demonstrated for the surfactant nonilphenolethoxylate (Celentano et al. 1979). In R. esculenta skin, the surfactant inhibits active ion transport-coupled water flow. The mechanism for the change in skin permeability was thought to occur as a result of the surfactant affecting protein structure and activity (Ibid 1979).

d) DIOXIN

(See also III e) Phenoxy acid Herbicides)

Long-Term Effects

No mortality of R. catesbeiana tadpoles could be attributed to dosages of 0.025-1.000 mg/kg dioxin applied intra-peritoneally (Neal et al. 1979). Histopathological examination of the tadpoles after metamorphosis revealed no lesions. After adult R. catesbeiana were administered dosages of up to 0.500 mg/kg i.p. dioxin, no mortality occurred during the 35 day observation period. Also no histopathological lesions occurred due to the dioxin (Ibid 1979).

VII METALS

a) CADMIUM

Short-Term Effects

Bioassays to determine the toxicity of cadmium have produced contrasting results (Birge et al. 1979, Canton and Slooff 1982) (Table 1). These discrepancies may be due to inherent differences in flow-through and static techniques (Birge and Just 1975, Birge et al. 1979, Hall and Swineford 1980, 1981), species sensitivity and life stage sensitivity.

Miller and Landesman (1978) exposed X. laevis to varying concentrations of cadmium and observed at 0.001 mg/L 92% of the embryos survived but were severely malformed. In the presence of magnesium (2 mg/L) the toxic effects of cadmium were greatly reduced (Ibid 1978).

Long-Term Effects

Development and survival were severely affected by lengthy exposures to cadmium (Birge et al. 1977, Manson and O'Flaherty 1978) (Table 2). The most severe effects were noted by Birge et al. (1977) when R. pipiens eggs were exposed to cadmium at the cleavage stage (Table 2). No further development took place when the eggs were exposed to cadmium at this stage. At lower concentrations of cadmium survival was severely reduced (Table 2).

At low concentrations (0.03 mg/L) no mortality occurred, though at levels above 0.09 mg/L cadmium, development was inhibited (Canton and Slooff 1982) (Table 1).

Long-term exposures of Notophthalmus viridescens to 2.0-6.75 mg/L of cadmium caused mortality and retarded forelimb regeneration sometimes leading to abnormal limb regeneration (Manson and O'Flaherty 1978) (Table 2). Mortality was 82% at 6.75 mg/L after 51 days exposure, compared to no mortality in the controls. The deaths were partly attributed to weight loss, since all salamanders exposed to cadmium frequently refused food. All salamanders had their forelimbs removed at the elbow before exposure, in order to assess the effect of cadmium on regeneration. Limb regeneration was retarded, but after 76 days, development was similar to controls although severe abnormalities of the forelimbs were more common in the cadmium treated groups (Ibid 1978).

Residues

In areas located near a zinc mill in Poland, five species of amphibians were analyzed for cadmium residues (Dmowski and Karolewski, 1979) (Table 4). The residues in organisms from contaminated areas were higher than the control location. Food chain accumulation was observed through the different trophic levels in this study.

Canton and Slooff (1982) observed a concentration factor of 130 times in X. laevis exposed for 50 days to a solution of cadmium in water (Table 4).

Mechanism for Toxicity

Cadmium exerts its toxicity on amphibians in a number of ways. There is strong evidence to suggest that cadmium effects epithelial cells by inhibiting active ion transport by combining with sulfhydryl groups in the cell membrane and decreasing membrane potential (Kanno et al. 1978, Arita et al. 1979, Arhem 1980, Takada and Hayashi 1981a,b) (Table 2). The degree of inhibition of ion transport however, varies between different epithelial tissue (Hillyard and Gonick 1978, Takada and Hayashi 1978a,b) (Table 2). Cadmium appears to affect nerve tissue, at least in the heart, by suppressing the release of acetylcholine (Hayashi and Takayama 1978) (Table 2). Cadmium has been shown to cause a depression in the rod receptor cells in the retina of R. catesbeiana (Fox and Sillman 1979). Cadmium had a much greater effect on the rod cells than either Pb or Hg (Ibid 1979) (Table 2).

Treatment of R. nigromaculata eggs, prior to the first cleavage, at a level of 4 mg/L or higher, has been shown to reduce the number of primordial germ cells in the later developmental stages (Hah 1978) (Table 2). Reduction of primordial germ cells can lead to smaller gonads and partial or complete sterility (Bounoure et al. 1954).

b) LEAD

Short-Term Effects

Lead was highly toxic to Gastrophryne carolinensis eggs between fertilization and 4 days posthatching in continuous flow bioassays (Birge et al. 1979) (Table 1).

X. laevis, exposed during the embryo stage to 0.001-10 mg/L lead in static bioassays, resulted in 100% mortality at 10 mg/L and 82% deformities with 18% mortality at 0.001 mg/L (Miller and Landesman 1978). The toxic effect however was reduced when 2.0 mg/L or greater of magnesium was added (Ibid 1978).

Adult anurans appear to be much less sensitive to lead (Kaplan et al. 1967). R. pipiens adults exposed to lead nitrate in daily renewable bioassays had a 96 hour LC50 of 105 mg/L (Table 1). Frogs

exposed to lead had a permanent loss of semi-erect posture (Table 2). Other toxic effects observed were a sloughing of the integument, muscular twitching, salivation, sluggishness and death (Table 2).

Long-Term Effects

Exposure of R. nigromaculata eggs to lead, prior to the first cleavage, resulted in a reduction of the number of primordial germ cells later in development (Hah 1978) (Table 2).

Dilling and Healey (1926) reported lead salts had a severe inhibitory effect on the development of frogs causing a slow down in growth and abnormal development (Table 2).

Residues

Field collected R. esculenta from the River Toce had higher lead levels in tadpole tissues than the adult stage (Baudo 1976). He interpreted the difference in lead levels in the tadpole and adult to be the result of the differences in the lead levels of their foods. Tadpoles primarily eat vegetation which had higher lead levels than the insects eaten by adults. Adult females had high levels of lead in their liver (7.56 mg/kg) while the highest levels in males were found in the skin (3.66 mg/kg) (Ibid 1976).

Other studies have found similar results for lead accumulation. The liver and heart were found to have the highest lead concentrations in frogs (species not reported) in Katowice, Poland (Dec et al. 1979) (Table 2). Similarly, after X. laevis had ingested worms containing lead for four weeks, adults had the highest lead residues in the liver (30.75 mg/kg wet wt), kidney (81.32 mg/kg) and bone (55.98 mg/kg) with lower levels in the skin and muscle (less than 12.0 mg/kg) (Ireland 1977) (Table 4). Ireland (1977) compared lead levels in X. laevis with Peromyscus and concluded that since amphibians had much higher levels, amphibians may be important in concentrating lead in the food web.

Dmowski and Karolewski (1979) suggested amphibians pose a potential hazard due to food chain accumulation. Residue levels in amphibians from an area close to a zinc mine in Poland were high due to accumulation via prey species (Ibid 1979) (Table 4).

Mechanism for Toxicity

Binding of lead in amphibian skin has been shown to be the result of the affinity of melanin for metals (Ireland et al. 1979) (Table 2). X. laevis adults exposed to similar lead levels were either dark adapted to increase the melanophore number, or light adapted to decrease melanophore number. Toads that were dark adapted accumulated significantly more lead than light adapted toads, implicating melanin as the lead binding site in the skin (Ibid 1979) (Table 2).

Lead exerts its toxic effects on amphibians in a number of ways. Kaplan et al. (1967) concluded that a reduction in red and white blood cells in R. pipiens adults exposed to lead nitrate was likely the result of an inhibition of the hematopoietic tissues.

Discolouration of the liver reported for Necturus (Dawson 1933), R. catesbeiana tadpoles (Barrett 1947) and R. pipiens adults (Kaplan et al. 1967), is thought to indicate either that phagocytosis of red blood cells is occurring or that lead is being metabolized and excreted through the gallbladder (Kaplan et al. 1967) (Table 2).

Lead can also affect vision. Dissected R. catesbeiana retina treated with lead caused a depression of the amplitude of the rod receptor cells (Fox and Sillman 1979). The effect was reversible when lead levels were reduced (Ibid 1979).

The main physiological mechanism of lead toxicity is thought to be the binding of lead to sulfhydryl groups and disruption of sulphur bridges (Celentano et al. 1979). This would change the structure of membrane proteins leading to changes in ionic channels (Ibid 1979).

c) MAGNESIUM

Short-Term Effects

Magnesium is an essential element that can compete with other metal ions and reduce their toxicity to developing embryos (Miller and Landesman 1978). Magnesium at 2.0-200 mg/L greatly reduced teratogenicity in X. laevis tadpoles exposed to mercury, lead, cadmium and manganese. Since magnesium levels in amphibians are constant regardless of environmental levels, there may be competition for some common carrier mechanism that transports divalent cations into the embryos, thereby reducing metal toxicity (Ibid 1978).

d) MERCURY

Short-Term Effects

Tests to determine the toxicity of twenty-two metals to amphibians revealed mercury to be the most toxic to Gastrophryne carolinensis eggs (Birge et al. 1979) (Table 1). The toad was more sensitive to mercury than three species of fish that were also tested. Ghate and Mulherkar (1980b) reported 100% mortality of Microhyla ornata eggs at the gastrula stage when exposed to mercuric chloride at concentrations above 3 mg/L (Table 1). Mercuric chloride also retarded development,

caused blisters on the body and curvature of the spine. The tadpole stage had a slightly lower resistance to mercuric chloride. Sublethal doses caused the tadpoles to be sluggish and distension of the body cavity was observed (Ibid 1980b) (Table 2). X. laevis exhibited many of these symptoms when they were treated with mercury at the egg stage (Miller and Landesman 1978).

Birge et al. (1977) examined the survival of G. carolinensis eggs treated with mercury before hatching. At the lowest concentration of mercury (0.146 ppm) 61% of the hatched embryos died within four days of hatching. Eleven percent of the hatched embryos exhibited gross congenital deformities at hatching (Table 2).

Studies using R. pipiens embryos at different stages revealed death and serious developmental defects can be induced by trace amounts of methylmercury (Dial 1976) (Table 2). Frog gastrulae appeared to be the most sensitive embryonic stage of the three stages tested.

Long-Term Effects

Mercury caused severe damage to R. nigromaculata germ cells in eggs prior to the first cleavage (Hah 1978) (Table 2). Further proliferation of primordial germ cells was much reduced. Loss of primordial germ cells can result in reduced gonads and complete or partial sterility (Bounoure 1954).

Residues

Mercury residues can be accumulated by most aquatic biota (Cox et al. 1975). Cox et al. (1975) found Ranidae tadpoles to accumulate moderately high levels (up to 6.41 mg/kg) when exposed to low levels of mercury. Less than 1% of the mercury residues in the tadpoles was in the methyl form (Ibid 1975) (Table 4).

In Yugoslavia mercury residues were measured in a number of amphibian species from sites with varying levels of mercury contamination (Byrne et al. 1975). The liver had the highest concentrations ranging from 0.19 mg/kg in frogs collected in uncontaminated areas, to 25.9 mg/kg in R. temporaria from a site near a mercury plant. The kidney also had high residue levels with lung, egg and muscle tissue accumulating almost an order of magnitude less mercury. The liver and kidney contained only a small fraction of the mercury in the methyl form, whereas the muscle contained almost 100% methylmercury (Ibid 1975) (Table 4).

Dustman et al. (1972) analyzed organisms from lake St. Clair to determine their mercury content. Great Blue herons (Ardea herudias) and common terns (Sterna hirundo) had the highest levels while frogs

(R. pipiens and R. catesbeiana) had the lowest (Table 4). Though frogs form part of the diet of herons, the high residue levels were believed to have been caused by the fish they consumed which contained high levels of total mercury (Ibid 1972).

Accumulation of mercury in the liver and kidneys is common in other animals. In mammals mercury binds to metallothionien and there is evidence that this is the primary binding site in amphibians as well (Mehra et al. 1980).

Mechanism for Toxicity

Mercury has a high affinity for sulfhydryl groups and may affect enzymes and proteins associated membranes with the most severe effects occurring at nerve membranes (Dial 1976, Arhem 1980, Ghate and Mulherkar 1980b).

Mercury has been reported to exert its toxic effect on amphibians in a number of ways. Lesions in the rods in eyes have been shown to occur in the perfused retina of R. catesbeiana when exposed to mercury (Fox and Sillman 1979) (Table 2).

Chang et al. (1975) after observing complete mortality of R. pipiens tadpoles within 48 hours of exposure to low levels of mercury, (0.05 ppm), speculated that the distention of the body cavity (Chang et al. 1975, Ghate and Mulherkar 1980b) and mortality was the result of a disturbance of the osmotic regulator system caused by mercury.

Mercury at low concentrations (0.01-0.07 mg/L) can inhibit cell growth while at higher concentrations (0.1 mg/L) actually promote cell growth and tissue development (Chang et al. 1976). The actual mechanism for this dose dependent effect is not yet understood.

e) SELENIUM

Short-Term Effects

Although selenium is an essential trace element it can be acutely toxic in high concentrations. X. laevis embryos (at the gastrula stage) and tadpoles exposed to low levels of sodium selenite in daily renewal bioassays had increased survival compared to controls, but mortality rapidly increased at 2.0 mg/L and higher (Browne and Dumont 1979). Although toxicity of selenium to tadpoles was not high (Table 1) 50% of those hatched from exposed eggs exhibited abnormalities in development. Exposures to selenium during the egg stage caused spine curvatures, tail flexures and malformed heads. Tadpoles treated with selenium exhibited erratic swimming behaviour and sluggishness as well as epithelial blisters, abdominal edema and degeneration of muscle cells (Browne and Dumont 1979, 1980).

Although toxicity for the egg stage was not measured, eggs experienced a lower rate of mortality than tadpoles. The gelatinous coating of the egg was thought to present a permeability barrier affording partial protection to the egg.

In continuous flow bioassays, G. carolinensis eggs were much more sensitive to selenium with a 96 hour LC50 of 0.09 mg/L (Birge et al. 1979) (Table 1).

Residues

X. laevis eggs exposed to 5 mg/L of selenium exhibited a rapid uptake for the first 24 hours (Browne and Dumont 1979). Up to 50% of these residues were retained in tadpoles for extended periods of time (Ibid 1979).

f) ZINC

Short-Term Effects

Zinc was the second most toxic metal of twenty-two metals tested using G. carolinensis eggs (Birge et al. 1979) (Table 1). B. boreas tadpoles exhibited higher resistance to zinc concentrations. Concentrations of 39 mg/L of zinc caused complete mortality within 12 hours whereas exposure to 0.1 mg/L caused no mortality even after 61 days of exposure (Porter and Hakanson 1976) (See Acid Mine Drainage Table 2).

Birge et al. (1977) found G. carolinensis eggs exposed to sediment bound zinc, did not suffer high mortality or teratogenesis at hatching (Table 2).

Residues

In laboratory exposures, the zinc residues accumulated by T. granulosa had a half life of 3 1/3 years (Willis and Valett 1978). Loss of zinc was temperature dependent with higher temperatures shortening the half life. The skin, muscle, blood and liver had the highest residue levels and together these tissues accounted for 70% of the total body burden (Ibid 1978) (Table 2). Dec et al. (1979) stated that zinc accumulated in the heart of frogs taken from the environs of a steel plant in Poland (Table 3).

Mechanism for Toxicity

Zinc appears to exert its toxic effect in a manner similar to other metals. In myelinated nerve fibres zinc has been shown to slow the kinetics of the potassium system and alter sodium permeability (Arhem

1980). Zinc can also change the sodium and potassium kinetics across epithelial cell membranes (Kanno et al. 1978) (Table 2). These results were thought to indicate that changes in membrane permeability are in part due to zinc combining with sulfhydryl groups in the cell membrane (Kanno et al. 1978) and in part due to the tendancy of the zinc to form complexes with certain ligands.

g) OTHER METALS

Short-Term Effects

In continuous flow bioassays Birge et al. (1979) measured the 96 hour LC50s of a number of metals to G. carolinensis eggs (Table 1).

Components of acid mine drainage (iron, zinc, copper) were tested to determine their toxicity to amphibians (Porter and Hakanson 1976). Larvae were able to tolerate 20 mg/L of iron but at 30 mg/L there was complete mortality. Copper was toxic to B. boreas at 3.7 mg/L. When zygotes were placed in a solution of mine drainage 100% mortality occurred within 12 hours (See Acid Mine Drainage, Table 2).

R. pipiens eggs exposed in static bioassays to copper sulfate at low levels, exhibited no toxic effects (Landé and Guttman 1973) (Table 2). The tadpole stage was much more sensitive (Ibid 1973) (Table 1). Higher mortality in the tadpole stage has also been reported by Kaplan and Yoh (1961).

Residues

Residues of chromium, manganese and copper measured in R. esculenta from the Toce River, Italy, were 0.42-9.22 mg/kg, 23-958 mg/kg and 7.63-32.4 mg/kg respectively in the tadpole stage (mean annual concentrations: chromium 0.01 mg/L; copper 0.005 mg/L; manganese 0.06 mg/L) (Baudo 1976). Neometamorphosed frogs had significantly lower chromium and manganese levels (0.47 mg/kg and 25.0 mg/kg respectively) than the tadpole stage. The significant drop in chromium and manganese residues in the neometamorphosed frog was attributed to a change in the metal levels in their food, as the tadpoles shift from being primarily herbivorous to being mainly insectivorous adults. In adults copper was most highly concentrated in the liver (61-63 mg/kg) while chromium and manganese had the highest concentrations in the skin (1.27-2.47 mg/kg and 30.3-67.9 mg/kg respectively) (Ibid 1976).

Mechanism for Toxicity

Nickel and copper have been shown to slow down the kinetics of the potassium system in myelinated nerve fibres with copper having the greatest negative effect (Arhem 1980). Manganese and cobalt had no

effect. All four metals, however did alter sodium permeability with copper causing the greatest decrease and nickel the smallest. The toxicity of these metals is thought to be due to a similar mechanism as described for zinc, where the metals combine with sulfhydryl groups in cell membranes and alter membrane permeability (Ibid 1980).

Pytasz et al. (1980) reported a decrease in the general metabolic rate of frogs in an area close to a metallurgic worksite (See Metals, Table 2).

VIII RADIOACTIVE ISOTOPES

a) CESIUM

Residues

Radiocesium content was measured in Hyla cinerea adults collected from a contaminated swamp in South Carolina (Dapson and Kaplan 1975) (Table 3). Variability in body burdens was attributed to an inverse relationship between body burden and body size plus a large degree of variability in radiocesium content of food items. The biological half-life for unfed adult frogs at 20-30°C was 30.1 days (Ibid 1975) (Table 3).

b) COBALT

Long-term Effects

Two B. canorus adults affixed with tags containing cobalt^{60} and released in the field for 41 weeks exhibited no ill-effects from the exposure (Karlstrom 1957) (Table 3).

c) STRONTIUM

Short-Term Effects

Birge et al. (1979) reported strontium to be one of the less toxic metals tested using G. carolinensis in flow-through toxicity tests (Table 1).

Long-Term Effects

T. granulosa adults exposed to radiostrontium for 25 days at 10°C exhibited an initial rapid uptake for 72 hours which declined without reaching equilibrium at day 21 (Willis and Valett 1971, 1976) (Table 2). The biological half-life was 136 days which was similar to the half-life in acute exposures using intraperitoneal injections. Significant concentrations were found in the blood, liver, skin and

muscle although the bone contained the largest portion. The biological half-life of radiostrontium for R. pipiens at 10°C after intraperitoneal injections was 22 days (Ibid 1971) (Table 2).

d) IONIZING RADIATION

The effects of ionizing radiation on amphibian development have been comprehensively reviewed by Brunst (1965).

IX OTHER COMPOUNDS

a) INDUSTRIAL EFFLUENTS

X. laevis embryos exposed to sour water from a coal-gasification process exhibited high mortality and teratogenic effects (Dumont and Schultz 1980) (Table 1). The teratogenic effects ranged from tail flexure to gross distortions of the longitudinal axis, as well as the optic, cephalic, visceral and pericardial tissues. Loss of balance and reduced pigmentation were also common. The sour water was typical of most coal gasification processes and contained straight chain carboxyl acids, and varying amounts of aromatic hydrocarbons, aromatic amines and sulphur compounds (Ibid 1980).

R. ridibunda collected near the Katowice ironworks in Poland had significantly reduced metabolic rates that were inversely related to the distance from the mill (Pytasz et al. 1980). The reduction in metabolism was directly related to metal residues in tissues (Ibid 1980) (See Metals, Table 2).

b) ACID DRAINAGE

Short-Term Effects

Runoff from some rock types (particularly those containing pyrite), can be very acidic and have elevated metal levels which are highly toxic to amphibians (Huckabee et al. 1975, Mathews et al. 1976, Mathews and Morgan 1982). Eighty-three percent mortality of Leurognathus marmoratus larvae occurred in Anakeesta acid metal leachate (Huckabee et al. 1975). The 72 hour LC50 for L. marmoratus larvae exposed to the leachate from the same rock deposit was 33% (Mathews and Morgan 1982) (Table 1). The combined effects of acidity plus elevated metals, particularly zinc, copper and aluminum, were considered to be the toxic components of the leachate (Huckabee et al. 1975, Mathews and Morgan 1982). The toxicity of the leachate was reduced by the addition of sodium hydroxide (NaOH) (Mathews et al. 1976). The sodium hydroxide raised the pH and reduced the metal levels by decreasing solubility and inducing the formation of metal complexes. Neutralized leachate had a 72 hour LC50 of 113% (Mathews et al. 1976).

Porter and Hakanson (1976) attributed the absence of B. boreas from a creek in Colorado to a combination of toxic effects from acid mine drainage that was entering the stream. The stream water was non-toxic only after a 1,000 fold dilution. The toxic components of the drainage were hydrogen ion, copper, zinc and iron (Ibid 1976) (See Acid Mine Drainage, Table 2).

X WATER QUALITY

a) NITRITE

Short-Term Effects

Toxicity values for Ambyostoma texanum larvae suggest that the salamander is extremely sensitive to nitrite (Huey and Beitinger 1980) (Table 1). Nearly 100% mortality was observed at levels as low as 2.5 mg/L nitrite. However, when chloride concentrations were increased, larvae suffered no mortality in exposures to 10 mg/L nitrite. Chlorides appeared to compete with nitrite in ionic uptake on respiratory surfaces, resulting in reduced uptake of nitrite (Huey and Beitinger 1980).

b) pH

Acidity is an important water quality parameter that can affect amphibians directly and indirectly by influencing the toxicity of other contaminants. Only the direct effects will be discussed in this section. Indirect effects of pH are referred to in the section for the chemical where an effect has been noted.

Short-Term Effects

The direct short term effects of elevated hydrogen ion concentration have been observed at all lifestages. Fertilization of R. pipiens eggs was reduced below pH 6.5 (Schlichter 1981) (Table 2). Below pH 4.9 complete failure of egg development was attributed to low sperm motility. Fertilization was more susceptible to depressed pH than either activation or cleavage. The optimal pH for normal egg development was concluded to be above pH 6.0 (Ibid 1981). Gosner and Black (1957) estimated levels of pH that would allow 50% or more of embryos to survive in lab bioassays with tap water acidified with hydrochloric acid. The species and pH levels were as follows:

Acris crepitans	4.7
H. versicolor	4.3
Rana catesbeiana	4.3
R. palustris	4.3
H. crucifer	4.2
R. clamitans melanota	4.1

R. pipiens	4.1
Pseudacris nigrata	4.1
R. sylvatica	3.9
Hyla andersoni	3.8
R. virgatipe	3.8

Gosner and Black (1957) also reported the observed lethal level for all these species (Table 2).

Acid stressed embryos exhibited morphological and behavioural abnormalities that appeared to be a result of shrinkage of the perivitelline membrane. Embryos were cramped in the eggs and had difficulty hatching (Ibid 1957). Short-term pH depressions were also toxic to developing embryos. R. sylvatica eggs exposed for short periods to pH 3.5-3.7 exhibited increased mortality with the degree of toxicity dependent upon the lifestage and length of exposure (Ibid 1957).

A. maculatum and A. jeffersonianum had marked reductions in hatching success in lab and field exposures to pH less than 6.0 (Pough 1976, Pough and Wilson 1977) (Table 3, Table 2). Tolerable pH for A. maculatum was 6 to 10 while A. jeffersonianum tolerated pH 7 to 9 (Pough and Wilson 1977) (Table 2). A. maculatum experienced greater than 60% mortality in ponds with pH 6.0. Developmental abnormalities at pH 6.0 included stunted gills, swelling near the heart, failure of yolk plug retraction and curved spines (Pough 1976).

Optimal development of amphibians is dependent upon both pH and temperature (Punzo 1983). Punzo (1983) by exposing Ambystoma texanum to varying temperature and pH values discovered that optimum hatching occurred within a temperature range of 10 to 15°C with a pH range of 7 to 9 (Table 2). Beyond these ranges hatching success dropped to no survival at the extremes tested (Punzo 1983).

Dunson and Connell (1982) observed a marked inhibition of hatching of X. laevis eggs at a pH of 3.9 (sulfuric acid) and a pH of 4.3 (bog water) (Table 2). In natural breeding ponds A. maculatum hatching was completely arrested at a pH of 3.6 caused by acid precipitation (Nielsen et al. 1977) (Table 3). However, Cook (1983) found no significant correlation between pond pH and embryonic mortality for two species of salamander inhabiting ponds in an area of acidic rainfall (Table 3).

R. catesbeiana eggs and larvae exposed at sites in a stream draining a bog which exhibited a natural pH gradient from 3.9 to 6.6, showed a positive relationship between mortality and pH (Saber and Dunson 1978) (See Bog Water, Table 3). R. catesbeiana embryos and larvae between stage 18 and 26 (Gosner 1960) were extremely sensitive to pH, whereas R. catesbeiana and R. clamitans larvae beyond stage 26

were not sensitive. Mortality of R. catesbeiana and X. laevis was also greater in coloured water than clear water at the same pH, indicating some other toxic component of the bog water (Saber and Dunson 1978) (Table 3).

B. bufo and R. Temporaria exposed to pH 3.75-8.0 exhibited dramatic increases in mortality between pH 4.25 and 4.75 (Beebee and Griffin 1977) (Table 3). B. calamita was much more sensitive with increased mortality of eggs and larvae below pH 6.0 and complete mortality at pH 4.75. Two abnormalities were observed preceding some instances of mortality in the acidic exposures, an inhibition of growth beyond stage 5 (Taylor and Kollros 1946) and a bloating of the body late in development (Beebee and Griffin 1977) (Table 3).

Long-Term Effects

pH affects the distribution of many amphibian species (Gosner and Black 1957, Cooke and Frazer 1976, Pough 1976, Beebee and Griffin 1977, Pough and Wilson 1977, Hagstrom 1977, 1980, Strijbosch 1979, Beebee 1983) (Tables 2 and 3). Most species avoid acid waters, although since there is a range in species sensitivities to acidity, some species such as R. sylvatica and R. clamitans which are more tolerant, will be found in more acidic habitats (Gosner and Black 1957, Saber and Dunson 1978). Other species such as A. jeffersonianum will be found only in circumneutral habitats (Pough and Wilson 1977). Although vegetation and other physical/chemical parameters are important in amphibian distribution, pH was found by Strijbosch (1979) to be one of the most important habitat characteristics. Amphibians can detect soil pH gradients and so can actively select acceptable habitat acidity (Mushinsky and Brodie 1975).

Mechanism for Toxicity

Fromm (1981) examined osmoregulation across R. pipiens skin in vitro at varying pH levels. He found at low pH that the outer skin surface experiences a decrease in the flux of sodium ions into the skin, reducing overall sodium transport. He concluded that there was an ionoregulatory failure as a result of decreased uptake rather than an excess loss of ions.

Disruptions of ionic balance were speculated by Gosner and Black (1957) and Pough (1976) to be a cause of egg mortality. Gosner and Black (1957) suggested that this was the cause of shrinkage of the perivitiline membrane during egg development (Table 2).

c) SALTS

Short-Term Effects

The toxicity of three chloride salts (potassium chloride, sodium chloride, calcium chloride) were measured using R. breviceps tadpoles in static bioassays (Mahajan et al. 1979). Tadpoles were most sensitive to potassium chloride with 73% mortality in 72 hours when exposed to 3000 mg/L. Sodium chloride above 5000 mg/L was highly toxic. Calcium chloride did not produce significant mortality until exposure to concentrations of 6000 mg/L and higher. All observed mortality was associated with respiratory failure and possible osmoregulatory imbalances resulting in sluggishness and loss of balance (Ibid 1979).

Sea water is highly toxic to amphibians. R. pipiens adults died within one hour of being placed in sea water (Bentley and Schmidt-Nielsen 1971) (Table 3). Post mortem analysis revealed that the frogs had accumulated large amounts of sodium through the skin and by drinking (Ibid 1971).

XI EVALUATION OF AMPHIBIANS AS MONITORS

a) CHEMICALS REPRESENTING A POTENTIAL HAZARD TO AMPHIBIANS

Wildlife toxicological concerns can be divided into two broad areas, direct concerns for wildlife health and the use of wildlife as indicators of enviornmental quality. In both cases, it is important to be able to carry out controlled laboratory experiments and as well as field studies.

The relative toxicity of various chemicals to amphibians are summarized in Table 5. Among the most toxic are the salts of heavy metals. The most sensitive life stage to metal toxicity is the egg followed by the larval and then adult stage (Birge et al. 1979). Birge et al. (1979) reported that treatment of the egg stage with metals (mercury, cadmium) produced mortality and teratogenisis in larvae at hatching (Table 2). Mercuric chloride, at less than 1 ppm, caused mortality, abnormal development and retarded growth (Ghate and Mulherkar 1980b) (Table 2). Though differences in species sensitivity have been observed, tolerance to pollutants must also be a function of the sensitivity of specific life stages. Birge et al. (1979) reported the sensitivity of G. carolinensis eggs to cadmium, but X. laevis larvae resistance was two orders of magnitude greater (Canton and Slooff 1982) (Table 2). Adult R. pipiens were more resistant to lead than G. carolinensis eggs as reported by Birge et al. (1979) (Kaplan et al. 1967) (Table 2). Thus different life stages display greater resistance to contaminants than others within the life cycle of one species.

Metals are not only toxic to amphibians but are also readily accumulated in body tissues and can be concentrated as they are passed up the food web (Byrne et al. 1975, Baudo 1976, Browne and Dumont 1979). Metals can be concentrated in amphibian tissues many times environmental levels and in some instances the metals are retained for long periods of time (Ireland 1977, Browne and Dumont 1979, Canton and Slooff 1982). Dial (1976) stated that in most environmental situations mercury stress would not be a factor due to low environmental levels, but that at point sources, such as industrial effluent disposal, populations R. pipiens could be seriously affected due to their acute sensitivity to mercury.

Of the insecticides the cyclodienes are the most toxic (Table 5). In particular endosulfan (Korschgen 1970, Gopal et al. 1981) endrin, toxaphene (Sanders 1970, Hall and Swineford 1981, 1982), and dieldrin (Sanders 1970) are the most toxic (Table 1). Cyclodiene toxicity not only can result in mortality but can have long-term effects on behaviour, morphology and development (Rane and Mathur 1978, Hall and Swineford 1980, Marchal-Segault and Ramade 1981). There is some indication that the tadpole stage is more sensitive to cyclodienes than the egg stage. For lindane X. laevis experienced greater mortality in exposures as tadpoles than during egg stage (Marchal-Segault and Ramade 1981) (Table 2). Hall and Swineford (1980) reported R. sphenocephala experienced no mortality at the egg stage but increased mortality at the tadpole stage when exposed to endrin (Hall and Swineford 1980).

Larval amphibians have exhibited an ability to store organochlorine insecticides in their fatty tissues, most importantly the tail (Cooke 1970, 1973a, 1974, Licht 1976b, Lyons et al. 1976). During resorption of the tail at metamorphosis, these residues are mobilized and result in mortality among young frogs (Cooke 1970). Thus the hazard presented by some organochlorines affects all stages of the amphibian life cycle up to the adult stage, where mobilization and excretion of residues is more efficient (Niethammer et al. 1984).

Organophosphate insecticides presented the widest range in toxicity to amphibians (Table 5). Many organophosphates had extremely high toxicity (carbophenthion, parathion, malathion) (Sanders 1970) while others such as orthene were toxic only in large amounts (Lyons et al. 1976, Geen et al. 1984) (Table 1).

Symptoms of organophosphate poisoning were abnormal behaviour and development (Lyons et al. 1976, Marchal-Segault 1976), and alterations in thermal resistance (Johnson and Prine 1976) (Table 2).

Organophosphate residues can be concentrated above background levels (Lyons et al. 1976). Parathion and fenthion can be accumulated to levels lethal to ducks (Hall and Kolbe 1980) suggesting that organophosphate concentration in the food web could represent a serious hazard at higher trophic levels.

Carbamate, organochlorine (except the cyclodienes) and pyrethroid insecticides were all moderately toxic to amphibians resulting in behavioural and developmental abnormalities as well as mortality (Cooke 1970, 1979, Sanders 1970, Rzehak 1977, Jolly et al. 1978, Harri et al. 1979) (Tables 1 and 2).

Cooke (1972b) suggested that the levels of some insecticides necessary to produce slight hyperactivity in amphibian larvae are not encountered in the field in Britain and that hazards from insecticides would only occur due to misuse or accidents.

Acidification alone can impair egg and larval development (Gosner and Black 1957, Pough 1976, Pough and Wilson 1977, Schlichter 1981). The lowest pH that embryos can survive are listed on page 82. In view of the acidification of lakes and other smaller water bodies in eastern North America and much of Europe the effect on a number of species of amphibians should be a matter of concern. In addition to the direct effects of increasing acidification there are chronic effects on the ecology of the pond, changes in habitat structure, predator-prey relationships, food resources etc., which may profoundly affect anuran populations.

Amphibians which breed in temporary ponds may be the group most vulnerable to acidic precipitation. These temporary bodies of water often refill with rainwater and snowmelt during the early spring. Acidic water entering a dry pond is not diluted and may encounter little buffering as it enters through surface flow rather than as ground water. Semkin and Jeffries (1986) found that ionic concentration in the snowmelt was 2 to ten times greater than that of pre-melt material, the so-called "acid-shock".

The herbicides and fungicides include chemicals with a wider range in their toxicity to amphibians (Table 5). Malachite green had a 96 hour LC of 0.068 mg/L while DSMA had a 96 hour TL50 of 271 mg/L. Behavioural and developmental abnormalities have been observed after short or long term exposures to some herbicides (Arias and Zavanella 1979, Scorgie and Cooke 1979, Ghate and Mulherkar 1980) (Table 2). Herbicides may exert indirect effects by altering the environment, for example the use of 2,4-D, which has low toxicity to amphibians, has been considered to reduce the diversity of prey species in an area and thus cause a reduction in amphibian numbers (Cooke 1972b).

A symptom of toxicity common to many of the compounds discussed in this review is an alteration to some degree of normal behaviour. This has been shown to put amphibians at a selective disadvantage which could result in depletion of some natural populations by overpredation (Cooke 1971, Lyons et al. 1976, Hall and Kolbe 1980, Mohanty-Hejmadi and Dutta 1981) (Table 2).

TABLE 5
RANKING OF TOXICITY OF POLLUTANTS BASED ON THE 96 HR LC50

CHEMICAL	SECTION	SPECIES	STAGE	96 HR LC50 mg/L	RE
Mercury	Metals	Gastrophryne carolinensis	egg	0.001	19
Endosulfan	Insecticide	Rana tigrina	tadpole	0.0018	5
Endrin	Insecticide	Rana catesbeiana	tadpole	0.002	10
Zinc	Metals	Gastrophryne carolinensis	egg	0.01	19
Carbophenthion	Insecticide	Pseudacris triseriata	tadpole	0.028	2
Chromium	Metals	Gastrophryne carolinensis	egg	0.03	19
Arsenic	Metals	Gastrophryne carolinensis	egg	0.04	19
Cadmium	Metals	Gastrophryne carolinensis	egg	0.04	19
Copper	Metals	Gastrophryne carolinensis	egg	0.04	19
Lead	Metals	Gastrophryne carolinensis	egg	0.04	19
Aluminum	Metals	Gastrophryne carolinensis	egg	0.05	19
Cobalt	Metals	Gastrophryne carolinensis	egg	0.05	19
Germanium	Metals	Gastrophryne carolinensis	egg	0.05	19
Nickel	Metals	Gastrophryne carolinensis	egg	0.05	19
Toxaphene	Insecticide	Rana sphenocephala	larval	0.032-0.054	8
Malachite green	Fungicide	Bufo sp.	tadpole	0.068	13
Selenium	Metals	Gastrophryne carolinensis	egg	0.09	19
Thallium	Metals	Gastrophryne carolinensis	egg	0.09	19
Tin	Metals	Gastrophryne carolinensis	egg	0.09	19
Dieldrin	Insecticide	Pseudacris triseriata	tadpole	0.10	2
Trifluralin	Herbicide	Bufo woodhousii	tadpole	0.10	2
Guthion	Insecticide	Bufo woodhousii	tadpole	0.13	2
TDE	Insecticide	Bufo woodhousii	tadpole	0.14	2
Aldrin	Insecticide	Bufo woodhousii	tadpole	0.15	2
Strontium	Metals	Gastrophryne carolinensis	egg	0.16	19
Malathion	Insecticide	Pseudacris triseriata	tadpole	0.20	2
Antimony	Metals	Gastrophryne carolinensis	egg	0.30	19
Methoxychlor	Insecticide	Pseudacris triseriata	tadpole	0.33	2
DEF	Herbicide	Bufo woodhousii	tadpole	0.42	2
Heptachlor	Insecticide	Bufo woodhousii	tadpole	0.44	2
Furanace	Fungicide	Rana pipiens	larval	0.77	17
DDT	Insecticide	Pseudacris triseriata	tadpole	0.80	2
Parathion	Insecticide	Pseudacris triseriata	tadpole	1.0	2
Piperonyl butoxide	Insecticide	Pseudacris triseriata	tadpole	1.0	2
Nitrite	Nitrite	Ambystoma texanum	larval	1.090	6
Hydrothol 191	Herbicide	Bufo woodhousii	tadpole	1.2	2
Manganese	Metals	Gastrophryne carolinensis	egg	1.42	19
Naled	Insecticide	Pseudacris triseriata	tadpole	1.7	2
2,2-DPA	Herbicide	Limnodynastes peroni	tadpole	2.0	1
Lindane	Insecticide	Pseudacris triseriata	tadpole	2.7	2
Tungsten	Metals	Gastrophryne carolinensis	egg	2.90	19
Amitrole-T	Herbicide	Adelotus brevis	tadpole	3.0	1

Benzene hexachloride	Insecticide	Bufo woodhousii	tadpole	3.2	2
Acridine	Organics	Xenopus laevis	egg	3.6	18
Fenitrothion	Insecticide	Rana clamitans	tadpole	4.9	7
Permethrin	Insecticide	Rana catesbeiana	tadpole	7.033	16
Molinate	Herbicide	Bufo woodhousii	tadpole	14.0	2
Fenoprop	Herbicide	Limnodynastes peroni	tadpole	22.0	1
Paraquat	Herbicide	Bufo woodhousii	tadpole	26.0	2
Quinoline	Organics	Xenopus laevis	egg	79.0	18
Picloram	Herbicide	Adelotus brevis	tadpole	95.0	1
Sodium arsenate	Herbicide	Adelotus brevis	tadpole	96.0	1
Dicamba	Herbicide	Limnodynastes peroni	tadole	106.0	1
Matacil	Insecticide	Rana clamitans	tadpole	118.0	7
Aniline	Organics	Xenopus laevis	tadpole	150.0	18
2,4,5-T amine	Herbicide	Limnodynastes peroni	tadpole	169.0	1
2,4-D amine	Herbicide	Adelotus brevis	tadpole	200.0	1
DSMA	Herbicide	Limnodynastes peroni	tadpole	271.0	1
Pyridine	Organics	Xenopus laevis	tadpole	1090.0	18

References: 1=Johnson 1976, 2=Sanders 1970, 3=Marchal-Segault 1976, 4=Dumont and Schultz 1980, 5=Gopal et al. 1981, 6=Huey and Beitinger 1980, 7=Lyons et al. 1976, 8=Hall and Swineford 1980, 9=Wohlgemuth 1977, 10=Hall and Swineford 1981, 11=Mathews et al. 1976, 12=Landé and Guttman 1973, 13=Bills et al. 1977, 14=Browne and Dumont 1979, 15=Canton and Slooff 1982, 16=Jolly et al. 1978, 17=Marking et al. 1977, 18=Davis et al. 1981, 19=Birge et al. 1979.

b) TOXICITY TESTING TECHNIQUES

Several species of amphibians are readily available from the field or from commercial distributors. Adult anurans can be housed at low temperatures, without food, until they are to be used. They can be induced to spawn readily (Rugh 1962, Browne and Dumont 1979). Spawn can be collected from the field and developed under laboratory conditions. Development in lab aquaria can easily be observed and amphibians do not generally require a large amount of water or space. Some studies have shown that anurans require less attention during these processes than urodeles. Urodeles require live food and can be difficult to handle, making anurans more attractive as a test organism (Slooff and Baerselman 1980).

Amphibians have several other attributes that make them useful test organisms. A short life cycle which involves physiological, histological and anatomical changes (Cooke 1981), allow observations of the complete cycle over a short period of time (Birge et al. 1975). They respond to environmental contaminants in many different ways and these can be monitored over the life of the organism.

Some concerns that are evident after reviewing the amphibian toxicological literature about methods and techniques that should be evaluated prior to any toxicity testing are as follows:

1. Section of species. There is a certain degree of species variability in response to environmental pollutants (e.g. Sanders 1970, Cooke 1972a,b, Rzehak et al. 1977, Birge et al. 1979) (Table 1). Although it is generally recommended that the most sensitive species be used in any toxicity testing, so little is known about the sensitivity of most amphibians that it is difficult to choose a single indicator species. X. laevis and R. pipiens are often used in toxicity testing. X. laevis does not naturally occur in Canada and may not reflect the sensitivity of native species. Whenever possible a number of species should be tested and results from single species should not be generalized to cover all amphibians until species sensitivities have been more extensively investigated (Hall and Swineford 1980, 1981).

2. Lifestages sensitivity. There is a wide range in sensitivity of lifestages to the same toxicant (e.g. Cooke 1972a,b, Wohlgemuth 1977, Lyons et al. 1976, Hall and Swineford 1980, Davis et al. 1981). In general, adults are more tolerant than either the egg or larval stages. The egg and larval stages exhibit wide ranges in sensitivity (Hall and Swineford 1980, Davis et al. 1981). The sensitive stages vary greatly between toxicants presumably dependent upon the physiological processes that are occurring in the organisms during the developmental stage and the physiological

mechanism causing toxicity (Davis et al. 1981). One process that has not been considered, except with respect to pH, is toxic effects on fertilization and egg cleavage. Schlichter (1981) found fertilization of R. pipiens eggs to be highly sensitive to pH 5.8. Species may be experiencing high mortality during fertilization which is not measured in standard egg bioassays. In all bioassays as many lifestages as possible should be tested and in all instances, the exact stages when exposures were intiated and terminated should be reported.

3. Toxic effect. Mortality, as measured in bioassays, is not the only toxic effect that can be induced by a pollutant. Behavioural, morphological and developmental aberrations that occur during chronic dosing at levels much lower than the LC50, can also be important measures of toxicity (e.g. Cooke 1970, 1973a, 1981, Lyons et al. 1976, Rzehak 1977, Harri et al. 1979, Hall and Swineford 1980). Residues are an indirect toxic effect that should be considered, particularly for pollutants that have little effect on amphibians but that may accumulate and pass up the food web (Rosato and Ferguson 1968, Collins et al. 1973, Baudo 1976).

4. Dilution water. Although dilution waters should vary to represent conditions where the pollutant is a concern, the water characteristics should always be reported, particularly pH, alkalinity and oxygen content. Generalizations cannot be made about the effects in all water qualities. pH is an important parameter that can be toxic by itself (Gosner and Black 1957, Pough 1976) and also can influence toxicity (Hashimoto and Nishiuchi 1981). Dissolved organic carbon levels are also important because some toxicants, such as metals, can interact with organics and modify the toxicity (Baker and Schofield 1980). Hardness or alkalinity is closely linked with pH, but alone it can influence toxicity (Porter and Hakanson 1976). For example, metal toxicity is greater in soft water than in hard water at the same pH (Ibid 1976).

5. Temperature. Toxicity can also vary with temperature during exposures (Licht 1976b, Pough and Wilson 1977). Temperature should always be reported and experiments should be conducted at a temperature comparable to what would be encountered in field conditions.

6. Holding technique. With only a few exceptions, aquatic amphibians inhabit lentic waters, often small temporary pools or ponds with very little water flow. Flow-through experiments may therefore not represent an accurate field situation. To maintain toxicant levels and avoid build up of waste product a flow-through system is often preferred to static bioassays. Hall and Swineford (1981) found

flowing water, with a five hour replacement time, caused moderate mortality to R. septentrionalis tadpoles and in later experiments reduced the flow to a 24 hour replacent (Ibid 1981). In comparison with Sanders' (1970) methods Hall and Swineford (1980) found that continuous-flow methods yielded lower LC50 estimates than static methods. However, flow-through methods with a rapid renewal time do not duplicate conditions in the field. Further investigation is necessary to fully assess the implications of static and flow-through bioassays on different life stages.

7. Rearing density and body size. Rearing density can affect size of tadpoles with high densities (50 tadpoles/L) producing tadpoles half the size of those reared at low densities (10 tadpoles/L) (Cooke 1979). The significance of size is that large individuals can be more resistant to pollutants than small individuals. For example body size was negatively correlated with T. cristatus sensitivity to Maneb (a fungicide) (Zaffaroni et al. 1978) and copper toxicity to R. pipiens tadpoles (Landé and Guttman 1973). Bioassays should therefore include either a random sample of sizes or should be designed to incorporate any effects of body size.

8. Observation period. The observation period to observe lethality after exposure to different chemicals needs to be determined for different groups of pollutants. Hall and Swineford (1980) reported much lower LC50s for toxaphene based on a 30 day observation period than Sanders (1970) did based on a 4 day observation period. Hall and Swineford (1980) attributed their lower LC50 mainly to the longer observation period because most mortalities were observed after the 4 day dosing between 1 and 26 days after exposure.

There are no generally accepted standardized techniques for toxicity testing using amphibians. The wide range of bioassay methodologies used in measuring LC50s makes comparison between studies very difficult. The United States EPA (1975) has published a detailed methodology which is recommended for use in toxicity testing of amphibians. However, it is designed mainly for fish, and to a lesser extent, aquatic invertebrate testing, and it does not fully address amphibian toxicity testing.

c) AMPHIBIANS AS MONITORS OF ENVIRONMENTAL POLLUTANTS

Ideally indicator species should be both representative of a specific trophic level within the ecosystem and capable of being used, at reasonable cost, under laboratory conditions. The latter requirement has already been considered under section XI b).

The requirements for indicator species were listed by Moore (1966) as follows:

(a) The species should be widely distributed, relatively abundant and easy to collect.

(b) If monitoring is to be carried out by chemical analysis of organs, these should be large enough for adequate samples.

(c) It should be possible to ascertain the age of the individual.

(d) The level of residues in the species should be between the limit of detection and the limit of crude toxicological effect.

(e) If measurements of local changes are required, the species must be sedentary. If the degree of contamination of a large area is to be measured, species with more extensive ranges can be used. In either case the range of the indicator species must be known.

Dumpert and Zietz (1984) considered that the platanna (X. laevis) could be used as an indicator species for determining embryotoxic effects of environmental chemicals. It met their criteria that it was available in sufficient numbers and at an acceptable cost at any time of the year and was adequately sensitive.

Cooke (1972a) has stated that under normal usage patterns that most insecticides would probably not be likely to reach levels high enough to induce toxic effects in the field. Meeks (1968) and Niethammer et al. (1984, 1985) believed that adult anurans would not be useful as indicators of organochlorine compounds as they do not accumulate residues in proportion with environmental levels.

The toxicity and toxic effects of organophosphate insecticides to amphibians varies greatly (Table 1) and due to the low persistence of these compounds in the environment, results are difficult to compare. Pearce and Price (1977) reported that spraying operations using fenitrothion represented little hazard to amphibians when used according to recommended field application rates. In contrast, Johnson and Prine (1976) reported that fenthion even when used at half the usual concentration caused marked physiological changes. These results, the apparent tolerance to cholinesterase inhibitors by amphibians (Potter and O'Brien 1964) and the generally low toxicity of organophosphates (Edery and Schatzberg-Porath 1960), indicates amphibians would not be adversely affected by many organophosphate compounds in the environment.

Arias and Zavanella (1979) presented data indicating the feasibility of using forelimb regeneration in newts as a model for assessing the risks incurred by the use of pesticides. However, the applications of this method may yield results difficult to apply to environmental situations.

Birge et al. (1973, 1975, 1976, 1977, 1979) have studied the effects of metal contamination on amphibians and the possibility of using amphibians as bioassay and bioindicator species based on the sensitivity of some amphibian species. Birge et al. (1975) considered that amphibians are an ideal test organism. The authors noted that due to differences in species sensitivity, that conclusions should be based on results from two or three test species. Birge et al. (1979) noted significant differences between urodele and anuran sensitivity based on individual ecological requirements, and concluded that sensitivity to environmental stress may be related to individual life cycles and requirements.

The sensitivity of amphibians to metallic contaminants may allow environmental monitoring of these compounds using amphibians as indicators, once standardized techniques and representative species have been established (Birge et al. 1975, 1979, Niethammer et al. 1985).

Amphibians can be used to monitor the effects of acidification. The survival of embryos has been related to pH, under both experimental and field conditions (see Section X b)). Fish are widely used monitor species for the impact of acidification. Amphibians could be used to broaden the approach and might be particularly valuable for assessing the impact on temporary water bodies.

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TABLE 1
ACUTE TOXICITY STUDIES

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	72 hr	96 hr	Effects[d]	Remarks	References
Acephate (OP)	- *Pana clamitans*	tadpoles	A	housed at 21 ± 1	-	-	6633 (5857-6775)	-	-	-		95% fiducial limits in parentheses.	Lyons et al., 1976
	- *Asellus aquaticus*	69 day old larvae	renewed daily	22 (7.0)	-	6 mg/L hardness as $CaCO_3$	-	-	-	8816			Cowan et al., 1984
Acetone	- *Asellus racicavus*	3-4 weeks post hatch	A	20 ± 1	-	-	-	30,000	-	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	24,000	-	-			
Acridine	- *Xenopus laevis*	mid-blastula embryos	A	Room temp.	-	98.5 mg/L	-	10.9 (3.8-39.3)	8.7 (4.1-18.6)	3.6 (1.9-6.7)	Teratogenicity EC50 for mid-blastula embryos: 48 hr-6.5 (3.1-13.7); 72 hr-2.6 (0.7-10.2); 96 hr-2.6 (0.6-7.0). Abnormalities include exogastrulation, edema, formation of blisters.	95% confidence limits in brackets.	Davis et al., 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	72 hr	96 hr	Effects[d]	Remarks	References
Acridine (cont'd)	- *Xenopus laevis*	tailbud embryos	A	Room temp.	-	98.5 mg/L	13.2 (8.5-20.6)	7.1 (5.3-9.5)	5.5 (4.6-6.5)	-		95% confidence limits in brackets.	Davis et al., 1981 (cont'd)
		free swimming larvae	"	"	"	"	6.2 (5.5-7.0)	5.6 (4.7-6.3)	4.9 (4.1-5.9)	4.5 (3.7-5.5)			
Aeration	- *Xenopus laevis*	tadpoles	B	17.2 ± 1 (7.39 ± .32)	-	40.6-49.5 mg/L as $CaCO_3$	-	-	-	0.887 (.866-.908)		95% confidence interval in parentheses.	Maicumbo et al., 1987
							24 hr.	48 hr.		96 hr.			
Aeration-50 (propellant)	- *Asellus communis*, *A. aquaticus*	larvae	A	20.0-24.0 (7.8-8.2)	APHA et al., 1971	105-232 mg/L as $CaCO_3$	>10	6.7		3.5			Slooff, 1984
			"	"	"	16-18 mg/L as $CaCO_3$	>10	>10		5.2			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Aldrin (OC)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	2.0 (1.6-3.1)	0.68 (0.17-2.7)	0.15 (0.03-0.78)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
Allylamine	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	1.8	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	-	-	-	-	-	5.0	-			
								7 day				
Aluminum (N)	- Crassostrea virginnica	embryo-larval	B	- (7.0-7.8)	Birge & Black 1977a,b	195.0 ± 9.4 ppm hardness as $CaCO_3$		0.05		Exposure from fertilization to 4 days posthatch.	N = 14.	Birge et al., 1979a

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Anamycin (C)	- Rana clamitans	tadpoles: small 8.2 ± 1.1 cm mean body length; large 10.9 ± .1 cm body length	A	housed at 21 ± 1	-	-	267 (232-342) 234 (223-246	206 (191-220)	118 (112-125)		95% fiducial limits in parentheses.	Lyons et al., 1976
2-Amino-ethanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	220	-		N_{est} 10 per concentration.	de Zwart and Slooff, 1987
1-Amino-2-propanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	420	-		N = 10 per concentration.	de Zwart and Slooff, 1987
4-Aminopyridine (avian frightening agent)	- Rana catesbeiana	larvae	A	24	ASTM Committee E-35 on Pesticides 1980	hardness 24 mg/L as $CaCO^{3}$	7.2 (6.6-7.8)	-	2.4 (2.0-2.9)		95% confidence intervals in brackets.	Working and Champion, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	72 hr	96 hr	Effects[d]	Remarks	Reference
Acetrole-I (H)	- Asellus brevis	1-2 week old tadpoles	A	21-22	Sanders 1970	-	5.2 g/L	5.0 g/L	-	3.0 g/L			Johnson, 197-
Aniline	- Ambystoma maculatum	3-4 weeks post hatch	A	20 ± 1	-	-	-	440	-	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	140	-	-			
	- Xenopus laevis	mid-blastula embryos	A	Room Temp.	-	98.5 mg/L	1400 (325-6100)	660 (470-940)	490 (330-730)	550 (350-850)	Teratogenicity EC50 for mid-blastula embryos: 24 hr-360 (180-740); 48 hr-560 (360-910); 72 hr-460 (280-750); 96 hr-370 (150-900). Abnormalities included concentration, edema, formation of blisters.	95% confidence limits in brackets.	Davis et al., 1981
		tailbud embryos	"	"	"	"	1420 (1100-2300)	1370	1150	940			
		free swimming larvae	"	"	"	"	-	1420[e]	540 (350-830)	150 (105-200)			
							30 min	5 hr		7 day			
Anthracene	- Rana pipiens	embryos - stage 25	A	-	-	-	0.063	0.025		-		LC50 determined for given length of exposure to sunlight.	Kagan et al., 1984

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 30 min	5 hr	7 day	Effects[d]	Remarks	Reference
Antimony (H)	- Centrophrys carolinensis	embryo-larval	B	- (7.0-7.8)	Birge & Black 1977a,b	195.0 ± 5.4 ppm hardness as $CaCO_3$	-	-	0.30	Exposure from fertilization to 4 days post hatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Arsanilic Acid (C)	- Bufo bufo japonicus	tadpole	-	-	-	-	-	75	-			Hashimoto and Nishiuchi, 1981
								7 day				
Arsenic (H)	- Centrophrys carolinensis	embryo-larval	B	- (7.0-7.8)	Birge & Black 1977a,b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.08		Exposure from fertilization to 4 days post hatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Azinphos-methyl (GP)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	19.3 ± .3 (7.1)	-	30 ppm methyl orange	0.68 (0.45-1.6)	0.31 (0.17-0.56)	0.13 (0.05-0.33)		N = 10. 95% confidence limits in brackets.	Sanders, 1970

Table 1 - Amphibian Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Bacillus sphaericus 1593M	- Rana bufo	tadpoles 5.2 mg 16 mg	A	- (7.2)	American Public Health Assoc. 1971	-	220 470	- -	- -		N = 10 per concentration. LC50 given in concentration of protein (ug/ml).	Mathavan and Yelpandi, 1984
Bacillus sphaericus 2362	- Rana bufo	tadpoles 5.2 mg 16 mg	A	- (7.2)	American Public Health Assoc. 1971	-	125 300	- -	- -		N = 10 per concentration. LC50 given in concentration of protein (ug/ml).	Mathavan and Yelpandi, 1984
Benthiocarb (H)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	3.5	-			Hashimoto and Nishiuchi, 1981
Benzene	- Ambystoma opacicrepum	3-4 weeks post hatch	A	20 ± 1	-	-	-	370	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	190	-			
	Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	190	-		N = 10 per concentration.	de Zwart and Slooff, 1982

Table 1 - Amphibian Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Benzene Hexachloride	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	19.5 ± .5 (7.1)	-	30 ppm methyl orange	13 (6.3-40)	7.1 (4.8-10)	3.2 (0.30-6.3)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
Beryllium Sulfate	- Ambystoma opacum	larvae	A	23.5 ± 1.5	APHA 1965	hard = 400 mg/L; soft = 20 - 25 mg/L	31.5 23.7	31.5 4.21	31.5 3.15	In ppm Be	N = 120.	Slonim and May, 1975
	Ambystoma maculatum	larvae	"	"	"	hard	18.2 21.2	18.2 18.2	18.2 18.2		N = 80.	
						soft	>10 >12	>10 >7	8.81 5.65			
g-BHC	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	24	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
5-Benzylthrin (PY)	- Rana pipiens	adults 25-30 g	-	20	Subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50 = 1.7 (1.3-2.4)	N = 25-30. 95% confidence interval in parentheses.	Cole and Casida, 1983

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
3-Bromopropene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	0.66	-		N = 10 per concentration.	de Zwart and Slooff, 1987
1-Butanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	1200	-		N = 10 per concentration.	de Zwart and Slooff, 1987
2-Butanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	1530	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Cadmium (II)	- Rufo melanostictus	tadpoles 1.8-2.2 cm	A	29-34 (7.1-7.6)	APHA et al. 1976	120-160 mg/L as $CaCO_3$	19.81 (17.39-29.15)	11.91 (9.99-13.95)	8.18 (6.96-9.53)		95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Ray, 1987
	- Microhyla ornata	tadpoles	re-newed daily	25.5-26.0 (6.86-6.96)	-	97.6-98.0 ppm					Ranges in parentheses. Animals not fed during test.	Rao and Madhyastha, 1987
		1 week old					2.62 (2.4-2.8)	2.48 (2.0-2.8)	1.58 (1.4-1.8)			
		4 weeks old					2.78 (2.4-3.0)	2.66 (2.2-2.8)	1.81 (1.4-2.0)			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Cadmium (cont'd)	- Rufo regularis	adult females 25 ± 4.3 g	-	-	Single i.m. injection $CdCl_2$	-	-	-	-	24 hr - 22 48 hr - 18 96 hr - 6.2		Hiley et al., 1986a
								7 day				
	- Centropyxus cornuta	embryo-larval	S	- (7.8-7.8)	Birge & Black 1977a,b	195.0 ± 9.4 ppm hardness as $CaCO_3$		0.04		Exposure from fertilization to 4 days post hatch	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
	- Xenopus laevis	3-4 weeks	C	20 ± 1	Dutch Standardization Organization 1980	1.7 mval/L hardness	4 (16)	3.2 (12.8)	-	No Toxic Effect Level for 24 or 48 hr -2.2.	N = 10. Values are corrected (based on actual test concentrations); uncorrected values are in parentheses.	Canton and Slooff, 1982a
Cadmium Nitrate	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	1.3	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	32	-			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]				Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	72 hr	96 hr			
Cadmium Nitrate (cont'd)	- Xenopus laevis	3-4 weeks old larvae	A	20 ± 1	-	-	-	20.2	-	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Captan (F)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	3.0	-	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
Carbaryl (C)	- Rana tigrina	tadpoles weight: 0.1 g 0.82 g 1.2 g	A	-	-	-	12.8 - -	8.25 - -	- - -	6.3 6.2 11.7	Median Resistance Time for 0.5 g tadpole: 1230 min at 10 ppm 110 min at 30 ppm.	N = 10 per group. LC_{50} is weight dependent factor.	Marian et al., 1983
	- Bufo bufo	26-37 day old tadpoles	A	20 ± 1	-	-	20.5-21.8	18.2-20.8	16.8-20.4	-	Development slowed.	95% confidence intervals.	Marchal-Segault, 1976
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	7.2	-	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
							24 hr	48 hr		96 hr			
Carbophenthion (DP)	- Pseudacris triseriata	1 week old tadpoles	A	15.5 ± 5 (7.1)	-	30 ppm methyl orange	0.10 (0.039-0.20)	0.05 (0.010-0.25)		0.028 (0.004-0.09)		N = 10. 95% confidence limits in brackets.	Sanders, 1970

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Carbomethyl-succinate	- Xenopus laevis	3-4 weeks	semi-static	21 ± 2	-	-	-	-	1,000-3,200		Test solutions corrected for oxygen content.	Canton and Slooff, 1982b
Carton	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	5.6	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
1-Chloro-3-bromopropane	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	41	-		N = 10 per concentration.	de Zwart and Slooff, 1987
3-Chloro-2-methylpropene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	10	-		N = 10 per concentration.	de Zwart and Slooff, 1987
4-Chloro-2-picolinic Acid (H)	- Pseudacris triseriata	1 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	18 (9.0-27)	12 (6.0-24)	6.0 (2.0-18.0)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
3-Chloro-1,2-propanediol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	181	-		N = 10 per concentration.	de Zwart and Slooff, 1987
3-Chloro-1-propanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	2750	-		N = 10 per concentration.	de Zwart and Slooff, 1987

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
1-Chloropropene	- Tanytarsus luevis	3-4 week old larvae	A	20 ± 1	-	-	-	0.34	-		N = 10 per concentration.	de Zwart and Slooff, 1987
								7 day				
Chromium (VI)	- Cyprinodon variegatus	embryo-larval	S	- (7.8-7.8)	Birge & Black 1977a,b	199.8 ± 5.4 ppm hardness as $CaCO_3$		0.05		Exposure from fertilization to 4 days post hatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
	- Salmo gairdneri	Yearling 1.8-2.2 cm	A	29-34 (7.1-7.4)	APHA et al. 1976	125-140 mg/L as $CaCO_3$	57.97 (49.14-65.48)	53.43 (49.21-61.67)	49.29 (43.63-56.39)	In high concentrations surfacing, erratic body movement, loss of equilibrium noted.	95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Ray, 1987
Citrex-3-3	- Tanytarsus luevis	3-4 weeks	Semi static	21 ± 2	-	-	-	-	10,000 10,000		Test solutions corrected for pH and oxygen content.	Canton and Slooff, 1982b
								7 day				
Cobalt (II)	- Cyprinodon variegatus	embryo-larval	S	- (7.8-7.8)	Birge & Black 1977a,b	199.8 ± 5.4 ppm hardness as $CaCO_3$		0.05		Exposure from fertilization to 4 days post hatch.	N = 14.	Birge et al., 1979a

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Cobalt (^{60}Co)	- Limnodrilus hoffmeisteri		-	25	Cobalt-60 gamma radiation from a Theratron unit at dose rate of 0.3 Gy/min.	-	-	-	-	LD50 values (Gy):	N = 100 - 120	Panter, 1986
		Fertilized egg (.75 hr)								0.6	40 day observation.	
		late cleavage (10 hr)								3.3	40 day observation.	
		tail bud (33 hr)								9.9	40 day observation.	
		heartbeat (3 days)								18.6	40 day observation.	
		early limb bud (9-10 days)								20.2	40 day observation.	
		Sex development (40-50 days)								26.9	40 day observation.	

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Cobalt (^{60}Co) (cont'd)		metamorphosis (60 days)	-	25	0.3 Gy/min	-	-	-	-	18.3	160 day observation.	Panter, 1986 (cont'd)
		young frog (80 days)								18.7	160 day observation.	
Copper (II)	- Bufo melanostictus	tadpoles 1.8-2.2 cm	A	29-34 (7.1-7.6)	APHA et al. 1976	120-160 mg/L as $CaCO_3$	.843 (.731-.936)	.446 (.36-.55)	.32		95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Ray, 1987
	- Microhyla ornata	tadpoles	re-newed daily	25.5-26.0 (6.86-6.94)	-	97.0 98.0 ppm					Range in brackets. Animals not fed during tests.	Rao and Madhavendra, 1987
		1 week old					5.61 (5.4-5.8)	5.31 (5.0-5.6)	5.[illegible] (4.7-5.1)			
		4 weeks old					6.04 (5.6-6.4)	5.74 (5.4-6.2)	5.[illegible] (5.0-[illegible].8)			

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Copper (cont'd)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977a,b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.06		Exposure from fertilization to 4 days post hatch.	N = 16.	Birge et al., 1979a
								72 hr				
Copper Sulfate	- Rana pipiens	larval-immediately post hatch	A	19.4 ± 1.2 (7.73 ± 0.5)	-	290 ± 32 mg/L as $CaCO_3$		0.15			N = 15 per concentration. Survival time was positively correlated with weight of tadpole.	Landé and Guttman, 1971
								30 day				
	- Rana pipiens	adults	A	20	-	-		.00168				Kaplan and Yeh, 1961
							24 hr	48 hr	96 hr			
	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	1.7	-		N = 10 per concentration.	de Zwart and Slooff., 1987

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
o-Cresol	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	40	-		N = 10 for each species. Organisms not fed during expt.	Sloff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	38	-			
Cyanatryn (H)	- Rana temporaria	tadpoles	No test methodology specified				-	-	38			Maddox et al., 1974
1,5,9 - Cyclododeca-decatriene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	1.6	-		N = 10 per concentration.	de Zwart and Sloff, 1987
1,3,5 - Cycloheptatriene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	41	-		N = 10 per concentration.	de Zwart and Sloff, 1987
1,5 - Cyclo-octadiene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	24	-		N = 10 per concentration.	de Zwart and Sloff, 1987

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
1R, cS - Cypermethrin (FY)	- Rana catesbeiana	tadpoles 10-15 g	-	20	intraperi-toneal injection	-	-	-	-	24 hr LD50: trans 0.20 (0.06-0.41) cis 0.04 (0.01-0.24)	N = 32-45. 95% confidence limits in parentheses.	Cole and Casida, 1981
	R. pipiens	adults 20-30 g	-	20	subcuta-neous injection into the dorsum	-	-	-	-	24 hr LD50: trans 0.45 (0.41-1.0) cis 0.16 (0.12-0.24)	N = 28-34. 95% confidence limits in parentheses.	
2,4-D (H) (2,4-dichloro-phenoxyacetic acid)	- Bufo melanostictus		A	Approx. 25 (8.3)	APHA 1980	220 mg/L total hardness	13.77 (11.81-16.05)	9.09 (8.23-9.91)	8.05 (7.29-8.81)	Incipient Lethal Level - 6.1.	95% fiducial limits in parentheses.	Verdia et al., 1984
2,4-D amine	- Adelotus brevis	1-2 week old tadpoles	A	21-22	Sanders 1970	-	255	228	200			Johnson, 1976
	Limnodynastes peronii	1-2 week old tadpoles	"	"	"	"	321	300	287			
	Bufo marinus	2 week old tadpoles	"	"	"	"	346	333	286			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
2,4-D amine (cont'd)	- Pseudacris triseriata	1 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	100	-	100		N = 10.	Sanders, 1970
2,4-D iso-octyl ester (Agroxone S)	- Irituru cristatus carnifex	adults	re-newed daily	-	-	-	-	-	-	LT50 values (lethal time for 50% of individuals) hrs conc / male / female 150 / 14 (6.8-28.7) / not determined 125 / not determined / 52 (38.3-78.9) 75 / 102 (89.1-122.3) / 132 (105.6-165.0) 50 / 440 (321.9-601.5) / -	Animals immersed in Agroxone S (37% 2,4-D; 63% diluents and emulsifiers); concentrations for LT50's are 2,4-D concentrations. N = 10 per concentration 95% confidence limits in parentheses.	Zaffaroni et al., 1986a

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]				Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	72 hr	96 hr			
2,4 - D sodium	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>40	-	-			Hashimoto and Nishiuchi, 1981
DDT (DC)	- Rana temporaria	adults	-	10-15	Oral administration of DDT dissolved in olive oil.	-	-	-	-	-	96 hr LD50 (estimated from graph) - 24; 20 day LD50 - 7.6.	Toxicity was increased by starving.	Harri et al., 1979
	- Bufo bufo	26-37 day old tadpoles	A	20 ± 1	-	-	0.7-1.3	0.5-0.8	0.3-0.5	-	95% confidence intervals.	Development is slowed. DDT more toxic than Savin or Fenthion.	Marchal-Segault, 1976
	- Pseudacris triseriata	one week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	1.4 (0.91-2.8)	0.90 (0.40-1.5)	-	0.80 (0.50-2.3)		N = 10. 95% confidence limits in brackets.	Sanders, 1970

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
DDT (cont'd)	Bufo woodhousii fowleri	tadpoles: 1 week old weight, $\bar{x}$ = 15 mg	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	5.3 (2.9-9.9)	1.8 (0.99-3.3)	0.75 (0.28-2.0)		N = 10.	Sanders, 1970 (cont'd)
		2-3 weeks old weight, $\bar{x}$ = 54 mg					5.4 (2.9-10)	1.3 (0.32-5.3)	-			
		4-5 weeks old weight, $\bar{x}$ = 74 mg					2.4 (0.75-8.0)	1.0 (0.58-6.5)	1.0 (0.02-3.6)			
		6 weeks old weight, $\bar{x}$ = 350 mg					2.2 (0.58-15)	0.61 (0.28-0.6)	0.18 (0.03-0.6)			
		7 weeks old weight, $\bar{x}$ = 600 mg					1.6 (0.98-2.8)	0.75 (0.41-1.1)	0.03 (0.004-0.4)			
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	31			Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
DDT (H)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	1.2 (0.98-2.6)	0.76 (0.46-0.82)	0.42 (0.16-1.1)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
cis - Deltamethrin (PY)	- Rana pipiens pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: 0.35 (0.19-0.62)	N = 20-34. 95% confidence limits in parentheses.	Cole and Casida, 1981
	R. catesbeiana	tadpoles 10-15 g	-	20	intraperitoneal injection	-	-	-	-	24 hr LD50: 0.13 (0.05-0.34)	N = 32-45. 95% confidence limits in parentheses.	
DFP (OP) (diisopropyl fluorophosphate)	- Bufo viridis	adults 18-43 g	-	23	injection into dorsal lymphatic sac	-	-	-	-	7 day LD50: 1450.	N = 25 minimum.	Edery and Scheiberg-Porath, 1960
Diazinon (OP)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	14	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Dicamba (H)	- Adelotus brevis	1 to 2 week old tadpoles	A	21-22	Sanders 1970	-	220	202	185			Johnson, 1976
	Limnodynastes peronii	1 to 2 week old tadpoles	"	"	"	"	205	166	106			
2,4-Dichloroaniline	- Xenopus laevis	<2 days	semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed.): Mortality - 1 Development - 0.32 Growth - 1		Slooff and Canton, 1983
1,3-Dichloropropene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	63	-		N = 10 per concentration.	de Zwart and Slooff, 1987
1,3-Dichloropropene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	450	-		N = 10 per concentration.	de Zwart and Slooff, 1987

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Dieldrin (OC)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.5 ± .3 (7.1)	-	30 ppm methyl orange	1.1 (1.4-2.2)	0.60 (0.09-1.2)	0.15 (0.02-0.47)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	one week old tadpoles	"	"	"	"	0.23 (0.08) 0.41	0.22 (0.080-0.40)	0.10 (0.03-0.28)			
Disefox (OP)	- Bufo viridis	adults 18-45 g	-	25	injection into dorsal lymphatic sac	-	-	-	-	7 day LD50-1,410.	N = 25 minimum.	Ebory and Schatzberg-Porath, 1960
Dimethoate (OP)	- Xenopus laevis	<2 days	semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed): Mortality - 1 Development - 32 Growth - 32		Slooff and Canton, 1983

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Dimethoate (cont'd)	- Rana temporaria	adults	renewed daily	23 ± .2 (7.3-7.8)	-	60-70 mg/l hardness				Significant glycogen elevation in liver, muscle and kidney noted.	95% confidence limits in parentheses.	Mudgal and Patil, 1987
		male					51.4 (49.6-53.5)	43.3 (41.6-45.1)	39.0 (37.5-40.5)			
		female					-	44.3 (43.4-47.1)	36.0 (35.8-36.2)			
1,2-Dimethylbenzene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	73	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Dimethylhydrazine (unsymetrical, UDMH) (propellant)	- Ambystoma mexicanum, A. opacum	larvae	A	20.5-23.5 (7.8-8.2)	APHA et al. 1971	185-232 mg/l as $CaCO_3$	>100	93	26			Slonim, 1986
		"	"	20.5-23.5 (6.3-6.9)	"	16-18 mg/l as $CaCO_3$	>135	>135	100			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
1,1-Dimethylhydrazine	- Xenopus laevis	embryos	A	-	-	-	-	-	-	EC50 (teratogenesis) for exposure from blastula to hatching: 7.9.		Greenhouse, 1977
2,6-Dimethylquinoline	- Xenopus laevis	early cleavage to mid-blastula embryos	A	20	-	-	-	-	6.5 (5.6-7.5)		N = 25. 95% confidence limits in brackets. Reduced motility, development retarded, pigmentation lighter.	Dumont et al., 1979
Dinitro-o-cresol	- Xenopus laevis	<2 days	Semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed): Mortality - 0.32 Development - 0.32 Growth - 0.32	N = 75.	Slooff and Canton, 1983
Dinocap (M)	- Rana rana temporaria	tadpoles	-	-	-	-	-	0.55	-			Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
3,6-Dioxa-1,8-octanediol	- Xenopus laevis	3-6 week old larvae	A	20 ± 1	-	-	-	7047	-		N = 10 per concentration.	de Zwart and Slooff, 1987
2,2-DPA (H)	- Adelotus brevis	1-2 week old tadpoles	A	21-22	Sanders 1970	-	11.1 g/L	9.2 g/L	6.2 g/L			Johnson, 1976
	Limnodynastes peronii	1-2 week old tadpoles	"	"	"	"	3.3 g/L	2.5 g/L	2.0 g/L			
DMC-1339 (avian control chemical)	- Rana sphenocephala	larvae	A	16	ASIN Committee E-35 on Pesticides 1980	Hardness 24 mg/L as $CaCO_3$	63 (60-66)	-	44 (42-47)		95% confidence intervals in brackets.	Marking and Chandler, 1981
DMC-1367 (avian control chemical)	- Rana sphenocephala	larvae	A	16	ASIN Committee E-35 on Pesticides 1980	Hardness 24 mg/L as $CaCO_3$	41 (40-42)	-	32 (30-34)		95% confidence intervals in brackets.	Marking and Chandler, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
DMC-2698 (avian control chemical)	- Rana sphenocephala	larvae	A	16	ASIN Committee E-35 on Pesticides 1980	Hardness 24 mg/L as $CaCO_3$	-	-	>10			Marking and Chandler, 1981
DSMA (H) (disodium methyl arsonate)	- Adelotus brevis	1-2 week old tadpoles	A	21-22	-	-	600	525	453			Johnson, 1976
	Limnodynastes peronii	1-2 week old tadpoles	"	"	"	"	324	310	271			
Endosulfan (EC)	- Rana tigrina	tadpoles	A	28 ± 2 (6.9-7.2)	-	95-100 mg/L	0.0021 (0.0018-0.0025)	0.0020 (0.0017-0.0023)	0.0018 (0.0014-0.0022)	96 hr presumable Noncipus conc.-.00399.	95% confidence limits in brackets. Frogs more susceptible than fish and aquatic insect.	Gopal et al, 1981
	- Bufo melanostictus		A	Approx. 25 (8.3)	APHA 1980	220 mg/L total hardness	0.1619 (.1639-.1213)	0.1368 (.1370 .1318)	0.1230 (.1270-.1184)	Incipient Lethal Level - 0.105.	95% fiducial limits in parentheses.	Pardio et al., 1984
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	9.0	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Endothall (N) (mono(n,n-dimethylalkylamine) salt)	- Bufo woodhousii	4-5 week old tadpoles	A	19.9 ± .5 (7.1)	-	38 ppm methyl orange	3.2 (1.7-5.5)	1.8 (0.93-3.2)	1.2 (0.46-3.4)		N = 18. 95% confidence limits in brackets.	Sanders, 1970
Endrin (EC)	- Rana sphenocephala	eggs	B	20 (7.2-7.5)	-	total hardness 100 ppm as $CaCO_3$	0.025	-	-	Exposed for 24 hr, observed at 96 hr.	N = 20-30 per level.	Hall and Swineford, 1980
		sub-adults	"	"	"	"	-	-	0.085		N = 5-10 per level.	
		larvae	"	"	"	"	-	-	0.036	Exposed for 96 hr, observed at 192 hr.	N = 20-30 per level.	
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	12	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
			B	20 (7.2-7.5)	Hall and Swineford 1980	hardness 100 ppm as $CaCO_3$				24 hr EC50 (conc. producing behavioural aberrations in 50% of animals): 0.013 (0.009-0.020)	Animals were exposed for 96 hr, mortality was determined after 192 hr. 95% confidence limits in brackets.	Hall and Swineford, 1981
	Rana sphenocephala	larvae	"	"	"		-	-	0.009 (0.006-0.014)			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Endrin (cont'd)	N. cyanoleuca	larvae	"	"	"	"	-	-	0.002 (0.001-0.005)	>0.040	Some differences between results using static or flow through test methods noted.	
	N. sitvatica	larvae	"	"	"	"	-	-	0.014 (0.023-0.055)	<0.014	Removal of pesticides from water by test animals was significant.	
	Bufo americanus	larvae	"	"	"	"	-	-	0.010 (0.007-0.014)	0.008 (0.006-0.010)		
	Acris crepitans	larvae	"	"	"	"	-	-	0.010 (0.008-0.011)	0.023 (0.016-0.031)		
	Ambystoma maculatum	larvae	"	"	"	"	-	-	0.018 (0.0-0.031)	0.018 (0.014-0.021)		
	Ambystoma opacolatum	larvae	"	"	"	"	-	-	0.014 (0.009-0.009)	0.048 (0.043-0.053)		

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]				Effects[d]	Remarks	Reference
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	72 hr	96 hr			
Endrin (cont'd)	- Bufo woodhousii	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	0.57 (0.33-1.3)	0.46 (0.31-0.68)	-	0.12 (0.30-0.66)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	-	-	-	-	0.29 (0.18-0.45)	0.29 (0.18-0.45)	-	0.18 (0.09-0.30)			
	- Rana catesbeiana	tadpoles 2-5 g	B	17.4-21.2 (8.08-8.06)	APHA 1980; USEPA 1976				0.0025 (.0021-.0029)			Range in parentheses.	Thurston et al., 1985

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	Reference
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	72 hr			
Endrin (cont'd)	- Rana temporaria	tadpoles: with external gills (2 days)	A	20 (6.0)	-	2.21-2.39 ml 1N-HCl/L	2.0086 (1.2106-3.1967)	0.7098 (0.3982-1.2535)	0.4253 (0.2999-0.6061)		Tadpoles lay paralysed on the bottom of aquaria. Fully mobile. Initial period of agitation, thereafter a loss of balance and paralysis, lying on bottom of aquaria.	Mehlquanth, 1977
		with internal gills, no limbs (8 days)					0.9886 (0.5890-1.7181)	0.2268 (0.1344-0.3932)	0.505 (0.0207-0.0903)			
		with internal gills, no limbs (15 days)					1.1375 (0.7217-1.9508)	0.6149 (0.3433-1.1796)	0.2875 (0.1537-0.4482)			
		with internal gills, (45 days)					0.2417 (0.1163-0.4691)	0.0748 (0.0308-0.1488)	0.0243 (0.0151-0.0351)			
		with pelvic limbs (86 days)					0.1685 (0.0793-0.3187)	0.0396 (0.0182-0.0776)	0.0172 (0.007-0.0257)			
		with pelvic and thoracic limbs (107 days)					0.1133 (0.0648-0.2256)	0.0299 (0.0158-0.0533)	0.0147 (0.0067-0.0210)			

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Eptam (H)	- Unspecified	Tadpoles 1-1.5cm	Re-newed daily	17-20 (7.7-8.2)	-	-	-	16.8	-	Value is LC_{50} of the active ingredient.		Paravaichenko, 1975
1,2-Ethanediol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	326	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Ethyl Acetate	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	150	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	180	-			
								10 day				
Ethylene Diamine	- Xenopus laevis	Tadpoles 5-12 days old	A	22 ± 1	-	-		250		Treatment was continuous for 10 days; LC50 was determined at 10 days post exposure.		Birch and Pramlal, 1986

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		10 day				
Ethylene Thiourea	- Xenopus laevis	Tadpoles 5-12 days old	A	22 ± 1	-	-		180		Treatment was continuous for 10 days; LC50 was determined at 10 days post exposure.		Birch and Pramlal, 1986
							24 hr	48 hr	96 hr			
Ethyl Propionate	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	56	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	56	-			
Fenitrothion (OP)	- Rana cimitora	Tadpoles 22.4 mm	A	Housed at 21 ± 1	-	-	9.9 (8.9-9)	7.8 (7.1-8.5)	6.9 (6.3-5.3)		95% fiducial limits in brackets.	Lyons et al., 1976
	- Bufo bufo japonicus	Tadpoles	-	-	-	-	-	9.8	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Fenoprop (H)	- Adelotus brevis	1-2 week old tadpoles	A	21-22	Sanders 1978	-	77	60	54			Johnson, 1976
	Limnodynastes peroni	1-2 week old tadpoles	"	"	"	"	35	27	22			
	Bufo marinus	2 week old tadpoles	"	"	"	"	60	42	34			
S - Fenpropathrin (PY)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: 0.27 (0.12-0.63)	N : 28-34. 95% confidence limits in parentheses.	Cole and Casida, 1983
Fenthion (OP)	- Bufo bufo	26-37 day old tadpoles	A	20 ± 1	-	-	2.1-2.6	2.0-2.2	1.8-2.2	95% confidence intervals.	Development slowed.	Marchal-Ségault, 1976
	- Rana pipiens Bufo boreas	1 day old larvae	-	-	-	-	-	-	5.8			Lewis et al., 1985

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Fenthion-acetate (F)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	0.33	-			Hashimoto and Nishiuchi, 1981
SS - Fenvalerate (PY)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: 0.13 (0.09-0.19)	N : 28-34. 95% confidence limits in parentheses.	Cole and Casida, 1983
Fenthion (F)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	4.2	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
Fluvalinate (valinate)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>40	-			Hashimoto and Nishiuchi, 1981
Fluorothene	- Rana pipiens	embryos stage 25	A	-	-	-		1 hr				
								0.09			LC50 determined for 1 hour of exposure to sunlight.	Kagan et al., 1985

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Fossil fuels:	- Xenopus laevis	mid-late blastula embryos	A	-	-	-				EC50 (conc. that induced teratogenesis in 50% of survivors):		Dumont et al., 1983
Coal-derived fuel oil blend							-	-	1.48%	0.96%		
Shale-derived crude							-	-	6.97%	3.36%		
Coal-gasifier electrostatic-precipitator tar							-	-	0.83%	0.48%		
Aromatic petroleum crude							-	-	33.38%	31.10%		
										Little effect on growth rate; pigmentation and motility reduced by some of the materials.		

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Fossil fuels (Coal-gasification Electrostatic Precipitator Tar Fractions):	- Xenopus laevis		A	Room temp.	-	-				EC50 (conc. which causes malformations in 50% of survivors); EC0 (no effect conc.):	95% confidence intervals in parentheses.	Schultz et al., 1983
Raw Tar		blastula embryos					-	-	3.13- (2.32- 4.23)	96 hr EC50=0.70 (.31-1.57); EC0=0.50.		
		newly hatched froglets					-	-	163			
Ether Soluble Acid Fraction		blastula embryos					-	-	5.26 (4.22- 6.55)	96 hr EC50=2.48 (1.49-4.11); EC0=1.00.		
		froglets					-	-	425			
Ether Soluble Base Fraction		blastula embryos					-	-	5.30 1.67- 17.07)	96 hr EC50=1.02 (.62-1.67); EC0=0.50		
		froglets					-	-	299			

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Fossil fuels (tar fractions) (cont'd)			A	Room temp.	-	-						Schultz et al., 1983 (cont'd)
Mono/di Aromatic Fraction		blastula embryos					-	-	17.91 (13.20-99.00)	96 hr EC50-21.47 (15.79-29.20); EC0<10.00.		
		froglets					-	-	776			
Poly Aromatic Fraction		blastula embryos					-	-	6.76 (5.21-8.77)	96 hr EC50-2.55 (1.48-4.38); EC0<1.00.	Malformations in head and gut, edema and growth retardation observed.	
		froglets					-	-	529			
Furanace (fish bactericide)	- Rana pipiens	larvae	A	16	Committee on Methods for Toxicity Tests with Aquatic Organisms 1975	total hardness 21 mg/L as $CaCO_3$	6.98 (5.55-8.57)	-	0.770 (0.590-1.01)		95% confidence limits in brackets. Larvae were more sensitive than eggs.	Marking et al., 1977

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Germanium (H)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977a,b	hardness 195.5 ± 5.4 ppm as $CaCO_3$		0.05		Exposure from fertilization to 4 days post hatch.	N = 16.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Heptachlor (OC)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	19.5 ± .5 (7.1)	-	30 ppm methyl orange	0.05 (0.19-3.8)	0.76 (0.23-2.7)	0.44 (0.30-0.65)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
n - Heptanol	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	52	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	44	-			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Hydrazine (propellant)	- Aedes aegypti, A. aquarius	larvae	A	22.2-24.0 (7.8-8.2)	APHA et al. 1971	185-232 mg/L as $CaCO_3$	>10	8.0	5.3			Slonim, 1986
			"	22.2-24.0 (6.3-6.9)	"	16-18 mg/L as $CaCO_3$	>10	5.2	2.3			
	- Xenopus laevis	embryos	A	-	-	-	-	-	-	EC50 (teratogenesis) for exposure from blastula to hatching - 11.48-12.5.		Crawshaw, 1977
Bis(2-Hydroxyethyl) amine	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	1174	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Bis(2-Hydroxyethyl) ether	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	7065	-		N = 10 per concentration.	de Zwart and Slooff, 1987

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Bis(3-Hydroxyethyl) ether	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	3181	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Bis(2-Hydroxypropyl) amine	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	418	-		N = 10 per concentration.	de Zwart and Slooff, 1987
IBP(S-benzyl diisopropyl) phosphorothioate	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	10	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
Iron Methanearsonate (H)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>40	-			Hashimoto and Nishiuchi, 1981
Cis - Kasugamycin (FY)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: 1.2 (0.63-2.4)	N = 28-34. 95% confidence limits in parentheses.	Cole and Cassida, 1983
Kasugamycin (F)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	6.4	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]			Effects[c]	Remarks	References
Lead (N)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.04		Exposure from fertilization to 4 days posthatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Linear Alkylbenzene Sulfonate	- Xenopus laevis	3-4 weeks	semi-static	21 ± 2	-	-	-	-	56-100		test solutions corrected for pH and oxygen content.	Canton and Slooff, 1982b
Lindane (OC)	- Bufo woodhousii fowleri	4-5 weeks old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	14 (5.8-41)	5.4 (3.1-9.6)	4.4 (1.8-5.6)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	4.0 (2.7-6.1)	3.8 (2.50-5.7)	2.7 (1.4-4.3)		"	
Malachite Green (F)	- Notophthalmus viridescens	adults	S	16 (7.5)	Committee on Methods for Toxicity Tests with Aquatic Organisms 1975	total hardness 20 mg/L as $CaCO_3$	3.90 (3.67-4.38)	-	1.83 (0.672-1.58)		95% confidence limits in brackets.	Bills et al., 1977

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c] 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
Malachite Green (cont'd)	Rana pipiens	larvae	S	16 (7.5)	Committee on Methods for Toxicity Tests with Aquatic Organisms 1975	total hardness 20 mg/L $CaCO_3$	0.380 (0.351-0.412)	-	0.173 (0.149-0.200)			Bills et al., 1977 (cont'd)
	Bufo sp.	larvae	"	"	"	"	0.355 (0.235-0.276)	-	0.0680 (0.0530-0.0860			
Malathion (OP)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	1.9 (1.4-3.5)	0.58 (0.25-1.3)	0.42 (0.09-0.98)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	0.56 (0.24-0.96)	0.32 (0.18-0.68)	0.20 (0.09-0.27)		"	
								15 day				
	- Rana pipiens	adults approx. 9 cm	renewed daily	24 (5.3-6.5)	-	-		150			N = 24 per concentration.	Kaplan and Glazewski, 1965

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Maneb (F) (Manganese ethylenebis-dithiocarbamate)	- Triturus cristatus	adults median length - 7.4 cm (males), 7.8 cm (females)	renewed daily	10 ± 2 (7.4)	-	Hardness 300 mg/L as $CaCO_3$	-	-	-	LT50 (lethal time for 50% of individuals) hr: Conc. Male Female 125 8.4 (7.6-9.2) 28.5 (16.4-49.6 100 28.0 (16.0-49.0) 19.5 (9.5-40.2) 75 19.0 (7.0-51.3) 25.5 (11.6-55.8) 50 76.0 (57.6-100.3) 168.0 (85.7-329.3) 25 255.0 (147.4-441.1)	N = 12 (6 males, 6 females) for each concentration. 95% confidence limits in parentheses. Females seemed less susceptible than males possibly due to larger body size.	Zafferoni et al., 1978

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Manganese (M)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		1.42		Exposure from fertilization to 4 days posthatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
	- Microtyle ornata	tadpoles: 1 week old	renewed daily	23.5-26.0 (6.86-6.94)	-	97.0-98.0 ppm	16.62 (16.2-17.0)	16.03 (15.6-16.4)	14.84 (14.6-15.4)		Range in parentheses. Animals not fed during test.	Rao and Madhyastha, 1987
		4 weeks old					17.36 (17.0-17.0)	16.52 (16.0-16.8)	14.33 (14.0-14.8)			
MCPA (M) Sodium salt (2-methyl-4-chlorophenoxy acetic acid)	- Triturus cristatus carnifex	adults	semi-static	10 ± 2	-	-	-	-	-	LT50 (lethal time for 50% of individuals) hr: Conc. Male Female 3200 17 (16.0-18.0) 21 (19.4-22.7) 1600 35 (31.0-40.9) 45.5 (41.0-50.0)	95% confidence limits in parentheses. Animals not fed during test.	Zafferoni et al., 1986b

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Mercury	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.001		Exposure from fertilization to 4 days posthatch.	N = 14. Toad is more sensitive than goldfish and rainbow trout.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
	- Bufo melanostictus	tadpoles 1.8-2.2 cm	A	29-34 (7.1-7.6)	APHA et al. 1976	120-160 mg/L as $CaCO_3$	.0528 (.0836-.0615)	.0856 (.0409-.0567)	.0836 (.0368-.0585)	In high concentrations surfacing, erratic body movement, loss of equilibrium noted.	95% confidence limits in brackets. Animals not fed 24 hr prior to or during test.	Khangarot and Ray, 1987
	- Microhyla ornata	tadpoles: 1 week old	renewed daily	25.5-26.0 (6.86-6.96)	-	97.0-98.0 ppm	2.04 (1.8-2.2)	1.68 (1.4-1.8)	1.12 (0.9-1.3)		Range in brackets. Animals not fed during test.	Rao and Madhvasthe, 1987
		4 weeks old					2.41 (2.0-2.8)	2.07 (1.8-2.4)	1.43 (1.0-1.8)			
	- Bufo regularis	adult females 25-30 g	-	-	-	-	-	-	-	96 hr LD50 following single i.m. injection of Hg^{+2} - 5.60		Hilmy et al., 1986b

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Mercury (cont'd)	- Gastrophryne carolinensis	embryo-larval	renewed every 12 hr	19 - 22 (7.0-7.8)	-	hardness 90-105 ppm $CaCO_3$		1.3 (0.9-1.9)		Exposure from fertilization to 4 days post-hatch.	95% confidence limits in brackets.	Birge et al., 1979b
	Hyla chrysoscelis	"	"	"	"	"		2.4 (1.5-3.4)				
	H. squirella	"	"	"	"	"		2.4 (1.5-3.8)				
	H. gratiosa	"	"	"	"	"		2.5 (1.7-3.4)				
	H. versicolor	"	"	"	"	"		2.6 (1.2-4.2)				
	H. crucifer	"	"	"	"	"		2.8 (1.9-3.9)				
	Rana pipiens	"	"	"	"	"		7.3 (4.8-10.8)				

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]:	7 day		Effects[d]	Remarks	References
Mercury (cont'd)	Acrid cricotiana blanchardi	embryo-larval	renewed every 12 hr	19-22 (7.0-7.8)	-	hardness 98-105 ppm $CaCO_3$		10.4 (8.5-12.6)		Exposure from fertilization to 4 days post-hatch.	95% confidence limits in brackets.	Birge et al., 1979b (cont'd)
	Bufo boreas	"	"	"	"	"		34.8 (28.3-51.1)				
	B. d. canorus	"	"	"	"	"		48.0 (29.6-92.2)				
	B. hemiophrys	"	"	"	"	"		59.9 (33.8-65.9)				
	B. fowleri	"	"	"	"	"		65.9 (46.8-84.8)				
	B. woodhousei	"	"	"	"	"		67.2 (56.3-79.5)				
	Ambystoma tigrinum	"	"	"	"	"		107.5 (72.5-193.3)				

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Mercury (Mercuric chloride)	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	0.4	-	-	N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	0.1	-	-		
	- Hyalella azteca	embryos in gastrulation	A	21-25 (7.9)	-	51 mg/L as $CaCO_3$	-	-	0.1784		N = 10 embryos per concentration.	Chapin and Heitmuller, 1980a
		tadpoles 8-10 days post-hatch	"	"	"	"	-	-	0.1184			
	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	0.1	-		N = 10 per concentration.	de Zwart and Slooff, 1983
Methiocarb (seven repetitions)	- Rana sphenocephala	larvae	A	16	ASTM Committee E-35 on Pesticides 1980	hardness 24 mg/L as $CaCO_3$	8.5 (7.9-9.1)	-	8.7 (7.8-9.6		95% confidence intervals in brackets.	Herring and Chandler, 1981
Methanol	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>48	-			Hashimoto and Nishiuchi, 1981

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Methoxychlor (MC)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	0.76 (0.52-1.1)	0.11 (0.02-0.60)	-		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	0.44 (0.30-0.65)	0.42 (0.29-0.62)	0.33 (0.19-0.57)			
1-Methyl-4 (tert)butylbenzene	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	5.0	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Methyl Parathion	- Rana crassiphilus	adults	renewed daily	23 ± .2 (7.3-7.8)	-	60-70 mg/L hardness				Significant variations observed in muscle, liver and kidney glycogen levels.	95% confidence intervals in parentheses.	Mukhali and Patil, 1987
		male					28.0 (21.6-42.0)	17.7 (16.65-18.8)	8.0 (7.7-8.3)			
		female					37.0 (30.2-45.4)	24.0 (22.0-26.2)	11.5 (11.2-12.2)			

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
4-Methyl-2-pentanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	656	-		N = 10 per concentration.	de Zwart and Slooff, 1987
2-Methyl-1-propanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	10.3	-		N = 10 per concentration.	de Zwart and Slooff, 1987
2-Methyl-2-propanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	2430	-		N = 10 per concentration.	de Zwart and Slooff, 1987
2-Methyl-quinoline	- Xenopus laevis	early cleavage to mid-blastula embryos	A	20	-	-	-	-	26.6 (22.6-31.1)		95% confidence limits in brackets. N = 25. Development retarded, reduction in motility.	Dumont et al., 1979
Mevinphos (MP)	- Bufo americanus	adults	-	-	injected via lymphatic sac	-	-	-	-	96 hr LD50: 850		Juarez and Guzman, 1984a
Mevacarbate (C)	- Rana catesbeiana	adults male	-	-	single dose - insertion of gelatin capsules to the level of the proventriculus	-				Estimated 14 day LD50 = 700-800.	Animals fasted for 2-24 hr prior to test. N = 14 males. Bullfrog more resistant than birds and mammals.	Tucker and Crabtree, 1969

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]				Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	72 hr	96 hr			
Mine Drainage (acidic)	- Lepomis/Hus macrochirus	larvae	S	12 (3.9)	-	-	53% leachate	47% leachate	33% leachate	-	LT50 ± SE (time of exposure causing 50% mortality): 33% leachate - 61 ± .93 hr; 45% leachate - 35 ± .66 hr; 63% leachate - 21 ± 1.82 hr; 100% leachate - 19 ± 2.15 hr.	Mortality rate was a function of exposure time and concentration.	Mathews and Morgan, 1982; Mathews et al., 1976
	- Salmonidae	embryos	A		modified Dumont et al. 1983						96 hr EC50 malformation:	Metal concentrations at mixing (ppm):	Dawson et al., 1985
				- (3.4)			-	-		13.1-15.8	13.1-13.48	Fe: 192.0-433.0; Zn: 178.0-235.0; Pb: <0.005-0.008; Cd: 0.055-0.104; Cu: <0.04	
				- (3.3-3.9)			-	-		27.3-34.3	30.3-33.3%	Fe: 414.0-533.0; Zn: 47.2-73.0; Pb: <0.005-0.008; Cd: <0.005-0.004; Cu: <0.04-0.04	

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[b]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Phlorate (H)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.3 ± .5 (7.1)	-	30 ppm methyl orange	33 (19-60)	28 (12-59)	14 (4.2-34)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
								10 day				
Malaw (F)	- Lepomis larvae	tadpoles 5-12 days old	A	22 ± 1	-	-		2		Treatment was continuous for 10 days; LC50 was determined at 10 days post exposure.		Birch and Prahlad, 1984
							24 hr	48 hr	96 hr			
Malaf (DP)	- Pseudacris triseriata	1 week old tadpoles	A	15.3 ± .5 (7.1)	-	30 ppm methyl orange	2.2 (0.85-6.0)	2.0 (0.90-5.0)	1.7 (0.90-3.2)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
Naphthalene	- Lepomis larvae	larvae 3 weeks old	S	20 (7.0-7.1)	-	-	-	-	2.1 (1.3-3.8)	6 hour EC50 for loss of pigment - 3.7 (3.1-4.5).	95% confidence intervals in brackets.	Edmisten and Bantle, 1982
									2.1 (1.6-2.9)	6 hour EC50 for absence of swimming - 3.3 (2.8-5.3), 3.7 (3.2-3.3).		

Table 9 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr / 7 day	Effects[d]	Remarks	References
Nickel (Ni)	- Coregonus clupeaformis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm Hardness as $CaCO_3$			0.05	Exposure from fertilization to 4 days posthatch.	N = 16.	Birge et al., 1979a
	- Rufa salmonoides	tadpoles 1.8-2.2 cm	A	29-34 (7.5-7.6)	APHA et al. 1976	120-160 mg/L as $CaCO_3$	53.21 (49.36-58.37)	34.3 (32.9-37.21)	25.32 (22.8-28.62)		95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Ray, 1987
Nitrite	- Ambystoma tigrinum	larvae 0.49 ± 0.08 g	A	25 (7.0)	E.P.A. 1975	140 mg/L total Hardness	-	-	1.09 (0.68-0.2)	Exposure greater than 10 ppm produced immediate stress, air gulping and mucus production.	95% confidence limits in parentheses.	Huey and Beitinger, 1980
Nitrofen	- Rufa bufo japonicus	tadpoles	-	-	-	-	-	3.8	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
Nitrilotriacetic Acid	- Isonepa lineata	3-4 weeks	mass-static	21 ± 2	-	-	-	-	560-1000		Test solutions corrected for pH and oxygen content.	Canton and Slooff, 1982b

Table 9 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
p-Nitro-toluene	- Isonepa lineata	>2 days	mass-static	20 ± 1	-	-	-	-	-	100 day exposure no effect levels (the highest concentration at which no effect or lethality was observed): Mortality - 10 Development - 1.2 Growth - 32	N = 75.	Slooff and Canton, 1983
DNPA (DP)	- Rana sylvatica	adults approx. 9 cm	renewed daily	25 (6.2 - 6.6)	-	-		19 day 2,900			N = 24 per concentration.	Kaplan and Glaremont, 1963

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
PA-14 (avian repressing agent)	- Rana sphenocephala	larvae	A	12	ASTM Committee E-35 on Pesticides 1980	hardness 24 mg/L as $CaCO_3$	7.5 (6.3-9.0)	-	5.9 (5.3-6.5)		95% confidence intervals in brackets.	Marking and Chandler, 1981
				14			4.5 (4.2-4.7)	-	4.2 (3.6-4.7)			
				22			2.6 (2.0-3.6)	-	2.3 (1.8-2.9)			
Paraquat (SP)	- Bufo viridis	adults 18-45 g	-	16	injection into dorsal lymphatic sac	-	-	-	-	7 day LD50: 180	N = 25 minimum.	Edery and Scheideberg-Perath, 1980
	Rana ridibunda									91		

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Paraquat (M)	- Ambistoma tigrinum	1-2 week old tadpoles	A	21-22	Sanders 1970	-	320	319	262			Johnson, 1976
	Limnodynastes peronii	1-2 week old tadpoles	"	"	"	"	204	153	100			
	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	15.9 ± .5 (7.1)	-	30 mg/L methyl orange	56 (30-100)	25 (9.4-66)	24 (11-43)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"		43 (18-96)	37 (28-92)	28 (21-36)		"	
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	16	-		Obtained from formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Amphibian toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Parathion (EP)	- Bufo viridis	adults 18-45 g	-	16	injection into dorsal lymphatic sac	-	-	-	-	7 day LD50: 967	N = 25 minimum.	Ebory and Schatzberg-Porath, 1960
	- Pseudacris triseriata	1 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	1.6 (0.60-3.0)	1.4 (0.91-2.8)	1.0 (0.30-2.0)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
								15 day				
	- Rana pipiens	adults approx. 9 cm	renewed daily	24 (5.0-6.8	-	-		10			N = 24 per concentration.	Kaplan and Glazewski, 1965
							24 hr	48 hr	96 hr			
	- Bufo arenarum	adults	-	-	injected via lymphatic sac	-	-	-	-	96 hr LD50: 3,352		Juarez and Guzman, 1984a
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	7.2	-			Hashimoto and Nishiuchi, 1981

Table 1 - Amphibian toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[d]	Remarks	References
Pentachlorophenol	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	0.38	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Beernsteen, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	0.26	-			
	- Rana catesbeiana	tadpoles 2-5 g	R	17.2-18.2 (8.01-8.04)	APHA 1980, USEPA 1974	-			0.207 (.185-.231)		Range in parentheses.	Thurston et al., 1985
	- Xenopus laevis	<2 days	Semi-static	20 ± 1	-	-	-	-	-	100 day exposure no effect levels (the highest concentration at which no effect or lethality was observed): Mortality - 0.032 Development - 0.032 Growth - 0.032	N = 75.	Slooff and Canton, 1983
Pentachlorophenol - Sodium	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	0.25	-		Obtained from formulated product.	Hashimoto and Nishiuchi, 1981

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Experimental Conditions: Temp. °C (pH)	Experimental Conditions: Methodology	Experimental Conditions: Alkalinity	LC 50[c]: 24 hr	LC 50[c]: 48 hr	LC 50[c]: 96 hr	Effects[d]	Remarks	References
Permethrin (PY)	- Rana catesbeiana	tadpoles 2-3 g	S	17.3 18.0 (7.98-8.01)	APHA 1980, USEPA 1974	-	-	-	0.115 (.0538-.265)		Range in parentheses.	Thurston et al., 1985
	- Rana catesbeiana	tadpoles 6-8 mm 0.01 g	A	24 ± 1 (8.4)	Doudoroff et al. 1951	100 mg/L hardness	-	-	7.033 (6.129-8.733)		N = 10 per concentration. 95% confidence limits in brackets. Frogs more tolerant than fish or crayfish.	Jolly et al., 1978
(1R-permethrin)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: trans 7.5 (5.2-10.9); cis 0.14 (0.11-0.17)	N = 28-34. 95% confidence limits in parentheses.	Cole and Casida, 1983
Phenol	- Xenopus laevis	tadpoles	S	17.2 ± 1 (7.39 ± .22)	-	48.4-49.1 mg/L as $CaCO_3$	-	-	>91.1			Holcombe et al., 1983

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Experimental Conditions: Temp. °C (pH)	Experimental Conditions: Methodology	Experimental Conditions: Alkalinity	LC 50[c]: 24 hr	LC 50[c]: 48 hr	LC 50[c]: 96 hr	Effects[d]	Remarks	References
1R-Phenothrin (PY)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: trans greater than 20; cis 4.0 (3.9 - 9.2)	N = 28-34. 95% confidence limits in parentheses.	Cole and Casida, 1983
Phenyl Mercury Acetate	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	0.12			Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
N-Phenyl-2-naphthylamine	- Xenopus laevis	larvae	A	-	-	-	-	2.1-2.3	-			Greenhouse, 1977
		embryos	-	-	-	-	-	-	-	LC50 (teratogenesis) for exposure from blastula to hatching-4.57-4.8.		
								15 day				
Phosdrin (DP)	- Rana pipiens	adults approx. 9 cm	daily renewal	24 (6.8-8.4)	-	-		12			N = 24 per concentration.	Kaplan and Glassmanis, 1965
Phosphamidon (DP)	- Bufo arenarum	adults	-	-	injected via lymphatic sac	-	-	-	-	96 hr LD50: 1,195		Juarez and Guzman, 1984a

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Picloram (H)	- Adelotus brevis	1-2 week old tadpoles	A	21-22	Sanders 1970	-	143	123	95			Johnson, 1976
		4 week old tadpoles	"	"	"	"	210	182	154			
	Limnodynastes peroni	1-2 week old tadpoles	"	"	"	"	120	114	105			
Piperonyl Butoxide (synergist)	- Pseudacris triseriata	1 week old tadpoles	A	15.5 ± .5 (7.1)	-	10 ppm methyl orange	1.8 (0.40-8.2)	1.3 (0.30-12)	1.0 (0.10-9.8)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
Potassium Dichromate	- Xenopus laevis	~2 days	semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed): Mortality - 1 Development - 3.2 Growth - 3.2	N = 75.	Slooff and Canton, 1983

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Propanil (H)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	2.3	-			Hashimoto and Nishiuchi, 1981
n-Propanol	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	4,000	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Ravensloven, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	4,000	-			
2-Propenyl Amine	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	12.4	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Bis (2-Propenyl) Amine	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	25.5	-		N = 10 per concentration.	de Zwart and Slooff, 1987
								1 hr				
Pyrone	- Rana pipiens	embryos stage 25	A	-	-	-		0.14			LC50 determined for 1 hour of exposure to sunlight.	Kagan et al., 1985
Irano - Pyrethrone (PY)	- Rana pipiens	adults 20-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: 5.8 (4.9-7.0) N = 28-34.	95% confidence limits in parentheses.	Cole and Casida, 1983

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
Pyridine	- Ambystoma gracilarum	3-4 weeks post hatch	A	20 ± 1	-	-	-	950	-		N = 10 for each species. Organisms not during expt.	Sloff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	1,400	-			
							24hr	48hr	72hr	96hr		
	- Xenopus laevis	mid-blastula embryos	A	Room Temp.	-	98.5 mg/L	3880 (2520-5760)	2570 (1880-3520)	2340	-	95% confidence limits in brackets.	Davis et al., 1981
		tailbud embryos					9550 (1710-53000)	3390 (3000-3830)	2820 (2350-3380	2440 (1710-3540)		
		free swimming larvae					1440[e]	1590 (1190-2110)	1200 (980-1460)	1090 (900-1340		

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	72 hr	96 hr	Effects[c]	Remarks	References
Quinoline	- Xenopus laevis	mid-blastula embryo	A	Room Temp.	-	98.5 mg/L	219	115 (53-267)	87 (65-167)	79 (51-123)	Teratogenicity EC50 for mid-blastula embryos: 24 hr - 71 (38-134); 72 hr - 34 (15-90); 96 hr - 29 (14-61).	95% confidence limits in brackets.	Davis et al., 1981
		tailbud embryo					280 (157-245)	148 (117-186)	129 (102-164)	129 (98-169)			
		free swimming larvae					135 (91-200)	117 (78-176)	107 (98-127)	95 (75-120)			
	- Xenopus laevis	early cleavage to mid-blastula embryos	A	20	-	-	-	-	-	24.3 (22.1-31.3)		N = 25. 95% confidence limits in brackets. Caused reduced motility, and retarded development.	Dumont et al., 1979

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
RRS - Resmethrin (FY)	- Rana catesbeiana	tadpoles 10-15 g	-	20	intraperitoneal injection	-	-	-	-	24 hr LD50: trans 5.6 (5.4-8.8); cis 1.2 (0.83-1.6)	N = 32-45. 95% confidence limits in parentheses.	Cole and Casida, 1983
	Rana pipiens	adults 20-30 g	-	-	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: trans greater than 60; cis 1.3 (1.0-1.6)	N = 28-34. 95% confidence limits in parentheses.	
Rotenone	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	0.33	-			Hashimoto and Nishiuchi, 1981
Rotenone (Noxfish-9% rotenone)	- Rana sphenocephala	larvae	A	16 ± 1 (6.6)	Committee on Methods for Toxicity Tests with Aquatic Organisms 1975	20 mg/L	0.588 (.694-.680)	-	0.500 (.423-.591)		95% confidence intervals in brackets.	Chandler and Marking, 1982
Rotenone (Rotenoids)	- Xenopus laevis	tadpoles	S	17.2 ± 5 (7.39 ± .22)	-	48.4-49.5 mg/L as $CaCO_3$	-	-	≥0.060			Holcombe et al., 1987

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
Salicylaldehyde	- Ambystoma mexicanum	3-4 weeks post hatch	A	20 ± 1	-	-	-	7.0	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baerselman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	7.7	-			
								7 day				
Selenium (IV)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.09		Exposure from fertilization to 4 days posthatch.	N = 10.	Birge et al., 1979a
								72 hr				
Selenium (sodium selenite)	- Xenopus laevis	tadpoles	renewed daily	23	-	-		8.0A (6.99-9.25)		LT50 (median survival time): 4.7 days in 2 ppm, 4.0 days in 5 ppm, 2.54 days in 10 ppm.	95% confidence limits in brackets. ≥2 ppm produced abnormalities.	Browne and Dumont, 1979
							24 hr	48 hr	96 hr			
Silver (II)	- Bufo melanostictus	tadpoles 1.8-2.2 cm	A	29-34 (7.1-7.4)	APHA et al. 1976	120-140 mg/L as $CaCO_3$	.0073 (.00637-.00809)	.0062 (.00500-.00700)	.0061 (.00764-.00641)	In high concentrations surfacing, erratic body movement, loss of equilibrium noted.	95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Ray, 1987

Table 1 - Amoko Toxicity (cont'd)

Contaminant[A]	Species	Stage/age	Experimental Conditions				LC 50[C]			Effects[C]	Remarks	Reference
			Flow[B]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Simazine (H)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>100	-		Obtained from formulated product.	Hashimoto and Nishiuchi, 1981
Sodium Aluminium Silicate	- Xenopus laevis	3-4 weeks	semi-static	21 ± 2	-	-	-	-	5,600-10,000		Test solutions corrected for pH and oxygen content.	Canton and Slooff, 1982b
Sodium Arsenite (H)	- Ambistoma gracile	1-2 week old tadpoles	A	21-22	-	-	152	119	96			Johnson, 1976
	Limnodynastes peronii	1-2 week old tadpoles	"	"	"	"	188	92	60			
	Bufo marinus	2 week old tadpoles	"	"	"	"	195	150	123			
Sodium Bromide	- Xenopus laevis	42 days	semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed): Mortality - 32 Development - 320 Growth - 320	N = 75.	Slooff and Canton, 1983

Table 1 - Amoko Toxicity (cont'd)

Contaminant[A]	Species	Stage/age	Experimental Conditions				LC 50[C]			Effects[C]	Remarks	Reference
			Flow[B]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Sodium Fluoroacetate (1080)	- Limnodynastes tasmaniensis	adults 2-5 g	-	-	Single i.p. injection	-	-	-	-	LD50 approx. 60 ppm. Time until death (median and range) - 78.4 hr (36.8-98.3)		McIlroy et al., 1985
Sodium or Methyl Fluoroacetate	- Rana pipiens		-	-	Subcutaneous injection	-	-	-	-	LD50 = 140.		Chenoweth, 1949
	Xenopus laevis		-	-	Subcutaneous or i.p. injection	-	-	-	-	LD50 >500.0.		
Sodium p-Fluoroacetamide	- Rana pipiens		-	-	Subcutaneous injection	-	-	-	-	LD50 = 25.0.		Chenoweth, 1949
								10 day				
Sodium Thiocyanate	- Xenopus laevis	tadpoles 9-12 days old	A	22 ± 1	-	-		2,000		Treatment was continuous for 10 days; LC50 was determined at 10 days post exposure.		Birch and Prahlad, 1986

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
Sour water from coal gasification plant	- *Xenopus laevis*	early cleavage to mid-blastula embryos (stages 2-6.5)	A	20 (6.4 for controls; 6.7 for 1% sour water)	-	-	0.69* (0.61-0.77)	0.36* (0.34-0.38)	-	*Levels in percent sour water content.	N = 200 per treatment. 95% confidence intervals in brackets. Reduction in motility, pigmentation and rate of embryonic development noted.	Dumont and Schultz, 1980
								7 day				
Strontium (II)	- *Gastrophryne carolinensis*	embryo-larval	S	- (7.8-7.8)	Birge & Black 1977 a, b	195.8 ± 5.4 ppm hardness as $CaCO_3$		0.16		Exposure from fertilization to 4 days posthatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
2,4,5-T amine (H) (2,4,5-trichlorophenoxyacetic acid)	- *Adelotus brevis*	1-2 week old tadpoles	A	21-22	Sanders 1970	-	228	205	200			Johnson, 1976
	Limnodynastes peroni	1-2 week old tadpoles	"	"	"	"	210	190	169			
	Bufo marinus	2 week old tadpoles	"	"	"	"	425	382	340			

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]: 24 hr	48 hr	96 hr	Effects[c]	Remarks	References
EDE (OC)	- *Bufo woodhousii fowleri*	4-5 week old tadpoles	A	15.5 ± .5 (7.1)	-	30 ppm methyl orange	0.70 (0.25-2.0)	0.32 (0.21-0.45)	0.14 (0.10-0.21)	N = 10. 95% confidence limits in brackets.		Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	0.61 (0.41-0.82)	0.58 (0.21-0.75)	0.48 (0.21-0.75)	"		
FEPP (OP)	- *Bufo viridis* *Rana ridibunda*	adults 18-45 g	-	25	Injection into dorsal lymphatic sac	-	-	-	-	24 hr LD50: 540 34	N = 25 minimum.	Fiory and Schatzberg-Porath, 1960
								15 day				
	- *Rana pipiens*	adults approx. 9 cm	daily renewal	24 (5.3-6.4)	-	-		40			N = 24 per concentration.	Kaplan and Glazenski, 1965
							30 min	2 hr				
α-Terthienyl	- *Rana pipiens*	embryos stage 25	A	-	-	-	0.11	0.018		LC50 determined for given length of exposure to sunlight.		Kagan et al., 1984

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
o-Terthienyl (cont'd)	- Hyla crucifer	tadpoles	-	-	-	-	.003	-	-	Initial incubation time was 1 hr, irradiation time (sunlight and UV) was 1 hr; survival determined 24 hr after after treatment.		Kagan et al., 1987
Tetrachlorophthalide	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	>40	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981
1R Tetramethrin (FY)	- Rana pipiens	adults 25-30 g	-	20	subcutaneous injection into the dorsum	-	-	-	-	24 hr LD50: trans greater than 20; cis 1.8 (1.2-2.6)	N = 28-34. 95% confidence limits in parentheses.	Cole and Casida, 1983
Tetrapropylene Benzene Sulphonate	- Xenopus laevis	<2 days	semi-static	20 ± 1	-	-	-	-	-	100 day exposure No Effect Levels (the highest concentration at which no effect or lethality was observed): Mortality - 3.2 Development - 10 Growth - 10	N = 75.	Slooff and Canton, 1983

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
TFM (Lamprecide) (3-trifluoro-methyl-4-nitrophenol	- Hyla versicolor	larvae	A	17 ± 1 (6.8-7.0)	-	hardness 44 mg/L as $CaCO_3$	-	-	1.98 (1.77-2.22)		99.4% TFM tested. 95% confidence intervals in parentheses.	Chandler and Marking, 1975
	Rana pipiens	larvae	"	"	"	"	-	-	2.76 (2.45-3.11)		Difficult to compare results of static vs flow through assays.	
	R. catesbeiana	larvae	8 renewed 3 times per day	16-17	-	"	-	-	3.55 (2.62-4.82)			
							7 day					
Thallium (M)	- Gastrophryne carolinensis	embryo-larval	8	- (7.0-7.8)	Birge & Black 1977 a, b	195.8 ± 3.4 ppm hardness as $CaCO_3$		0.11		Exposure from fertilization to 4 days posthatch.	N = 16.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Thiram (F)	- Xenopus laevis	47	A	20 ± .5 (7.5)	-	230 ppm $CaCO_3$ hardness	0.017	0.014	0.013			Saoud et al., 1983
		53	"	"	"	"	0.025	0.022	0.021			

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity		7 day				
Tin (M)	- Gastrophryne carolinensis	embryo-larval	S	- (7.8-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.09		Exposure from fertilization to 4 days post hatch.	N = 14.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Toxaphene (EC)	- Rana sphenocephala	eggs	S	20 (7.2-7.5)	-	total hardness 100 ppm as $CaCO_3$	-	-	0.651	Exposed for 96 hr, mortality determined at 15 days.	N = 20-30 per concentration.	Hall and Swineford, 1980
		larvae	"	"	"	"	-	-	0.065 (0.055-0.083)	"	N = 20-30 per concentration. 95% fiducial limits in brackets.	
		sub-adults	"	"	"	"	-	-	0.378 (0.190 1.125)	Exposed for 96 hr, mortality determined at 8 days.	N = 5-10 per concentration. Flow through method produced lower results than static method.	
	- Bufo woodhousii	4-5 week old tadpoles	A	15.5 ± 0.5 (7.1)	-	30 ppm methyl orange	.60 (0.30-1.2)	0.29 (0.20-0.42)	0.14 (0.06-0.35)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	1.7 (0.98-3.2)	0.70 (0.48-1.2)	0.50 (0.18-1.1)		"	

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[c]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Toxaphene (cont'd)	-		S	20 (7.2-7.5)	Hall and Swineford 1980	100 ppm as $CaCO_3$ hardness				24 hr EC50 (behavioural aberrations):	95% confidence limits in brackets.	Hall and Swineford, 1981
	Rana sphenocephala	larvae					-	-	0.130 (0.095-0.184)	0.193 (0.138-0.298)		
	R. catesbeiana	larvae					-	-	0.099 (0.071-0.166)	0.312 (0.229-0.635)	Disturbed equilibrium, abnormal posture, erratic swimming noted.	
	R. erythrea	larvae					-	-	0.199 (0.176-0.220)	0.036 (0.023-0.053)		
	Bufo americanus	larvae					-	-	0.034 (0.027-0.042)	0.038 (0.014-0.091)		
	Acris crepitans	larvae					-	-	0.076 (0.066-0.116)	>1.00		
	Ambystoma opacum	larvae					-	-	0.342 (0.265-0.470)	0.170 (0.122-0.235)		

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
Toxaphene (cont'd)	Ambystoma gracilicum fowleri	larvae	R	20 (7.2-7.5)	Hall and Swineford 1980	100 ppm as $CaCO_3$ hardness	-	-	0.034 (0.020-0.047)	24 hr EC50: 0.227 (0.215-0.234)	95% confidence limits in brackets	Hall and Swineford, 1981 (cont'd)
TPN (tetrachloroisophthalonitrile)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	0.14	-			Hashimoto and Nishiuchi, 1981
Trichloroethylene	- Ambystoma gracilicum	3-4 weeks post hatch	A	20 ± 1	-	-	-	48	-		N = 10 for each species. Organisms not fed during expt.	Slooff and Baarsmaman, 1980
	Xenopus laevis	3-4 weeks post hatch	"	"	"	"	-	45	-			
2,4,6-Trichlorophenol	- Xenopus laevis	tadpoles	R	17.2 ± 5 (7.39 ± .22)	40.6-69.5 mg/L as $CaCO_3$		-	-	1.30 (1.08-1.40)		95% confidence interval in parentheses.	Holcombe et al., 1987

Table 1 - Acute toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions				LC 50[c]			Effects[d]	Remarks	References
			Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	24 hr	48 hr	96 hr			
2-(2,4,5-Trichlorophenoxy) Propanoic acid, butyl ether ester (H)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	19.5 ± .5 (7.1)	-	30 ppm methyl orange	22 (15-32)	20 (13-28)	-	N = 10. 95% confidence limits in brackets.		Sanders, 1970
	Pseudacris triseriata	1 week old tadpoles	"	"	"	"	29 (16-32)	18 (10-31)	10 (6.8-18)			
Trifluralin (H)	- Bufo woodhousii fowleri	4-5 week old tadpoles	A	19.5 ± .5 (7.1)	-	30 ppm methyl orange	0.18 (0.10-0.30)	0.17 (0.10-0.31)	0.10 (0.06-0.49)		N = 10. 95% confidence limits in brackets.	Sanders, 1970
	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	14	-		Obtained from formulated product.	Hashimoto and Nishiuchi, 1981
3,5,5-Trimethyl-1-hexanol	- Xenopus laevis	3-4 week old larvae	A	20 ± 1	-	-	-	13.5	-		N = 10 per concentration.	de Zwart and Slooff, 1987
Trithion (OP)	- Rana pipiens	adults approx. 9 cm	renewed daily	24 (6.7)	-	-		15 day 199			N = 24 per concentration.	Kaplan and Glazmanski, 1963

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]			Effects[c]	Remarks	References
								7 day				
Tungsten (N)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		2.90		Exposure from fertilization to 4 days posthatch.	N = 16.	Birge et al., 1979a
							24 hr	48 hr	96 hr			
Yalen (N)		tadpoles 1-1.5 cm	renewed daily	17-20 (7.7-8.2)	-	-	-	19.1	-	Value is the LC_{50} of the active ingredient.		Perevozchenko, 1975
								7 day				
Zinc (N)	- Gastrophryne carolinensis	embryo-larval	S	- (7.0-7.8)	Birge & Black 1977 a, b	195.0 ± 5.4 ppm hardness as $CaCO_3$		0.01		Exposure from fertilization to 4 days posthatch.	N = 16.	Birge et al., 1979a
								15 day				
	- Rana pipiens	adults approx. 9 cm	renewed daily	24 (6.7)	-	-		155			N = 24 per concentration.	Kaplan and Glenzenski, 1965

Table 1 - Acute Toxicity (cont'd)

Contaminant[a]	Species	Stage/age	Experimental Conditions: Flow[b]	Temp. °C (pH)	Methodology	Alkalinity	LC 50[c]			Effects[c]	Remarks	References
								7 day				
Zinc (cont'd)	- Bufo valliceps	tadpoles 1.0-2.2 cm	A	29-30 (7.1-7.6)	APHA et al. 1976	120-140 mg/L as $CaCO_3$	47.26 (39.48-58.79)	25.65 (23.73-27.76)	19.86 (17.68-23.98)		95% confidence limits in brackets. Animals not fed 24 hr prior to or during tests.	Khangarot and Rao, 1987
	- Hyperolia arenicolor	tadpoles	renewed daily	25.5-26.0 (6.86-6.94)	-	97.0-98.0 ppm					Ranges in parentheses. Animals not fed during test.	Rao and Madhyastha, 1987
		1 week old					24.06 (23.6-24.4)	23.42 (23.0-23.8)	22.41 (22.0-22.8)			
		4 weeks old					25.42 (24.8-25.6)	24.38 (24.0-24.8)	23.08 (22.6-23.4)			
Zinc Sulphate	- Xenopus laevis	embryos blastula	static renewal	22-24 (7)	-	100 mg/L $CaCO_3$ hardness	-	-	34.5 ± 1.2	EC50 (malformation)-3.6 ± 0.5.	Mean ± confidence interval.	Dawson et al., 1988
Ziram (F)	- Bufo bufo japonicus	tadpoles	-	-	-	-	-	48	-		Obtained using formulated product.	Hashimoto and Nishiuchi, 1981

TABLE 2
SOMATIC EFFECTS - LABORATORY STUDIES

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Abate (OP)	- Bufo boreas	20	juveniles (T snout-vent length - 19.8 mm)	Exposure of hydrated toads to 60 ppb significantly lowered temperature tolerance.	23-26°C; 24 hr exposure.	Johnson and Prine, 1976
Acephate (OP)	- Rana catesbeiana		tadpoles	All tadpoles exposed to 5 ppm survived.	Continuous flow apparatus for 96 hr. Brain cholinesterase activity of mallards fed dosed tadpoles was 109.7% of controls; non-	Hall and Kolbe, 1980
	- Ambystoma gracile	200-400 per treatment	eggs, larvae	Egg hatch was not significantly affected by concentrations up to 798 mg/L; in first week post-hatch, mortality was directly related to concentration to which larvae exposed. Growth rate decreased and abnormalities increased when larvae were exposed to 382 and 798 mg/L.	The maximum surface water concentration of acephate used in insect control programs should be <1 mg/L and should have little effect on populations of A. gracile	Goan et al., 1984
	- Rana clamitans	7 per treatment	larvae	Significant decrease in mean activity time recorded at 1000 ppm but not at 500 ppm.	Temp. 21 ± 1°C. 24 hr. acclimation.	Lyons et al., 1976
Azadirachtin Extract	- Rana temporaria		tadpoles	>400 ppm caused 100% mortality, 200 ppm caused 50% mortality, 200 ppm produced no lethal effect during 1 week observation. At 10 ppm accelerated metamorphosis by 2 days in 80% animals observed; 200 did not complete metamorphosis, and were malformed.		Pavlov and Pavlovova, 1983

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS					REMARKS	REFERENCE
Aldrin (OC)	- Rana cyanophlyctis			Exp#	Wght of frog (gm)	Lgth of frog (mm)	TLM % concentration (ppm)	Length of time until death (min)	pH - 7.9. Room temp. - 19-26°C. Water temp. - 20-26°C.	Rana and Mathur, 1978
		6		1	3-4.5	26-35	0.006	180-215		
		2		2	2	21-25.5	0.03	75-95		
		3		3	6-8	32-43	0.06	140-180		
		4		4	3-4	30-35	0.09	85-155		
		5		5	6-5	36-38	0.1	90-110		
		6		6	3-10	27-46	0.125	55-120		
	- Acris crepitans	160	frogs 18-23 mm snout-vent length	Frogs taken from areas treated or untreated with pesticides and exposed to 30,000 or 50,000 mg/L aldrin in lab. Mortality recorded after 24 and 36 hr. Percent mortality was lower in frogs from treated fields than in frogs from areas with no or minimum prior treatment.					Areas treated with DDT, endrin, toxaphene, methyl parathion. Aldrin not used. Thus, resistance must represent a cross-resistance. Temp. 21-26°C.	Vinson et al., 1963
	A. gryllus	50		10,000 mg/L killed 40% of frogs. 50,000 mg/L killed 70% in 36 hours.						
	- Rana pipiens		abdominal skin	No changes in short-circuit current at 3×10^{-4} M.						Webb et al., 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Aldrin (cont'd)	- Bufo arenarum		embryos	5 and 15 ppm produced 100% mortality on day 15 and 10, respectively. 1 ppm was not toxic.	Exposed from fertilization, medium replaced daily.	Juarez and Guzman, 1984b
	- Rana pipiens		liver microsomes (adults)	Microsomal metabolism (apparent V_{max} and K_m values ± SE): App V_{max} = 0.036 ± 0.01 nmol/min/mg. App K_m = 4.3 ± 3 nmol/mg.		Ronis and Walker, 1985
	- Rana pipiens		adults 65 g, 8.9 cm long	Aldrin (ppm) / pH of solution at 25°C / No. of frogs dead at 30 days	Frogs placed in 200 cc of test solution.	Kaplan and Overpeck, 1966
		20		0.38 / 5.90 / 8	Neuromuscular changes produced: excessive thrashing and abnormal reactivity to stimulation were observed.	
		10		0.23 / 5.71 / 0		
		10		0.19 / 5.68 / 0		
Aldrin-transdiol	- Xenopus laevis		lateral-line organ	2.5×10^{-6} and 7.5×10^{-6} M failed to induce repetitive activity; caused marked increase in the rate of spontaneous firing, followed by a blockade.	Temp. 19-21°C.	Abbamonte et al., 1975a
			cutaneous touch receptors	Exposure to 10^{-5} M failed to induce repetitive nerve spikes; possible blocking effect.	No effects of temperature changes noted.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Aldrin - transdiol (cont'd)	- Rana esculenta R. temporaria		sartorius nerve-muscle	2.5×10^{-5} M produced increase in end-plate potential to 4 times control level at 20 min, after which amplitude declined and transmission completely blocked.	Temp. 10 ± 0.2°C. Hypothesis that aldrin-transdiol increases amount of calcium entering nerve terminal during nerve impulse.	Abbamonte et al., 1975b
	- Rana esculenta R. pipiens R. temporaria		motor end-plate	Concentrations of 10^{-5} to 10^{-4} exerted both pre- and post-synaptic actions; caused increase in end-plate potential frequency and decrease in their amplitude.	Temp 10 ± 0.2°C.	Abbamonte et al., 1976
	- Xenopus laevis		spinal chord	Application of 1 to 5 x 10^{-5} M in vitro caused potentiation of spinal reflex activity, increase in spontaneous activity of ventral and dorsal roots and reduction of spinal inhibitory mechanisms.	Temp. 10 ± 2°C.	Abbamonte et al., 1975c
Allethrin (PT)	- Xenopus laevis		lateral-line organ	Repetitive activity induced by application of 1-3 ppm allethrin for 20-40 min.		Abbamonte et al., 1975d
			cutaneous touch receptors	Produced repetitive activity after 5 min exposure to 10^{-5} M or 15 min to 10^{-6} M.	Slight increase in number of repetitive spikes observed when temperatures of preparation lowered from 19-21°C to 10-15°C. Negative temperature co-efficient of activity.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Allethrin (cont'd)	- Xenopus laevis		myelinated fibre from sciatic nerve	2×10^{-5} M hardly affected rate of rise, amplitude and declining phase of sodium current during test pulse. Upon repolarisation, large, slow sodium tail current apparent.	Temp. 15 ± 1°C. Concluded that allethrin acts on open sodium channels only.	van den Bercken and Vijverberg, 1980
	- Rana pipiens		nerve-muscle	Muscle contraction evoked by nerve stimulation was augmented somewhat at low concentrations (10^{-6} - 10^{-5} M) but was suppressed at higher concentrations ($1-5 \times 10^{-4}$ M).	Temp. 22°C.	Takeno et al., 1977
	- Rana esculenta R. temporaria Xenopus laevis		peripheral nervous system	Induced repetitive activity in sensory nerve fibres (0.3) - 3.3 uM), sense organs (1-10 uM) and in distal part of the motor fibres, leading to pronounced repetition in the motor end plate (0.1 - 10 uM).	May account for hyperexcitation and tremors leading to paralysis and death. Negative temperature coefficient of activity in lateral-line sense organ and motor nerve terminal.	van den Bercken, 1977
	- Xenopus laevis		adults	Exposure to 5 ppm caused excitation and convulsions within 10 min; excitation also occurred following exposure to 1 ppm for 30 min.		van den Bercken et al., 1973
			lateral line organ	2 ppm (6.6×10^{-6} M) in vitro caused pronounced repetitive activity.	Similarity between EDI and allethrin noted.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Allethrin (cont'd)	- Rana temporaria R. esculenta		motor-end plate	10^{-7} M produced pronounced repetitive activity in the motor end plate; a single nerve stimulus could provoke up to 13 end plate potentials.	Showed negative temperature coefficient.	Houtsmo et al., 1977
Alloxid (ICR)	- Bufo boreas	20	juveniles (T snout-vent length - 19.8 mm)	Exposure of hydrated toads to 100 ppb significantly lowered temperature tolerance.	23-24°C; 24 hr exposure.	Johnson and Prine, 1976
Aluminum (N)	- Rana catesbeiana	26 7	adults 100-300 g brain	Al^{+3} enriched water environment produced no measurable changes in ACHE levels in brain, in vivo or in vitro.	pH 4.6-6.6.	Marquis, 1982
	- Ambystoma maculatum	18-46 per treatment	embryos late blastula stage	Between pH 4.5 - 6.0 Al had no effects on hatching success; at pH 4.0, hatching success decreased from 11% at 0.1 ppm total Al to 7% at 0.4 ppm and 2% at 0.7 and 1.1 ppm. All larvae hatched at pH 4.0 died within a few days.	Exposure for 31 days at 21°C.	Dale et al., 1985
	Hyla crucifer	11-34 per treatment	embryos mid-blastula	No consistent effect on hatching success at any pH where hatching occurred (pH 4.5, 5.0, 6.0).	Exposure for 16 days at 21°C.	
	Bufo americanus	47-52 per treatment	embryos early gastrula stage	Hatching >90% at all pH's and total Al concentrations from pH 4.0 - 6.0.	Exposure for 11 days at 21°C.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Aluminum (cont'd)	Xenopus laevis	10 per treatment	embryos prior to 4 cell stage	% hatching (% of hatched larvae dead at end of expt. in parentheses) in oligotrophic brown water:	Exposure for 11 days at 21°C. At .05 - .15 ppm Al, high frequency deformed larvae found at pH 4.5; malformed larvae also found at pH >5.0 and high Al (=.35).	Dale et al., 1985 (cont'd)

Al Concentration (ppm)

pH	0.15	0.20	0.40	0.90
3.5	0 (-)	0 (-)	0 (-)	0 (-)
4.0	0 (-)	0 (-)	0 (-)	0 (-)
4.5	80 (100)	100 (30)	100 (0)	100 (100)
5.0	100 (0)	100 (0)	100 (0)	90 (100)
6.0	100 (0)	100 (0)	100 (0)	100 (50)

	- Rana sylvatica	100 per treatment	eggs within 24 hr of oviposition	Percent hatching success:	Aluminum toxicity dependent on pH. Egg mortality due to hydrogen ion stress was correlated with delay in time to hatch and reduction in egg size.	Clark and LaZerte, 1985

Nominal Al (ug/L)	Nominal pH		
	5.75	6.75	6.14
0	91	93	*81
10	89	95	*74
20	93	87	*67
50		92	*54
100		94	*47
200			*40

* indicates significant difference between treatments on either side of asterisk.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Aluminum (cont'd)	Bufo americanus	89-160 per treatment	eggs within 24 hr of oviposition	Percent hatching success:	Aluminum toxicity dependent on pH. Egg mortality due to hydrogen ion stress was correlated with delay in time to hatch and reduction in egg size.	Clark and LaZerte, 1985 (cont'd)

Nominal Al (ug/L)	Nominal pH		
	6.75	6.32	6.14
0	97		*75
			*
10	100		*56
20	94		*61
50	98		*53
100	99		*55
			*
200			34
0	90	84	*40
5		89	*41
		*	*
10		64	*18
20		69	*21
50	97	74	

* significant difference of treatments on either side of asterisk in that row or column.

	B. sylvatica B. americanus	50 per treatment	tadpoles	Survival not affected by exposure to 0-200 ug/L Al in combination with pH 6.14 - 5.75.	Tadpoles which had also been exposed during egg-stage survived as well as those which developed in egg-stage in non-acidic, low-aluminum water.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Aluminum (cont'd)	- Rana temporaria	12 per treatment	tadpoles	Growth depressed and metamorphosis delayed by 800 ug/L and several tadpoles died at forelog emergence. At 1600 ug/L small tadpoles died, large tadpoles metamorphosed at small size but without delay.	Tadpoles reared in given concentration Al to metamorphosis.	Cummins, 1986
Aminocarb (C)	- Rana clamitans	7 per treatment	larvae	Significant decrease in mean activity time recorded at 10 ppm but not at 5 ppm.	Temp. 21 ± 1° C. 24 hr acclimation.	Lyons et al., 1976
Ammonia (non-ionized)	- Xenopus laevis		embryos	Gastrulation is most resistant stage.		Cotta Ramusino, 1980
Aniline	Xenopus laevis	60 per treatment stage 10	tadpoles	All tadpoles dead within 10 sec and 2 days at 10,000 and 1000 ppm, respectively.	Exposure for 2 weeks.	Dumont, 1987
		10 per treatment		Anamnia exposed to 90 ppm died within 12 days; only those held at ≥10 ppm developed into toads. 1 ppm resulted in reduced body size in toads.	Exposure for 90 days.	
Arosurf 66-E_2 (monomolecular organic surface film)	- Hyla cinerea		tadpoles	Progressed normally from tadpole to adult in tank with 0.68 ml/m^2 added.		Hebber and Cochran, 1986
Atrazine (H) (Weedex A)	- Frog	25 per test	spawn	Spawn developed normally in various waters used except in water containing Weedex (0.05 g/L in 3 litres) or water containing mud from a contaminated pond.	Thought that Weedex had drained into a pond resulting in no hatching success for two years.	Hazelwood, 1969

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Bacillus thuringiensis (var. israelensis)	- Rana temporaria	10 per treatment	tadpoles	Exposure to 10 mg/L decreased weight, delayed metamorphosis by four days.			Lyophilized endotoxins from B.t.i. (28×10^{10} sp. /g) used.	Poulov, 1987a
Benzene	- Triturus cristatus		blood	Exposed for 1 minute to 250 ppm benzene in air:			Intensity of alterations was proportional to frequency of treatments. Onset of a modification was related to time interval following first treatment.	Gerovini and Seren, 1978
		35		Daily exposure for 43 days produced transitory alterations in hematological picture followed by a return to normal.				
		30		Twice weekly exposure for 99 days or alternate day exposure for 38 days produced same results as above but slower.				
				Alterations included increase in number of circulating white blood cells, decrease in red blood cells preceded by decrease in hemoglobin percentage and increase in size of red blood cells.				
Benzene Hexachloride (BHC)	- Rana pipiens		adults 65 g, 8.9 cm long	BHC (ppm)	pH of solution at 25°C	No of frogs dead at 30 days	Frogs placed in 200 cc of test solution. Frogs lethargic; made no attempt to escape handling.	Kaplan and Overpeck, 1966
		20		17.0	6.23	3		
		10		8.0	5.85	0		
		10		6.0	5.82	0		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Benzo(a)pyrene	- Rana pipiens		marrow cells	Three-fold increase in sister chromatid exchanges at 0.01 ug/ml in ambient water. Mitotic index reduced and cell cycling time lengthened in dose-dependent manner.		Goard and Soutter, 1986
	- Frog		eye lens epithelium	No micronuclei present in the epithelial cell cycle appeared unaffected in frogs immersed in ≤20.0 ug/ml for 1 week.	No evidence of genotoxicity.	Kung et al., 1987
	- Pleurodeles waltl	30 per dose	erythrocytes from larvae stage 53	Incidence of micronuclei in larvae exposed for 8 days markedly increased with dose up to 0.075 ppm, then more gradually reaching 150 per 1000 at 0.75 ppm, minimum dose which produced a significant increase was 0.01 ppm.	Exposure for <2 days did not result in increased incidence of micronuclei.	Grønfeld et al., 1986
Beryllium (N)	- Frogs		embryos	Development not interfered with until N/5000 to N/1000.		Dilling and Healey, 1926
Bidrin (OP)	- Frogs		sartorius muscle	Inhibition of membrane voltage response of muscle to carbamylcholine which was dependent upon concentration and temperature.		Dukin et al., 1978
Bioresmethrin (PY)	- Xenopus laevis		lateral- line nerve organ	5×10^{-6} M for 2-3 hr *in vivo* caused only weak repetitive activity.		Vijverberg et al., 1982
			peripheral nerves	5×10^{-6} M caused repetitive activity within first 2 hr of exposure.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium (N)	- Rana catesbeiana		abdominal skin	Decreased skin resistance upon epidermal application of 7 mM.		Arita et al., 1979
	- Rana catesbeiana		heart, cardiac nerves	Cd reduced acetylcholine release from cardiac nerve. 10 uM did not alter the compound action potential of nerve trunk or effect pacemaker activity of heart.	Cd may act on the cardiac nerve terminals where it suppresses the release of acetylcholine.	Hayashi and Takayama, 1978
	- Gastrophryne carolinensis	100	eggs	Percent mortality and teratogenesis at hatching (H) and 4 days posthatching (PH) of eggs exposed to sediment-bound cadmium:	Hardness - 200 mm as $CaCO_3$. pH - 7.5 - 8.0.	Birge et al., 1977

Cd added to sediment (ppm)	Concentration in sediment (ppm)	% mortality H	% mortality PH
0.1	1.14	27 (1)*	33
1.0	2.18	26 (9)	33
10.0	10.0	31 (7)	41
100.0	122.0	36 (11)	52

Percent mortality increased as frequency in experimental population/controls.

* Percentage of survivors bearing gross congenital deformities at hatching.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium (cont'd)	- Xenopus laevis	5 per treatment	lung	Histopathological lesions on lung; pneumonia found in animals from 0.3 ppm Cd^{+2}.		Canton and Slooff, 1982a
	- Bufo arenarum	60-120 per treatment	embryos 2-cell stage	In 0.50 - 4.00 ppm Cd^{+2} range high mortality, delayed development, significant alterations in the gastrulation and neurulation processes observed. In 0.03 - 0.25 ppm range lethality much lower, retarded growth, reduced body size, behavioural disorders, and malformations observed.	Exposure was continuous from 2-cell stage onward.	Perez-Coll et al., 1985
	- Xenopus laevis		embryos	Cannot tolerate >2 ppm; stages of organogenesis are most sensitive.		Cotta Ramusino, 1980
	- Rana temporaria		adults	Subcutaneous injections of 0.12 - 0.24 mg/100 g/day for 10 days did not cause death.		Vasil'eva et al., 1987
			skin epithelium	High external concentrations $CdCl_2$ (0.005%) inhibited activity of Na, K-ATPase in skin epithelium.		
	- Rana japonica R. nigromaculata Rhacophorus schlegelii	5 per treatment	adults	Injection (i.p.) of 0.225 mg Cd/kg body weight 11 times during 19 days induced metallothionein (MT): - induced MT consisted of a single isofore - induced MT was Cd, Cu-MT in contrast to native Cu-MT.	No effect on concentrations of 10 elements in the livers and kidneys except Cu.	Suzuki et al., 1986

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadium (cont'd)	- Bufo regularis	10 per group	adults female	A single i.m. injection of 6.2 mg Cd^{+2}/kg produced elevated serum activities of alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase, lactic dehydrogenase. Ca serum not effected; serum phosphorus, total protein, total bilirubin increased.	6.2 mg/kg = 96 hr LD50. 0.2 mmole/kg EDTA protected toads from mortality in up to 20 mg Cd^{+2}/kg.	Hiley et al., 1986a
	- Bufo regularis		liver, kidney, spleen, muscle, whole blood	Following a single injection of 6.2 mg Cd/kg: - Zn increased significantly in liver, kidney; decreased significantly in whole blood, no change in muscle, spleen - Mg increased in liver, spleen, whole blood; no change in kidney, muscle - Cu decreased in liver, muscle, whole blood but not in kidney, spleen - Ca increased in liver, muscle, spleen; decreased in whole blood; no change in kidney - Fe decreased in kidney, spleen, whole blood; no change in muscle; increased then decreased in liver.	Co-administration of 40 mg EDTA/Kg caused Zn, Cu to return to normal in most tissues and organs. EDTA therapy caused increase in Mg, Ca, Fe in most tissues and organs. 6.2 mg Cd/kg = 96 hr LD50.	Hiley et al., 1986d

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium (cont'd)	- Rana pipiens	100 per treatment	embryo-larval	Survival rates at hatching and 4 days posthatching were at or above 99% following exposure to Cd-enriched sediments (1.04 - 1074 ppm).	Concentrations in water were 1.0 - 76.5 ug/L.	Francis et al., 1984
	- Bufo regularis	20 per group	adults females, 25 ± 4.3 g	6.2 mg Cd^{+2}/kg i.m. injection caused decrease in erythrocyte count, haemoglobin content, haematocrit value, reaching minimum after 48 hr; then gradual increase after 96 hr.	6.2 mg/kg = 96 hr LD50. EDTA therapy caused immediate recovery.	Hiley et al., 1986c
	- Rana pipiens	100	embryos	Percent survival with treatment initiated at cleavage (Cl), Neurula (N) and Tail Bud (TB) stages:	Temp. 21.1 - 22.2°C.	Birge and Just, 1975b

Days	2.5 ppm	0.5 ppm			0.1 ppm		0 ppm		
	Cl	Cl	N	TB	Cl	N	Cl	N	TB
1	73	77	53	90	93	74	97	85	100
2	48	70	53	90	85	74	94	85	100
3	36	50	51	90	78	74	85	85	100
4	8	33	51	90	68	74	84	85	100
5	3	30	51	90	63	72	76	85	100
6	0	30	51	90	63	70	70	85	100
7		30	51	90	63	70	70	85	100

	- Rana catesbeiana		eye	12.5 uM Cd^{+2} depressed the amplitude of rod receptor potential in the retina by 90%, while leaving cone response unaffected. 5.0 uM decreased rod response 27%.	Effect reversible.	Fox and Sillman, 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium (cont'd)	- Rana nigromaculata		eggs, tadpoles	Exposure of eggs to 4 ppm Cd^{+2} resulted in partial reduction of primordial germ cells at the 9-12 mm body length stage; caused abnormalities in many embryos.	Treatment prior to the first cleavage.	Huh, 1978
	- Rana pipiens		epithelial membranes	10^{-3} M Cd^{+2} did not inhibit active sodium transport across isolated epithelial cell layer from frog skin or when applied to basal-lateral surface; inhibited transport across frog urinary bladder and large intestine.	Effects vary according to epithelium studied and pressure of potential carrier molecules.	Hillyard et al., 1979
	- Triturus pyrrhogaster		stomach mucous epithelial cells	10^{-4} M Cd^{+2} decreased membrane potential to 66% of the control value; effective membrane resistance and electrical coupling ratio not appreciably affected. 10^{-7} - 10^{-5} M did not affect electrical properties.	Action antagonized by cysteine and acetylpenicillamine.	Kanno et al., 1978
	- Notophthalmus viridescens	12 per group	adults		No mortality in controls. Cd caused an increase in frequency and severity of abnormal limb regeneration.	Munson and O'Flaherty, 1978

Nominal Cd^{+2} concentration (ppm)	Mortality (%) on Day 51	Limb Regeneration
2.25	55	onset delayed
4.5	45	onset delayed
6.75	80	onset delayed - longer than groups above

	- Xenopus laevis		myelinated nerve fibres	Decreased permeability constant in K and Na systems; reversibly shifted Na activation curve in positive direction along potential axis.		Arhem, 1980

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium (cont'd)	- Rana catesbeiana		abdominal skin	Activity of ouabain - sensitive, ATPase completely inhibited by 1 mM Cd^{+2}. From 10^{-7}-10^{-3} M larger decreases found in activities of ouabain - sensitive ATPase and ouabain - insensitive ATPase as Cd concentration increased.	Uptake of Cd into whole skin was about 150 uM/kg wet weight with 2 mM treatment for 20 mins.	Takada and Hayashi, 1978a
	- Rana catesbeiana		abdominal skin	2 mM epidermally applied for 20 minutes caused 41.3% increase in short circuit current; no change in SCC when 2 mM dermally applied.		Takada and Hayashi, 1978b
	- Rana catesbeiana		abdominal skin	1 mM increased short circuit current to 126%. Cd^{+2} acted uncompetitively on the binding reaction of Na^{+} with a Na^{+} entry channel.	No interaction between Cd^{+2} and ouabain effects on Na transport.	Takada and Hayashi, 1981a,b
	- Xenopus laevis		embryos	Hg (ppm) / Cd (ppm): 10: 200 D, 20 D, 2 D, .2 D, 0 D; 1: 200 M-SL 94, 20 M, 2 S 30, .2 D, 0 D 0; .5: 200 M 88, 20 M, 2 M-S 100, .2 S, 0 D 0; .1: 200 N 95, 20 N, 2 N 93, .2 M 96, 0 S 87; .01: 200 N 93, 20 N, 2 N 90, .2 SL 91, 0 S 91; .001: 200 N 94, 20 N, 2 N 90, .2 SL 91, 0 S 91. D - Death; S - Severely deformed; M - Moderately deformed; SL - Slight effects; N - No effects; numbers = % survival from blastula to feeding stage.	Exposure for 4 days at 18°C.	Miller and Landesman, 1978

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cadmium Chloride (see also Cadmium, Zinc sulphate)	- Rana tigerina	10 per group	ovaries	Effect of 3 i.p. injections of 200 ug each at 10-day intervals: control: Wt. ovary (g)/100 g body wt. 14.2 ± 1.2; Wt. oviduct (g)/100 g body wt. 2.8 ± 0.12. $CdCl_2$: 10.2 ± 0.9; 1.8 ± 0.33. p<0.05; p<0.02	In Cd-treated frogs, ovaries contained large number of yolkey atretic follicles.	Pramoda and Saidapur, 1986
	- Xenopus laevis	20 per test	tadpoles stage 50-58	Mortality (%) of pre-treated (P) and non pretreated (C) tadpoles: Cd/Cd: 50 P 0 0 0; 50 C 55 75 80; 80 P 21 30 30; 80 C 84 100 100; 100 P 26 35 35; 100 C 90 100 100. Zn/Cd: 50 P 4 35 36; 50 C 13 40 44; 80 P 4 5 5; 80 C 35 80 81; 100 P 0 5 5; 100 C 48 70 77	Pretreatment with 5.0 mg/l zinc sulphate or 2.5 mg/l cadmium chloride for 96 hr.	Woodall et al., 1988
Carbaryl (C) (Karbaton 75)	- Rana temporaria	50-75 per treatment	tadpoles stage 25	Exposure to 0.1 - 0.05% caused death within 10-24 hr; short exposure (30 or 15 min) caused rapid frantic swimming. Exposure to .01% for 24 hr or .001% for 10 days caused tail malformations, increased mortality. Tadpoles increased activity for 2 hr followed by gradual immobilization.		Rzehak et al., 1977

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Carbaryl (cont'd)	- Xenopus laevis	50-75 per treatment	tadpoles stage 19-20	Exposure to 0.1% for up to 2 hr caused contraction and separation of muscle fibres. Exposure to 0.05% for 4 hr or 0.01% for 10 hr produced bent tails, retarded growth; after 24 hr 0.01% group appeared moribund and heart beat was slow - 80% died in first week and remainder died in second week. Exposure to 0.0001% for 8 days produced bent tails, oedema, inhibited growth and development. Mortality was 20% in first week, 65% in second week.		
	- Rana tigrina	6 per group	adults 80 ± 5 g	Hepatic vitamin A storage significantly lowered and serum vitamin A levels elevated at 0.006% but not at 0.004%.	Starved for 24 hr before exposure.	Keshavan and Deshmukh, 1984
	- Rana pipiens		abdominal skin	2×10^{-4} M produced significant increases in the short-circuit current and decreases in the resistance.	Likely that the sodium permeability of the outer barrier was altered.	Webb et al., 1979
	- Rana tigrina	4-7 per treatment	tadpoles	Rates of feeding, defecation, excretion increased with increasing concentrations (0-5 mg/L); conversion rate decreased leading to production of small froglets. No mortality until appearance of forelimbs; during metamorphosis mortality was significant at 2 mg/L.	Older and larger tadpoles were more resistant.	Marian et al., 1983
	- Rana tigrina		pancreas serum	Exposure of frogs to 0.002-0.006% had no effect on amylase or lipase activities of either tissue.		Deshmukh and Keshavan, 1987
Carbofuran (C)	- Rana pipiens		nerve-muscle	Muscle contractions evoked by nerve or muscle stimulation were suppressed by 1×10^{-5} to 1×10^{-3} M. Response to indirect nerve stimulation was suppressed more effectively than response to direct muscle stimulation by 2.5×10^{-4} M.	Temp. 22°C.	Takano et al., 1977

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Chlorani (H,F)	- Xenopus laevis	30	embryos tail bud stage	Concentration (ppm)		24 hr Survival	Surviving embryos (1.25 - 1.75 ppm) showed alterations in the development of the otolith, optic cup and pigmentation. Movement was sporadically convulsive.	Anderson and Prahlad, 1976
				control		100%		
				0.50		100%		
				0.75		95%		
				1.00		90-95%		
				1.25		90%		
				1.50		65%		
				1.75		40%		
				2.00		5%		
				* taken from mortality curve.				
Chlordane (OC)	- Rana pipiens		adults 45 g, 8.9 cm long	Chlordane (ppm)	pH of solution at 25°C	No. of frogs dead at 10 days	Frogs placed in 200 cc of test solution. Neuromuscular changes produced; excessive thrashing and abnormal reactivity to stimulation were observed.	Kaplan and Overpeck, 1964
		20		0.50	5.79	8		
		10		0.38	5.75	0		
		10		0.25	5.78	0		
	- Bufo arenarum		embryos	5 and 15 ppm produced 100% mortality on day 20 and 14, respectively. 1 ppm was not toxic.			Exposed from fertilization, medium replaced daily.	Juarez and Guzman, 1984b
Chlordane (α- and γ-)	- Rana pipiens		abdominal skin	Produced no significant changes in short circuit current or resistance at 2×10^{-4} M.				Webb et al., 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Chlordimeform	- Rana nigromaculata		rectus abdominis muscle	Chlordimeform and its metabolites caused a slow contraction at 10^{-4} - 10^{-3} M. Chlordimeform and 1 metabolite strongly inhibited the acetylcholine-induced contraction of the muscle whereas 2 other metabolites were inactive to the ACh-induced contraction at 10^{-3} M.		Watanabe et al., 1976
			adults 20 g	I.p. injection of 300 mg/kg chlordimeform produced 80% mortality within 24 hr.		
	- Rana pipiens		sciatic nerve - sartorius muscle	0.1 mM suppressed amplitude of spontaneous miniature end-plate potentials (45% of controls after 22-25 min) without changing their frequency. 1 mM blocked miniature end-plate potentials completely. Resting membrane potential not affected.	Concluded that the block of neuromuscular transmission by chlordimeform is due primarily to a depression of the end-plate sensitivity to the transmitter.	Wang et al., 1975
	- Rana catesbeiana R. nigromaculata		rectus-abdominis muscle	10^{-3} M caused a slow contraction and 10^{-5} - 10^{-3} M inhibited the acetylcholine-induced contraction in a noncompetitive manner. In R. catesbeiana K^+-induced contraction was also inhibited in a noncompetitive manner.	Effect probably due to depression not only of sensitivity of endplate but also of excitability of cell membrane.	Watanabe et al., 1975

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
p-Chloroaniline	- Xenopus laevis	40 per treatment	embryos	All embryos in 100 ppm died within 3 weeks. Development in controls and $\leq$ 0.1 ppm was complete after 13 weeks; at this time 85% and 20% of animals held at 1 and 10 ppm, respectively, had completed larval development.	Exposure for 90 days.	Dumont, 1987
Chlorpyrifos (OP)	- Hyla regilla		tadpoles 3 weeks old	Thermal tolerance significantly lowered:	24 hr exposure. Recommended dosage rate for mosquito control in California-90 ppb.	Johnson, 1980
				Conc. (ppb) / Onset of Spasms (°C): Mean / Range Control: 37.5 / 36.9-37.8 25: 35.3 / 33.4-36.9 50: 34.1 / 30.8-36.2		
Chlorpyrifos-ethyl (OP)	- Bufo boreas	20	juveniles (X snout-vent length 19.8 mm)	Exposure of hydrated toads to 30 ppb significantly lowered temperature tolerance.	23-24°C; 24 hr exposure.	Johnson and Prine, 1976
Chlorpyrifos-methyl (OP)	- Bufo boreas	20	juveniles (X snout-vent length 19.8 mm)	Exposure of hydrated toads to 30 ppb significantly lowered temperature tolerance.	23-24°C; 24 hr exposure.	Johnson and Prine, 1976
Chromium (VI) (M)	- Rana tigrina	80 per treatment	tadpoles	All died within 72 hours at $\geq$ 2 ppm (lowest concentration tested). Abnormalities observed in pigmentation, tail fin and alimentary canal.	Tadpoles intolerant to levels permitted in irrigation water.	Abbasi and Soni, 1984
Cismethrin (PY)	- Xenopus laevis		lateral-line sense organ	$1-3 \times 10^{-6}$ M for 1-3 hr caused repetitive activity.	Temp. 8-22°C. Number of impulses per train and train duration increased with cooling.	Vijverberg et al., 1982
			peripheral nerves	5×10^{-6} M caused repetitive activity within first 2 hr of exposure.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cobalt (Co^{+2}) (M)	- Triturus pyrrhogaster		stomach mucous epithelial cells	Concentrations of 10^{-7} M to 10^{-4} M did not affect the electrical properties of the cells.		Kanno et al., 1978
	- Xenopus laevis		myelinated nerve fibre	Decreased permeability constant in K and Na systems; reversibly shifted Na activation curve in positive direction along potential axis.		Arhem, 1980
Copper (M)	- Frogs		embryos	In M/500,000, few tadpoles survived many days after leaving the spawn.		Dilling and Healey, 1926
	- Triturus pyrrhogaster		stomach mucous epithelial cells	10^{-4} M Cu^{+2} decreased membrane potential to 67% of the control value. Effect of ion increased as concentration increased from 10^{-7} to 10^{-4} M. Slight changes in the effective membrane resistance and electrical coupling ratio.	Action antagonized by cysteine and acetylpenicillamine.	Kanno et al., 1978
	- Bufo boreas		larvae	3.7 mg/L Cu^{+2} - all died within 12 hours. 0.02 mg/L - all metamorphosed.	Up to 61 days exposure.	Porter and Hakanson, 1976
	- Xenopus laevis		myelinated nerve fibre	Slowed down the kinetics of the K system; decreased permeability constant in K and Na systems; reversibly shifted Na activation curve in a positive direction along potential axis.		Arhem, 1980

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS				REMARKS	REFERENCE
Copper (cont'd)	- Rana pipiens		rectus abdominus muscle	10^{-6} M Cu had little effect; 10^{-6} M Cu + 10^{-5} ACh caused larger contractions than ACh alone. Exposure to Cu + ACh for 10-15 min resulted in spontaneous spasmodic contractions.					Miller and Mackay, 1983
Copper Chloride (Miedzan 50)	- Rana temporaria	500 per treatment	tadpoles just after hatching; after completion of operculum	Effect of Miedzan 50 on tadpoles:				Static tests.	Jordan et al., 1977
				Conc. (suspension)	Exposure (days)	Age	Result		
				0.05%	3	Young	Inhibition of growth, accumulation of pigment in liver and stomach cells. Most died; after transferred to tap water, some of survivors returned to normal.		
				0.05%	5	Young	Stoppage of growth, malformations.		
				0.01% and 0.05%	3 and 5	Old	Inhibition of growth usually leading to death. After transferred to tap water all surviving tadpoles died within 3 days.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS				REMARKS	REFERENCE
				Conc. %	pH	No. of frogs surviving			
						6 days	30 days		
Copper Sulfate	- Rana pipiens	14		0.1	4.9	0	0	Frogs in high concentrations increased their cutaneous output of mucus.	Kaplan and Yoh, 1961
		14		0.025	5.2	0	0		
		11		0.005	5.5	3	0	Temp. 20°C.	
		12		0.0025	5.6	2	0		
		17		0.0015	5.6	17	10		
		8		0.001	5.3	8	8		
		8		0.0005	5.6	8	8		
		8		0.0003	5.7	8	8		
		8		0.0001	5.6	8	8		
	- Rana pipiens		eggs, newly-hatched tadpoles	Fertilized eggs were not affected by 0.04-1.14 mg/L. 0.31 mg/L or more was fatal to tadpoles. Weights of tadpoles grown in 0.06 or 0.16 mg/L were lower than controls.					Londé and Guiteau, 1973
	- Rana cyanophlyctis	15 per treatment	adults	Progressive reduction in total number blood cells following i.m. injections of 100 and 200 ug $CuSO_4$ for 40 days. White blood cells increased for first 7 days then gradually decreased. Haemoglobin content reduced.				Frogs sacrificed at end of 7th, 14th, 30th, 40th day.	Patil and Shivaraj, 1984

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS		REMARKS	REFERENCE
Crude Oil	- Rana arvalis		embryos	oil (ml/L)	% embryonic mortality	Skeleton deformities and edema of abdominal cavity noted in hatched larvae.	Prostolova and Danilova, 1986
		363		control	10.5		
		244		0.25	29.7**		
		233		0.50	33.9*		
		222		0.75	69.2*		
		240		1.00	60.8*		
				* all hatched larvae died within a few days. ** 60% of hatched larvae died within first week.			
		40 per treatment	larvae just post-hatch	All exposed to 0.25 and 0.5 ml/L died within 20 days; 35% and 73%, respectively, had anomalies. Reduction in growth noted at 0.15 ml/L.			
		30 per treatment	larvae rear limb buds present	All exposed to 0.5 ml/L died within 22 days; those in 0.05 ml/L delayed metamorphosis and anomalies found in juveniles.			
Cyanazyn (A)	- Rana temporaria	20 per treatment	tadpoles weight 40 ± 5 mg hind limb buds present	No effect at 0.2 or 2 ppm pure cyanazyn for 24 hr; at 20 ppm behaviour changed: lethargic, displayed spasmodic twitching and a tendency to remain at the water surface, did not feed. Exposure to 20 or 200 ppm as slow release pellets led to behaviour similar to above: more active than controls for first 5 hr.			Scorgie and Cooke, 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Cypermethrin (PY)	- Xenopus laevis		lateral-line nerve organ	5×10^{-6} M induced long-lasting trains of nerve impulses.		Vijverberg et al., 1982
			sciatic nerves	Up to 10^{-5} M for >24 hr did not cause repetitive activity after a single stimulus.		
2,4-D (H)	- Rana temporaria		tadpoles hind limb paddles or hind legs	No mortality or behavioural changes at concentrations up to 10 ppm for 48 hr.		Cooke, 1972
	- Rana temporaria		tadpoles	1 mg/L of the sodium salt and 2 mg/L of the diethylamine salt significantly inhibited metamorphosis. An addition to thyroxin in a conc. of 1-5 mg/L hinders the action of the hormone stimulating metamorphosis.	24 hr exposure.	Buslovich and Borushko, 1976
DDD (DL)	- Rana pipiens		abdominal skin	2×10^{-4} M caused a decrease in skin resistance and an increase in the short-circuit current.	Suggests that DDD increased the sodium permeability of the outer barrier.	Webb et al., 1979
DDT (DC) (see also metabolite)	- Frog		single muscle fibres	Membrane hyperpolarization reduced and delayed in K^+-free medium in presence of 5.10^{-6}.	Action pH dependent.	Creveun and Agroscreaus, 1978

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
DDT (cont'd)	- Rana ridibunda		sartorius muscle single fibres	5.10^{-4} prevented Ca^{+2} binding in the globular phospholipidic micelle and facilitated Ca^{+2} removal from the laminar phospholipidic micelle.		Creveun et al., 1981
	- Rana pipiens	50 - Controls; 50 - Exp.		Injection with 0.2 ml DDT dissolved in alcohol to 100 ppm based on body weight caused increased activity (violent leg extensions) after 4 hr; increase maintained until death.	Controls injected with 0.2 ml ethyl alcohol. Controls and experimental frogs exhibited low activity levels for first 4 hr.	Isaacson, 1968
	- Xenopus laevis		lateral-line organ	Marked repetitive activity after exposure to 2-5 ppm for 18 hr.	Negative temperature coefficient of activity. Temp. 19-21°C.	Akkermans et al., 1975a
			cutaneous touch receptors	Showed short trains of spikes in response to a single mechanical stimulus after treatment with 10^{-5} M for 30-90 min.	Slight increase in number of repetitive spikes when temperature lowered from 19-21°C to 10-13°C.	
	- Xenopus laevis		myelinated nerve fibre from sciatic nerve	4×10^{-5} M induced a medium tail current which was similar to that induced by exposure to allethrin.	Temp. 15 ± 1°C. Suggests that allethrin and DDT share common mechanism of action.	van den Bercken and Vijverberg, 1980

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
DDT (Cont'd)	- Bufo arenarum	10 expt.; 10 control	tadpoles 31 days	Activity of beta-glucuronidase in tail fin treated with 1 ppm DDT (mean ± SE, umol/g/min): Treated = 0.03694 ± 0.00299 Ctrl = 0.00465 ± 0.00063. Plasma levels of thyroid hormones similar in 2 groups.	Exposed from fertilization to 31 days, medium replaced daily. Activity in treated tissue significantly greater than that in controls.	Juarez and Guzman, 1986
	- Bufo arenarum		embryos	5 and 15 ppm produced 100% mortality on day 16 and 12, respectively. 1 ppm reduced time to metamorphosis.	Exposed from fertilization, medium replaced daily. Room temp.	Juarez and Guzman, 1986b
	- Rana tigrina	6 per group	adults 80 ± 5 g	Exposure to 0.1; 0.2 and 0.3% solutions caused decreased vitamin A stores in liver; greater decrease with increase in time of exposure (24-96 hr). Increased serum vitamin A levels with increased DDT noted.	Starved for 24 hr before exposure.	Keshavan and Deshmukh, 1984

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS				REMARKS	REFERENCE
DDT (cont'd)	-			Tail regeneration (mm; mean ± S.E.):					Weis, 1975
				Exposure (days)	Controls	5 ppb	25 ppb		
	Rana pipiens	20 per group	tadpoles at limb bud stage	5	1.3 ± .07	1.2 ± .09	0.9 ± .06*		
				8	2.7 ± .21	1.8 ± .08*	1.4 ± .10* (40%)**		
				12	4.6 ± .3	2.6 ± .1 * (30%)	2.5 (95%)		
	R. catesbeiana	10 per group	tadpoles at limb bud stage	5	1.1 ± .07	1.0 ± .05	1.0 ± .06		
				7	2.9 ± .16	2.8 ± .16 (30%)	2.3 ± .12* (30%)		
				9	4.5 ± .21	4.5 ± .22 (30%)	3.8 ± .09* (30%)		
				* Significantly different from controls. ** % mortality.					
	- Rana hexadactyla		gastrocnemius muscle	Soaking in 25-80 ppm for 10 min caused significant decrease in shortening length, tetanic duration, half contraction time, half relaxation time.					Rajendra et al., 1980

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
ODT (cont'd)	- Rana temporaria		spawn	Fresh spawn exposed to 0.5 nominal ppm for 24 hr 5 days prior to hatch were hyperactive 8-13 days after hatching. Development and weight gain were retarded. Exposure to 0.5 ppm when eye beginning to pigment or when head and body of each tadpole distinguishable had no effect.		Cooke, 1972
	Rana temporaria Bufo bufo		tadpoles	Tadpoles most susceptible either just before or just after developing hind limb buds and at these and later stages they became hyperactive when tissues contained 2-4 ppm. Toads were more resistant than frogs at all developmental stages. Some toads survived despite tissue concentrations >1000 ppm. Abnormal growth noted in frogs treated with 0.02 or 0.1 ppm for 48 hr.	During tail resorption small frogs susceptible to tissue residues as ODT released from the fat.	
	Triturus vulgaris		tadpoles snout-anus 10-12 mm	Conc. (nominal ppm) / Exposure (hr) / Mortality % / Behaviour 0 / 24 / 0 / normal 0 / 48 / 0 / normal 0.025 / 24 / 0 / normal/frantic 0.025 / 48 / 0 / frantic 0.5 / 24 / 10 / moribund 0.5 / 48 / 33 / moribund		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
ODT (cont'd)	- Rana arvalicus	90 per treatment, except 30 for stage 28 embryos	embryos	No abnormal effects on morphology or hatching success of embryos at stages 11, 16 or 18 exposed to 0.025 ppm for 24 hrs; signs of ODT poisoning observed in embryos treated at stage 28.		Licht, 1981
	- Rana temporaria	160	tadpoles weight 363 ± 11 mg, snout-anus 10.5 - 12.5 mm	29% of frogs treated with 0.1 ppm ODT for 48 hr developed snout abnormalities; 7% died. Stages of poisoning: I - skin has rough appearance; II - appearance of hole in centre of snout; III - loss of mandible and subsequent metamorphosis halted.	All affected individuals that reached tail resorption stage died. 2 factors caused snout abnormality: - disruptive effect on development of skin glands above upper mandible - lower mandible striking inner surface above upper due to hyperactivity.	Osborn et al., 1981
	- Frogs, toads			6 of 13 animals which ate insects poisoned by 10 % ODT spray died.		Laqueur, 1949
	- Rana temporaria	24	adults weight 13-31 g, snout-urostyle 36-47 mm	Frogs subcutaneously injected exhibited sluggishness, irritability, extension and trembling of hind limbs, abnormal posturing, colour change and an increase in croaking. Treated frogs did not hide as often as controls. No significant changes were noted in food consumption.	Those poisoning symptoms may increase predation in the field.	Cooke, 1974

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
DDT (cont'd)	- Rana temporaria Triturus cristatus			Treated tadpoles became hyperactive. Newts made significantly more lunges at hyperactive tadpoles than at controls. Accumulation of DDT in newts had no visible effect.	Possibly more predation of tadpoles by newts in contaminated areas due to altered behaviour of frogs. Newts may be more resistant than frogs.	Cooke, 1971
	- Rana temporaria	160 large; 160 small	tadpoles with external gills Y weight (mg): large - 6.89 ± 44 small 363 ± 11	Exposure to 0.1 ppm for 2 days: - no toxic effects in large tadpoles. - hyperactivity, tendency to float near surface, deformities in small tadpoles	Small tadpoles had higher residues.	Cooke, 1979
	- Rana tigrina		pancreas	Amylase activity decreased significantly in frogs exposed to 0.2 and 0.3 % for 24-96 hrs; no effect at 0.1% exposure. Lipase activity increased significantly in frogs exposed to 0.1, 0.2, 0.3% for 24-96 hr.		Deshmukh and Keshaveen, 1987
			serum	Amylase activity increased significantly in frogs exposed to 0.2, 0.3 % for 24-96 hrs; no effect at 0.1% exposure. No effect on lipase activity at any concentration.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS		REMARKS	REFERENCE
DDT (cont'd)	- Rana temporaria		tadpoles	Mortality of tadpoles exposed to various concentrations of DDT:			Cooke, 1970
				Nominal Concentration (ppm)	Mortality (%)		
		40 per group	hind limb bud stages; snout-anus- 6.5 - 9.5 mm	0.0 0.01 0.1 1.0 10.0	<3 0 0 20 70	Temp. 20 ± 5°C. Exposure - 1 hr. Length of expt. - 14 days.	
		40 80 80	small-medium hind legs; snout-anus- 10-12 mm	0.0 1.0 10.0	<3 10 50	Temp. 23 ± 5°C. Exposure - 1 hr. Length of expt. - 22 days.	
	- Rana pipiens		abdominal skin	2×10^{-4} M did not produce changes in short circuit current or resistance.			Webb et al., 1979
	- Frogs		adults	All frogs treated with 150 mg/kg dissolved in olive oil and injected into the dorsal lymph sac died in 6-72 hr. Some died following injection of 10 mg/kg.			Ellis et al., 1944
DDVP (DP) (phosphoric acid 2,2 - dichlorovinyl dimethyl ester)	- Frog		ventricle muscle	Inhibition of membrane voltage response of muscle to carbamylcholine which was dependent upon concentration and temperature.			Dukes et al., 1978

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Defenuron (H)	- Rana temporaria	20 control; 20 expt.	tadpoles 2 days old	Delayed development found in tadpoles kept in 100 ppm for 80 days: controls - developed forelegs at 30 days, metamorphosed by 40 days. experimental - developed forelegs at 30 days, only 40% underwent complete metamorphosis, 40% developed into giant tadpoles without forelegs, 20% died.		Paulov, 1977
DEHP (Di-2-ethylhexyl-phthalate)	- Rana arvalis	Approx. 100 eggs per treatment	2-3 day old embryos	Hatching success decreased with increasing levels DEHP; approximately 50% hatched when exposed to sediments with 130 ug/g fresh weight. Survival rates of tadpoles did not differ.	DEHP added to sediment of model system. No abnormalities in development, no delays in hatching observed.	Larsson and Thurén, 1987
Deltamethrin (PY)	- Xenopus laevis		lateral-line nerve organ	Produced long trains of impulses in vitro at $1-5 \times 10^{-6}$ M for 3 hr.		Vijverberg et al., 1982
			sciatic nerves	Up to 10^{-5} M for more than 24 hr did not induce repetitive activity after a single stimulus.		
	- Rana esculenta		skin	Under open circuit conditions, 10^{-6} M did not provoke changes in Na^+ fluxes; at 10^{-5} M a slight inhibition of the J(o)Na$^+$ after 30 min noted, no change in Cl^- fluxes.		Salzbrun, 1983
	- Xenopus laevis		myelinated nerve fibres	10^{-6} M caused a frequency dependent depression of the action potential which was associated with a progressive membrane depolarisation brought about by summation of depolarising after-potentials.	Specifically affects the sodium channels of the nerve membrane.	Vijverberg and van den Bercken, 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
DFP (DP) (diisopropyl fluorophosphate)	- Bufo viridis	3	adults blood nervous tissue	Residual cholinesterase at time of death following injection of 2 LD50 into dorsal lymphatic sac: 20% 44%	LD50 - 1440 ppm.	Edery and Schatzberg-Porath, 1960
Dichlofenthion (DP)	- Rana pipiens		nerve-muscle	Only a small suppressive effect on muscle contraction evoked by direct or indirect stimulation at 1×10^{-3} M. No effect at lower concentrations.	Temp. 22°C.	Iskono et al, 1977
Dichlone (F,H)	- Xenopus laevis	30	embryos tail-bud stage	Concentration (ppm) / 24 hr Survival: control 100% 0.075 100% 0.1 95% 0.125 10-55% 0.15 15-20% >0.2 0%	Overall inhibition of growth and development; 0.1 - 0.15 ppm disrupted development of cephalic end of embryo.	Andersen and Pranlad, 1976
Dicrotophos (OP)	- Rana catesbeiana		tadpoles	* taken from mortality curve All tadpoles exposed to 5 ppm survived.	Continuous flow apparatus for 96 hr. Brain cholinesterase activity of Mallards fed dosed tadpoles was 78.1% of that of controls. None died.	Hall and Kolbe, 1980
Dieldrin (HEOD) (OC)	- Rana pipiens		adults 45 g, 8.9 cm long	Dieldrin (ppm) / pH of solution at 25° C / No. of frogs dead at 30 days: 20 — 0.10 — 5.77 — 10 10 — 0.08 — 5.61 — 0 10 — 0.05 — 5.59 — 0	Frogs placed in 200 cc of test solution. Neuromuscular changes produced; excessive thrashing and abnormal reactivity to stimulation were observed.	Kaplan and Overpeck, 1968

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Dieldrin (cont'd)	- Rana esculenta R. pipiens R. temporaria		motor end-plate	10^{-5} to 10^{-4} M caused no significant effects on frequency and amplitude of miniature end-plate potentials, or on end-plate membrane potential.		Akaike et al., 1974
	- Xenopus laevis		spinal chord	Application of up to 5×10^{-5} M *in vitro* failed to produce any significant effects.	Temp. 18 ± 2°C.	Akaike et al., 1975c
	- Rana temporaria		tadpoles	Treatment with 0.5 nominal ppm produced 9% mortality and frantic/enraged behaviour after 24 hr, 47% mortality after 48 hr. Snout abnormalities noted. 0.02 ppm had no effect.		Cooke, 1972
	Bufo bufo		tadpoles	Exposure to 0.5 ppm for 48 hr caused no mortality; enraged behaviour and snout abnormalities noted.		
	- Lymnodynastes tasmaniensis	Approx. 150 eggs per treatment	embryos	Skeith, also capsule and cephalic pigmentation abnormalities after exposure to 0.1 ppm for 7 hours; accelerated growth relative to controls. No effects with exposure to 0.01 ppm.	Observed abnormalities resemble those described for tadpoles exposed to chloramsi.	Brooks, 1981
	- Rana pipiens		abdominal skin	2×10^{-4} M caused significant increases in the short circuit current while decreasing the resistance.	Suggests altered the sodium permeability of the outer barrier.	Hobe et al., 1979
	- Xenopus laevis	1974	lateral-line organ	Treatment with 5×10^{-4} M for several hr produced no significant effect. Preparations taken from animals showing severe symptoms of poisoning after exposure to 3-5 ppm dieldrin did not show any sign of repetitivity.		Akaike et al., 1975a
			cutaneous touch receptors	No sign of repetitive activity. No effects of temperature changes noted.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS				REMARKS	REFERENCE
Dieldrin (cont'd)	- Bufo arenarum		embryos	Embryos developed in 0.2 and 2.0 mg/L died between 20-23 days; 25-30 % of those developed in 0.02 mg/L had front legs earlier than controls.					de Llamas et al., 1985
		3 per treatment		Treatment	Time Exposed	% Inhibition ACHE	% Inhibition CHE	All determinations made on 11 day embryos. Values are mean ± SE.	
				Unfertilized oocyte + 2 mg/L	1 hr	19.7 ± 1.8	17.2 ± 1.7		
				"	4 hr	22.3 ± 1.4			
				Recently fertilized oocyte + 2 mg/L	4 hr	22.7 ± 1.2	25.9 ± 1.3		
				"	continuous	26.7 ± 1.4	25.5 ± 1.8		
Diamfos (DP)	- Bufo viridis	3	adults	Monobasal cholinesterase at time of death following injection of 2 LD50 into dorsal lymphatic sac:				LD50 = 1410 ppm.	Edery and Schatzberg-Porath, 1960
			blood	0%					
			spinal nervous tissue	31%					

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS						REMARKS	REFERENCE
Dimethoate (OP)	- Rana tigerina		eggs tadpoles pre-limb, 29 mm		Development of Eggs			Dvpt. of Tadpoles		Temp. 30-38°C.	Dutta and Mohanty - Majumdi, 1978
				Conc. (%)	Time to metamorphosis (days)	Av. length at metamorphosis (mm)	No. Died	Time to metamorphosis (days)	Av. length at metamorphosis (mm)	Recommended dosage for spraying is .04%, higher than concentrations causing development delays.	
				Ctrl	39	16.8±0.96	0	20	16.0±0.74		
				0.0001	46	16.8±0.63	0	61	16.0±0.74		
				0.0002	54	16.0±0.82	0	64	16.0±0.54		
				0.0003	62	16.8±0.71	3	64	16.0±0.74		
				0.0004	62	16.8±0.74	3	64	15.0±0.54		
				0.0005	68	15.0±0.52	4	64	15.0±0.74		
				0.0006	63	14.5±0.52	4	64	15.0±0.54		
				0.0007	68	14.0±0.54	5	64	14.0±0.54		
				0.0008	63	13.5±0.57	6	67	14.0±0.74		
				0.0009	54	13.5±0.57	6	70	14.0±0.54		
				No eggs treated with 0.001 - 0.009% reached metamorphosis.							
	- Rufo melanostictus	60	tadpole tails	After 7 days in 0.05 ppm melanophore size increased significantly, puncto-stellate melanophores were visible and developed dendritic processes and branching.							Pandey and Ismar, 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS				REMARKS	REFERENCE
Dimethoate (cont'd)	- Rana tigerina		eggs, limb bud stage tadpoles	Conc. %	% metamorphosed		Mean time for metamorphosis (days)	Temp. 30-38°C.	Mohanty-Majumdi and Dutta, 1981
					eggs	limb bud	eggs	limb bud	Mean length at metamorphosis reduced at 0.0008 - 0.009% (eggs) and 0.007 - 0.01% (tadpoles).
				0.01	--	80	--	59	
				0.009	40	100	54	70	
				0.008	40	100	63	67	
				0.007	50	100	60	64	
				0.006	60	100	65	64	
				0.005	60	100	60	64	
				0.004	70	100	62	64	
				0.003	70	100	62	64	
				0.002	100	100	54	64	
				0.001	100	100	46	64	
				Control	100	100	35	18	
Dimethylhydrazine (compound used in jet and rocket fuel)	- Xenopus laevis		embryos				conc. (mg/L)	% malformed	Kinked tail was most frequent malformation. Most larvae with kinked tails metamorphosed into normal frogs. Other malformations included microcephaly, failure of stomodeum and edema.	Greenhouse 1976b
		100		1,2-dimethylhydrazine			control	3	
		100					10	6	
		100					20	6	
		100					40	5	
		100					50	100	
		100					80	100	
		100		1,1-dimethylhydrazine			control	3	
		100					1	3	
		100					2	10	
		200					5	50	
		100					10	67	
		100					20	100	
				Susceptible during neurulation.					

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Dimethylhydrazine (cont'd)	- Xenopus laevis	9-69 per treatment	larvae	1 mg/L 1,2 - DMH lethal to 100% larvae within 7 days; continuous contact with 0.1 mg/L allowed normal metamorphosis. Exposure to ≤1 mg/L 1,1 - DMH did not affect survival or metamorphosis; higher concentrations lethal.		Greenhouse 1976a
		50-100 per treatment	embryos	≤40 mg/L 1,2 - DMH not toxic if embryos transferred to uncontaminated water prior to hatching; ≥10 mg/L teratogenic. ≥ 5 mg/L 1,1 - DMH teratogenic.	Failure to elongate, tail kinks most common abnormalities.	
2,4-Dinitrophenol	- Rana temporaria		cutaneous pectoris nerve-muscle	10^{-4} M caused rise in miniature endplate potential frequency 30 min after application following reduction in first 20 min.		Statham et al., 1978
Dioctyldiphenylamine (component of jet fuel)	- Rana pipiens Xenopus laevis	200	embryos larvae	Up to 1 g/L not toxic to either species.	No deleterious effects on development. Controls had no malformed embryos.	Greenhouse, 1976a
Diquat (H)	- Rana pipiens	300 per treatment	eggs	After exposure to 100 ppm for 21 days eggs exhibited reduced viability and increased exogastrulation. None survived beyond 14 days post-hatch.	Temp. 18°C.	Bieber and Mitchell, 1978
	- Rana pipiens	200 per dose group	embryos: early gastrula and 15 days of age	% alive 16 days post-treatment after exposure at gastrula stage (G) or 15 days of age (15):	No abnormalities observed in embryos.	Dial and Dial, 1987

Diquat (mg/L)							
0		2	5	10	
G	15	G	G	G	15
81.8	90.0	79.5	70.9*	36.1*	78.3

* significantly different from controls.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS		REMARKS	REFERENCE
Diquat (cont'd)	- Xenopus laevis	30	embryos tail-bud stage	Concentration (ppm)	48 hr Survival*	0.75 - 2.0 ppm reduced body size and pigmentation; 1.5 ppm produced some distortion in body shape.	Anderson and Prahlad, 1976
				control	100%		
				0.5	95%		
				0.75	90-95%		
				1.0	70%		
				1.25	75%		
				1.5	70%		
				2.0	20%		
				* taken from mortality curve.			
Diquat/Rebem (H)	- Xenopus laevis	30	embryos tail-bud stage	0.75 - 1.25 ppm diquat/2.0 ppm rebem produced 100% mortality within 6 days.		Size of larvae reduced, melanin synthesis retarded, muscle fibres less developed.	Anderson and Prahlad, 1976
Distillery Effluent	- Rana catesbeiana	5 per treatment	tadpoles pre-hind limb emergence	Increase in concentration (0.03, 0.06, 0.12%) reduced period of limb bud emergence and tail resorption; elevated values for length of limbs, tail and body wt. of adults.			Hamilton et al., 1985
Edrophonium	- Rana pipiens		nerve-muscle	Neuromuscular transmission supressed at 5×10^{-4} and 1×10^{-3} M. Contraction evoked by muscle stimulation also suppressed. No effect at 1×10^{-4} M.		Temp. 22°C.	Tahone et al., 1977

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Endrin (OC)	- Rana catesbeiana	10	adults	Frogs force-fed mosquitofish which had been exposed to 2 ppm endrin solution for 7 days: - 100% mortality - Survival time 15.6 ± 5.03 (X̄ ± SD) - Ratio predator: prey mean weight 410:1.	Resistant mosquitofish tolerate endrin levels that will kill predators many times their own weight.	Reeseke and Ferguson, 1968
	- Rana pipiens		abdominal skin	2×10^{-4} M produced no significant effects on the short circuit current or resistance.		Nobb et al., 1979
	- Rana pipiens		adults 45 g, 8.9 cm long	Endrin (ppm) / pH of solution at 25°C / No of frogs dead at 30 days	Frogs placed in 200 cc of test solution.	Kaplan and Overpeck, 1966
		20		0.03 / 5.86 / 6	Neuromuscular changes produced: excessive thrashing and abnormal reactivity to stimulation were observed.	
		10		0.02 / 5.80 / 0		
		10		0.015 / 5.78 / 0		
Ethanol	- Rana temporaria	10 per series	tadpoles 8 days old	At 1000 ppm, all died within 10 min; at 900 ppm, tadpoles survived for up to 1 day; ~800 ppm had no effect.		Paulov, 1957b
		100	tadpoles	The body weight of tadpoles kept in 500 ppm from day 7 to day 22 increased compared to controls; onset of metamorphosis accelerated by up to 8 days; all died.		
		10	adults blood, muscles	6 day exposure to 1000 ppm led to enhanced enzymatic activities of aspartate aminotransferase and alanine aminotransferase.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Ethylenethiourea (degradation product of ethylenebis-dithiocarbamate fungicides)	- Microhyla ornata	50 per treatment	embryos gastrula stage	Conc. (ppm) / % dead at 96 hr / % abnormal at 96 hr	Abnormalities included wavy notochord, abnormal pigmentation and movement. Effects similar to those induced by dithiocarbamates.	Ghate, 1986
				0.0 / 0.0 / 0		
				5.0 / 0.0 / 22		
				10.0 / 2.0 / 100		
				20.0 / 10.0 / 100		
				30.0 / 10.0 / 100		
				40.0 / 90.0 / 100		
				50.0 / 90.0 / 100		
Ethylmethane-sulfonate	- Rana pipiens		marrow cells	7-fold increase in sister chromatid exchanges at 0.1 mg/ml in ambient water. Mitotic index reduced and cell cycling time lengthened in dose-dependent manner.		Gaard and Soutter, 1986
	- Frog		eye lens epithelium	Increase in pre-mitotic, G2, population and decrease in mitoses; micronucleated cells present in frogs smeared in 25-250 ug/L for 1 week.	Evidence of genotoxicity; suggests cell cycle arrested.	Kung et al., 1987
Eulan Wa New (mothproofing agent)	- Rana temporaria	20 per concentration (x 4)	tadpoles weight 21.0 ± 1.0 mg	Tadpoles exposed to 1.0 mg/L exhibited decreased movement for first 48 hr, slower development, abnormalities, decreased feeding rate and weight, 92.5% mortality. Tadpoles exposed to 0.1 mg/L showed slower development and no mortality.	Temp. range 11.5 - 15°C. Animals not fed during first 48 hr.	Osborn and French, 1981

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Fenitrothion (OP)	- Rana tigerina		tadpoles	Effects on feeding stage (F) and limb bud stage (L):	Temp. 30-38°C.	Mohanty-Hejmadi and Dutta, 1981
				Conc. % metamorphosed / mean time for metamorphosis (days): F: 0.00004 L: 20 F: L: 53.0; 0.00003 L: 40 L: 54.3; 0.00002 F: 40 L: 60 F: 51.0 L: 54.6; 0.00001 F: 40 L: 100 F: 56.5 L: 59.6; Control F: 80 L: 100 F: 31.5 L: 35.4	Mean length of metamorphosis reduced in limb bud tadpoles at 0.00004%.	
			eggs	Complete mortality at 0.0000001%.		
	- Rana clamitans	7 per treatment	larvae	Significant decrease in mean activity time recorded at 2 ppm but not at 1 ppm.	Temp. 21 ± 1°C. 24 hr acclimation.	Lyons et al., 1976
Fenoprop (H)	- Litoria ewingi; Limnodynastes tasmaniensis	20 per conc.	adults	No deaths observed in either species following exposure to 80-400 ppm for 96 hr.		Johnson, 1976
Fenpropathrin (PY)	- Xenopus laevis		lateral-line nerve organ	After 1 - 1.5 hr in vitro exposure to 5×10^{-6} M trains of impulses recorded with mean of 330 impulses per train.		Vijverberg et al., 1982
			sciatic nerves	Exposure to up to 10^{-5} M for more than 24 hr did not induce repetitive activity after a single stimulus.		
Fenthion (OP)	- Bufo boreas	20	juveniles (X snout-vent length = 19.8 mm)	Exposure of hydrated toads to 40 ppb significantly lowered temperature tolerance.	23-24°C; 24 hr exposure.	Johnson and Prine, 1976

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Fenthion (cont'd)	- Hyla regilla		tadpoles 3 weeks old	Thermal tolerance significantly lowered:	24 hr exposure. Recommended dosage rate for mosquito control in California - 100 ppb.	Johnson, 1980
				Conc. (ppb) / Onset of Spasms (°C): Mean / Range		
		34		Control: 37.5 / 36.9-37.8		
		42		25: 36.9 / 36.3-37.7		
		39		50: 36.7 / 36.2-37.9		
		43		100: 35.7 / 31.9-37.2		
	- Rana catesbeiana		tadpoles	All tadpoles exposed to 5 ppm survived. Bioconcentration factor: X 62.	Continuous flow apparatus for 96 hr. Brain cholinesterase activity of malloids fed dosed tadpoles was 16.9% of that of controls. All ducks died within 5 hr.	Hall and Kolbe, 1980
Fenvalerate (PY)	- Xenopus laevis		lateral-line nerve organ	Acted slowly; frogs had to be treated in vitro with 5×10^{-6} M for up to 23 hr before repetitive activity induced.		Vijverberg et al., 1982
			sciatic nerves	Up to 10^{-5} M for more than 24 hr did not induce repetitive activity after a single stimulus.		
Furanace (Fish bactericide)	- Rana pipiens		larvae	Exposure to 0.2 - 20 ppm for 96 hr produced no mortality after 7 days. Many in 10 and 20 ppm were immobilised during exposure but recovered.		Marking et al., 1977

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
HCE (OC) (dieldrin analogue)	- Rana pipiens		liver microsomes (adults)	Microsomal metabolism (apparent Vmax and Km values ± SE): App Vmax = 0.013 ± 0.001 nmol/min/mg App Km = 29 ± 8 nmol/mg.		Ronis and Walker, 1985
Heptachlor (OC)	- Rana pipiens		abdominal skin	2×10^{-4} M produced no significant effects on short circuit current or resistance.		Webb et al., 1979
	- Bufo arenarum		embryos	5 and 15 ppm produced 100% mortality on day 15 and 13, respectively. 1 ppm reduced time to metamorphosis.	Exposed from fertilization, medium replaced daily.	Juarez and Guzman, 1984b
Hexachlorophene (F) and other bisphenols	- Grass frogs		blood	1.2×10^{-4} M produced 50% hemolysis in washed, nucleated erythrocytes; 1.60×10^{-4} produced 50% hemolysis in nucleated erythrocytes in whole blood.	Temp. 22 ± 1°C.	Flores and Buhler, 1974
n-Hexane	- Bufo arenarum		embryos	5 and 15 ppm produced 100 % mortality on day 29 and 17, respectively. 1 ppm was not toxic.	Exposed from fertilization, medium replaced daily.	Juarez and Guzman, 1984b

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Hydrazine (component of jet fuel)	- Xenopus laevis	10-86 per treatment	larvae	Concentrations ≥1 mg/L produced irreversible toxic effects and death within 24-28 hr. 0.1 mg/L was not toxic to larvae.		Greenhouse, 1976a
		50-100 per treatment	embryos	Normal development in 5 mg/L but did not survive as larvae if not transferred to <1 mg/L at hatching. Malformations in some batches of eggs at 10 mg/L.		
	- Xenopus laevis		embryos	Concentration (mg/L) / % malformed	Kinked tail was most frequent malformation. Most larvae with kinked tails metamorphosed into normal frogs. Other malformations included tail duplication, microcephaly, failure of elongation and edema.	Greenhouse, 1976b
		100		Control: 2		
		100		1: 3		
		150		10: 35		
		100		25: 100		
		50		50: 100		
		150		100: 100		
				Susceptible during neurulation.		
Hydrazine Sulfate (compound used in jet and rocket fuel)	- Xenopus laevis		embryos	Concentration (mg/L) / % malformed	Kinked tail was most frequent malformation. Most larvae with kinked tails metamorphosed into normal frogs. Other malformations included microcephaly, failure of elongation and edema.	Greenhouse, 1976b
		100		Control: 0		
		100		10: 3		
		200		40: 100		
				Susceptible during neurulation.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Ionizing Radiation	- Assorted species/review		cells, tissues, organs, whole body	Effects include: suppressed mitotic activity of cells; formation of giant cells; formation of vacuoles; damage to nucleus; changes in skin epithelium and connective tissue; damage to muscle, spleen, CNS, liver, intestine, eyes; developmental abnormalities.		Brunet, 1965
	- Caudiverbera caudiverbera		larvae	Dose-effect relationship demonstrated between level of radiation (50-150 rads) and presence of micronucleated blood cells.	Exposure for 10 days.	Hermosilla and Carrasco, 1985
Iron	- Bufo boreas		larvae	144 mg/L - all died within 24 hr. 30 mg/L - all died within 20 days. 20 mg/L - all metamorphosed.	Greater than 39 days exposure to Fe^{+2}.	Porter and Hakanson, 1976
LAS (Linear chain alkylbenzene-sulphonate)	- Xenopus laevis		embryos	Caused malformations.	No levels given.	Cotta Ramusino, 1980
Lead	- Rana catesbeiana		eye	5.0 and 12.5 uM Pb^{+2} produced 9% and 20% decreases, respectively, in amplitude of rod response. No effect on cone potential.	Effect reversible.	Fox and Sillman, 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Lead (cont'd)	- Xenopus laevis		embryos	Mg (ppm) / Pb (ppm): 10 / 1 / .5 / .1 / .01 / .001 200: D / M / SL-M / M / M 0: 99 / 88 / 93 / 88 20: D / M-S / M / M / M / M 0: 81 / 100 / 78 / 96 / 96 2: D / M-S / M / M / M / M 0: 59 / 92 / 95 / 93 / 97 .2: D / S / M-S / M-S / SL / SL 0: 73 / 94 / 94 / 95 / 93 0: D / S / S / S / S / S 0: 59 / 100 / 74 / 96 / 82 D - Death; S - Severely deformed; M - Moderately deformed; SL - Slight effects; N - No effects; numbers = % survival from blastula to feeding stage.	Exposure for 6 days at 18°C.	Miller and Landesman, 1978
	- Rana nigromaculata		eggs, tadpoles	Exposure of eggs to 70 ppm resulted in partial reduction of primordial germ cells at the 9-12 mm body length stage. 70 ppm was lethal to tadpoles.	Treatment prior to the first cleavage.	Huh, 1978
Lead Acetate	- Rana catesbeiana	143	blood of tadpoles	25 ppm lead acetate or lead nitrate produced destruction of erythrocytes and necrosis of liver, spleen and intestinal mucosa; high mortality. Action of lead nitrate more rapid than action of lead acetate.		Barrett, 1967
	Xenopus laevis		embryos	Exposure to >1 mg/L always produced lethality and/or abnormalities. Main anomaly in embryos exposed before neurula stage was unfused neural tube; in those exposed after neurula stage main anomaly was edema in thorax and kidney tail.		Kanamura and Tanimura, 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Lead Acetate (cont'd)	- Rana hexadactyla		heart	Preperfusion by 10^{-7} M for 20 min augmented digoxin-induced cardiotoxicity: reduction in total digoxin exposure and time taken for systolic arrest.	Exposure to 6.5 mM Ca^{+2} attenuated this cardio-toxicity when prior to or simultaneously with Pb exposure.	Krishnamoorthy et al., 1987
	- Nocturus	13	blood	Newts maintained in bath of 20 cc of 1% lead acetate and 4 L water suffered destruction of mature erythrocytes which stimulated differentiation and proliferation of erythrocytes in blood.		Dawson, 1933
	- Frogs		eggs, tadpoles, adults	1-5 ppm can partly or fully inhibit germination of eggs. Adults killed at 100 ppm. 2 ppm was toxic to tadpoles.	Tadpoles with external gills were sensitive than those with internal gills.	Dilling and Healey, 1926
Lead Nitrate	- Frogs		eggs, tadpoles, adults	1-5 ppm can partly or fully inhibit germination of eggs. Adults killed at 100 ppm.	Tadpoles with external gills were sensitive than those with internal gills.	Dilling and Healey, 1926
				2 ppm was toxic to tadpoles.	Lead salts had higher inhibitory effects on germination of frog ovum than zinc, thorium, copper, beryllium or thallium.	
	- Rana esculenta		leg skin	10^{-5} M inhibited isosmotic, active transport-coupled volume flow.		Calentano et al., 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Lead Nitrate (cont'd)	- Rana pipiens	24 per group	adults 9 cm, 65 g	Total Concentration Pb (ppm)	No. of Frogs dead at 30 days		Temp. 8°C -sloughing of the integument -loss of postural tone -sluggishness -red and white cell counts decreased progressively as increasing lead concentrations -lead deposits in liver at 25 ppm.	Kaplan et al., 1967
				Control	0			
				25	1			
				50	3			
				100	7			
				150	17			
				200	22			
				300	24			
	- Rana catesbeiana	143	blood of tadpoles	25 ppm lead acetate or lead nitrate produced destruction of erythrocytes and necrosis of liver, spleen and intestinal mucosa; high mortality. Action of lead nitrate more rapid than action of lead acetate.				Barrett, 1967
Leptophos (OP)	- Rana pipiens		nerve-muscle	No effect on muscle contraction evoked by indirect or direct stimulation at up to 2.5×10^{-4} M.			Temp. 22°C.	Tokano et al., 1977
Leptophossan (OP)	- Hyla chrysoscelis		embryos	Conc. (ppm)	Mortality %	Abnormal Embryos (%)	Dermally exposed for 24 hr, no. survivors determined 120 hr after exposure.	Fulton and Chambers, 1985
		76		0	1	0		
		75		1.0	0	0		
		76		2.5	34	0		
		80		5.0	85	0		
		40		7.5	100	0		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Lindane (UC)	- Rana pipiens		ventral abdominal muscle	Initial rate of movement of lindane (10 ppm) through live or dead skin was 0.680 ppb/hr; at 0 hr, rates were 0.691 and 0.666 ppb/hr, respectively.		Kaiser and Duncan, 1972-73
	- Xenopus laevis		eggs	See table below	At 2 ppm 4 week old tadpoles showed up to 100% mortality.	Marchal-Segault and Ramade, 1981
	- Rana pipiens		cutaneous pectoris nerve-muscle	At 100 uM frequency of miniature end-plate potentials increased by 327 ± 24% (X ± S.E.) and amplitudes reduced to 21 ± 14% of controls after 30 min.	Suggest decreased amplitudes were reflection of decreased sensitivity of end-plate	Joy et al., 1987
			neuroblastoma cells	1-100 uM did not significantly modify action potentials elicited or affect calcium channels.	to ACh.	
	- Rana temporaria		cutaneous pectoris nerve-muscle	5×10^{-5} M caused marked increase in miniature end plate potential frequency.	Results suggest Ca^{+2} permeability and Ca^{+2} entry increased.	Publicover and Duncan, 1979

Trial	Control		Lindane		
	Water	Acetone	0.5 ppm	1 ppm	2 ppm
I No. eggs	30	30	30	30	30
No. eggs hatched	17	19	20	17	0
II No. eggs	30	15	45	45	45
No. eggs hatched	16	11	20	26	11
III No. eggs	75	75	75	75	75
No. eggs hatched	57	32	38	35	28
Total eggs	135	120	150	150	150
Total eggs hatched	70	62	78	76	67
Hatching rate (%)	52	52	52	51	31

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Lindane (cont'd)	- Rana temporaria		cutaneous pectoris muscle	5×10^{-3} M for 40 min caused extensive damage to myofilaments; 20 min exposure had little effect.	In the presence of Ca^{+2}, damage greater at 23°C than at 10°C.	Publicover et al., 1979
	- Rana pipiens		abdominal skin	2×10^{-4} M increased both the short circuit current and the resistance.	Likely that the sodium permeability of the outer barrier was altered.	Webb et al., 1979
Lithium Salts (Chloride and Sulphate)	- Rana pipiens and other species		eggs	Severity of effects increased with increased duration of exposure, concentration or temperature.	Microcephalic defects observed.	Hall, 1962
Magnesium (M) (see also lead, cadmium, mercury, manganese)	- Xenopus laevis	50 per group	embryos	Magnesium ions moderated the toxicity of lead, cadmium, manganese and mercury ions.	Tests run for 6 days at 18°C.	Muller and Landesman, 1978
Malathion (OP)	- Thio rugulis		tadpoles 3 weeks old	Thermal tolerance significantly lowered:	24 hr exposure.	Johnson, 1980

Conc. (ppb)	Onset of Spasms (°C)	
	Mean	Range
Control	37.5	36.9-37.8
25	36.9	35.9-37.9
50	36.4	35.2-37.2
100	36.4	35.1-37.5
200	36.1	35.2-37.7

Recommended dosage rate for anadromic control in California-500 ppb.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Malathion (cont'd)	- Microhyla ornata	100 per concentration	embryos yolk-plug stage	Percent abnormals:	Abnormalities include head, trunk and tail defects, behaviour, loss of balance, poor pigmentation and retarded growth.	Pawar et al., 1983
				Duration of treatment		
				conc. (ppm) | 24 hr | 48 hr | 72 hr | 96 hr | % mortality after 96 hr		
				0 | 0 | 0 | 0 | 0 | 0	Temp. - 25°C.	
				1 | 0 | 5 | 13 | 23 | 0		
				5 | 100 | 100 | 100 | 100 | 0		
				10 | 100 | 100 | 100 | 100 | 35		
				15 | 100 | 100 | * | | 100		
				20 | 100 | * | | | 100		
				* All animals died.		
	- Plethodon cinereus P. glutinosus	10 per treatment	adults	Brain cholinesterase was significantly inhibited at 3.6 kg/ha in P. glutinosus but not in P. cinereus. No changes in feeding, endurance or coordination.	Salamanders exposed to substrate treated with malathion.	Baker, 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Malathion (cont'd)	- Rana hexadactyla	6 per treatment	gastrocnemius muscle	Treatment | Twitch Amplitude (cm) | Period of Latency (s)	Results indicate that action of malathion is similar to that of a pharmacologically active eyo- and neuro-toxic agent.	Kowalyo et al., 1987
				I - Control: 2.6 ± .1 | .390 ± .021		
				A: 3.2 ± .3 * | .205 ± .022 *		
				10 ppm M: 2.2 ± .2 | .164 ± .018 *		
				20 ppm M: 1.8 ± .1 * | 1.025 ± .016 *		
				30 ppm M: 0.7 ± .2 * | .984 ± .020 *		
				20 ppm M + A: 2.4 ± .2 | 1.025 ± .016 *		
				II - Control: 1.9 ± .2 | .983 ± .025		
				A: 2.6 ± .3 * | .738 ± .036 *		
				20 ppm M: 0.4 ± .2 * | .287 ± .042 *		
				20 ppm M + A: 0.9 ± .2 * | .246 ± .032 *		
				A = Atropine, M = Malathion; values are mean ± SD. * significantly different from controls p <0.05.		
	- Rana pipiens	24 per concentration	adults approx. 9 cm length	Following exposure to concentrations of 50, 75, 100, 125, 150, 175 ppm for 15 days: - frequency of deaths rose with concentrations; 20 dead after 15 days at 175 ppm - progressive anemia and leucopenia occurred with successively higher concentrations - white blood cell count showed progressive neutropenia and lymphocytosis with increasing concentrations.		Kaplan and Glassmoke, 1965

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Malathion (cont'd)	- Rana tigerina		tadpoles limb bud stage	Conc. % / Percent metamorphosed / Mean length of metamorphosis / Mean time for metamorphosis in days 0.0007 — 20 — 16.2* — 30.0 0.0006 — 40 — 16.2 — 28.0 0.0005 — 60 — 16.2 — 27.0 0.0004 — 60 — 16.2 — 30.0 0.0003 — 60 — 16.3 — 27.3 0.0002 — 100 — 16.4 — 25.0 0.0001 — 100 — 16.6 — 27.4 Control — 100 — 16.7 — 19.6	Temp 30-36°C.	Mohanty-Hejmadi and Dutta, 1981
				* significantly different than controls.		
			tadpoles feeding stage	None metamorphosed at concentrations 0.0007% to 0.005%.		
			eggs	No tadpoles from eggs treated with 0.0007% to 0.004% metamorphosed.		
	- Rana catesbeiana		tadpoles	40% of tadpoles exposed to 5 ppm died.	Continuous flow apparatus for 96 hr. Brain cholinesterase activity of mallards fed dosed tadpoles was 90.6% of that of controls. None died.	Hall and Kolbe, 1980
	- Bufo arenarum	3 per treatment	embryos recently fertilized	47.3 mg/L produced 100% mortality within 5 days of exposure. 0.47 mg/L allowed larval development and metamorphosis. At 0.47 and 47.3 mg/L, an immersion time increased, inhibition of AChE augmented.	No behavioural effects.	de Llamas et al., 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Maneb 80 (F) (Manganese Ethylenebisdithiocarbamate)	- Triturus cristatus carnifex	36 Expt., 36-Ctrl.	adults	5 ppm caused delayed growth, reduced melanogenesis and malformations in regenerated limbs.	Temp 20 ± 1°C. Exposed for 4 days/week.	Arias and Zaverella, 1979
	- Xenopus laevis	30-60 per treatment	embryos yolk plug stage	Exposure (1-5 ppm) produced growth retardation, absence or reduction in melanogenesis in eye, shortened tail, distinct outward curvature. At higher concentrations organisms were unable to swim as well as control specimens.	Temp. 22 ± 1° C. Exposed for 1-10 days.	Bancroft and Prahlad, 1973
	- Triturus cristatus carnifex	190		No tumours were found in any animals at a concentration of 5 ppm. Some edema of the soft tissues and enlargement of spleen, vascular congestion in gut walls found; glycogen content increased in treated males.	Exposure for 19-23 weeks. Temp. 17 ± 1° C for 3 wks then 20 ± 1° C for following 2 wks, pH - 7.6.	Zaverella et al., 1979
	- Triturus cristatus carnifex	30 per treatment	adults	Effect of Maneb 80 on regeneration of limbs:	Female newts only. exposed 5 days/week percutaneously. Temp. 20 ± 1° C.	Zaverella et al., 1984

Treatment Group	No. of limbs examined	No. of limbs w/abnormalities: Slight	Severe
Tap water	10	6	1
Dispersant and wetting agent	10	3	2
5 ppm Maneb 80	10	0	10

Growth delayed in Maneb-exposed newts.
Severe proximal and distal limb abnormalities in all animals exposed to Maneb 80.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Manganese (M)	- Xenopus laevis		myelinated nerve fibres	Decreased permeability constant in K and Na systems; reversibly shifted Na activation curve in positive direction along potential axis.		Arhem, 1980
	- Xenopus laevis		embryos	Mn (ppm) / Hg (ppm): 10 / 5 / 1 / .1 / .01 / .001 250: N / N / N / N / N / N 94 / 98 / 92 / 94 / 84 / 86 20: SL / N / N / N / N / N 94 / 90 / 91 / 100 / 90 / 94 2: M-S / SL-M / N / N / N / N 96 / 96 / 88 / 93 / 96 / 87 .2: S / S / M-S / SL / SL / SL 72 / 50 / 92 / 74 / 96 / 93 0: S / 0 / S / S / S / S 56 / 0 / 79 / 87 / 87 / 92 0 - Deaths; S - Severely deformed; M - Moderately deformed; SL - Slight effects; N - No effects; numbers = % survival from blastula to feeding stage.	Exposure for 6 days at 18°C.	Miller and Landesman, 1978
Mercury (M)	- Bufo regularis	8-10 per group	adults females, 25-30 g	Single i.m. injection of 5.60 mg Hg^{+2}/kg produced significant decrease in serum alkaline phosphatase, lactic dehydrogenase activities. Total bilirubin, serum total protein, aspartate aminotransferase, alanine aminotransferase increased.	5.60 mg/kg = 96 hr LD50. 96 hr exposure. EDTA not a significant treatment.	Hilmy et al., 1984b

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (cont'd)	- Cyprinodon variegatus	100	eggs	Percent mortality and teratogenesis at hatching (H) and 4 days posthatching (PH) of eggs exposed to sediment-bound mercury: Hg added to sediment (ppm) / Concentration in sediment (ppm) / % Mortality: H / PH 0.1 / 0.146 / 45 (11)* / 61 1.0 / 1.188 / 41 (17) / 52 10.0 / 12.08 / 49 (16) / 64 100.0 / 122.83 / 47 (23) / 65	Hardness - 200 ppm as $CaCO_3$. pH - 7.4 - 8.0. Percent mortality expressed as frequency in experimental population/controls. * Percentages of survivors bearing gross congenital deformities at hatching.	Birge et al., 1977
	- Rana catesbeiana		eye	Hg^{+2} caused irreversible decrease in rod response amplitudes; did not affect cones.		Fox and Sillman, 1979
	- Xenopus laevis		myelinated nerve fibres	Hg^{+2} irreversibly shifted Na activation curve in positive direction along the potential axis.		Arhem, 1980
	- Rana nigromaculata		eggs, tadpoles	Exposure of eggs to 0.8 ppm Hg^{+2} caused damage to primordial germ cells and their proliferation rate thereafter seemed to be lower. Lethal to tadpoles. Abnormal tadpoles at 0.4 and 0.8 ppm.	Treatment prior to first cleavage.	Nah, 1978

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (cont'd)	- Xenopus laevis		embryos	Hg (ppm) / Hg (ppm): 1 / 1 / .05 / .01 / .001 200: 0 / N-S / N-S / N / N 0 / 79 / 84 / 93 / 90 20: 0 / N-S / N-S / N / N 0 / 61 / 96 / 94 / 97 2: 0 / N-S / N-S / N / N 0 / 75 / 88 / 93 / 89 .2: 0 / N-S / N-S / SL / SL 0 / 51 / 96 / 93 / 96 0: 0 / S / S / S / S 0 / 70 / 90 / 94 / 94 D - Death; S - Severely deformed; M - Moderately deformed; SL - Slight effects; N - No effects; numbers = % survival from blastula to feeding stage.	Exposure for 6 days at 18°C.	Miller and Landesman, 1978
Mercury (inorganic)	- Xenopus laevis		embryo-larval	Survival at 4 days post-hatch for embryos maintained in 0.16 and 0.36 ug/L averaged 46 and 28%, respectively. Adults had been maintained in 0.2 ug/L for 11 months prior to spawning.	Flow-through system, pH-7.3, water hardness = 102 ppm $CaCO_3$.	Birge et al., 1979b
Mercury (Mercuric Chloride)	- Rana pipiens	100 per treatment	embryos	Percent survival following 4 days of continuous exposure:	Flow through system.	Birge and Just, 1975a

Stage	Concentration (ppm)						
	10	1	0.1	0.01	0.001	0.0001	Control
Cleavage	0	0	0	0	93	94	95
Blastula	0	0	0	0	82	78	84
Gastrula	0	0	0	80	85	95	95
Neurula	0	0	0	93	88	94	100
Tail Bud	0	0	20	80	93	95	95

Table 2 : Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (Mercuric Chloride) (cont'd)	Rana pipiens	100 per treatment	larvae adults	Percent survival of adults (A) and larvae (L):	Flow through assay.	Birge and Just, 1975a (cont'd)

Exposure (days)	Concentration (ppm)													
	50		25		10		7.5		5.0		2.5		1.0		0.5	
	L	A	L	A	L	A	L	A	L	A	L	A	L	A	L	A
1	0	0	0	0	0	70	0	90	0	100	100	100	100	100	100	100
2						60		90		100	100	100	100	100	100	100
3						30		80		100	0	100	100	100	100	100
4						20		80		100	0	100	100	100	100	100
5						20		80		100	0	100	50	100	100	100
6						20		60		100	0	100	50	100	100	100
7						10		60		100	0	100	50	100	100	100
8						0		60		100	0	100	50	100	100	100
9						0		60		100	0	100	50	100	100	100
10						0		60		100	0	100	50	100	100	100

	- Hyperoolia arnoldi	50 per concentration	embryos	200 ug/L produced almost total mortality within 24 hr. 200-250 ug/L caused body blisters; hatched tadpoles were abnormal showing curved body axis, retarded growth, underdeveloped eyes. Most died within 72 hr. 100-150 ug/L produced low mortality, abnormalities were less severe. No significant changes noted at 50 ug/L.	pH 7.1. Temp. 21-25°C. 51 mg/l $CaCo_3$ alkalinity. Exposure 96 hr.	Dhote and Mulherkar, 1980b
			tadpoles (8 - 10 days)	50 ug/L caused total mortality within 24 hr. Survivors at 100-200 ug/L had distended body cavities and were sluggish after 24-48 hr.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (Methylmercury)	- Rana pipiens		ventral surface skin	Dimethylmercury did not alter the P.D. or the short circuit current across the skin at a concentration of 10^{-3} M. Methylmercuric chloride at 10^{-4} M reduced both the P.D. and S.C.C. Electrical resistance rose initially and then declined. The effects were irreversible. Methylmercuric chloride antagonized the action of vasotocin on osmotic water transfer and the O_2 consumption was depressed.		Yorio and Bentley, 1975
		5 per compound	adults	Injection into dorsal lymph sac of 20 mg/kg methylmercuric chloride or 18.4 mg/kg dimethylmercury caused no detectable toxic effects.		
		8		Immersion in 23 mg/L dimethylmercury for 7 days caused no detectable toxic effects.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Mercury (Methylmercury) (cont'd)	- Rana pipiens	200	tadpoles	Exposure	Concentration (ppm)			Chang et al., 1974
					0.001-0.01	0.05-0.09	0.5-1.0	Tadpoles raised in methylmercuric chloride water.
				1 hour	No observable effect	Irritative movements	Irritative movements, difficulties in breathing, escape attempts	
				8 hours	"	Irritative movements, abnormal swimming postures.	As above, 30% mortality	
				12 hours	"	Abnormal swimming postures, difficulties breathing.	75% mortality	Tadpoles injected with 0.025 ml of 0.1% solution methylmercury chloride on alternate days for 10 days showed extensive swelling and distension of their legs.
				24 hours	Appeared lethargic	40% mortality	100% mortality	
				48 hours	Lethargic, no other changes after 4 months of observation and exposure	100% mortality		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (Methylmercury) (cont'd)	- Rana pipiens	N=45 per concentration	embryos	Effects of methylmercury on development of embryos treated for 3 days:	Temp. 21 ± 1°C.	Dial, 1976

Conc. (ppb)	3 days post treatment: Abnormal/Surviving	% abnormal	% dead on 5th day
Stage: Blastula			
0	8/82	10	12
5	13/38	34	19
10	13/45	29	23
15	27/41	66	9
20	34/34	100	54
30	40/0	---	100
Stage: Gastrula			
0	8/81	10	20
5	3/39	8	20
10	8/41	20	31
15	30/36	83	60
20	10/10	100	91
30	2/2	100	100
Stage: Neural Plate			
0	6/83	7	13
5	10/45	22	29
10	18/42	43	30
15	13/45	29	22
20	11/39	28	60
30	31/0	---	100

Abnormalities observed were tail defects, exogastrulae, stunting and poor general development.

Death and severe defects occurred over a narrow range of concentrations and increased with exposure time and increased concentrations.

	- Xenopus laevis	100 per treatment	embryos	Conc. (ppm) / No. Hatched / Defective Dvpt. / Deaths: 0.1 — 0 — 37% — 63%; 0.05 — 18 — 61% — 22%; 0.01 — 18 — 38% — 11%; 0.002 — 14 — 21% — 21%; 0.0004 — 17 — 18% — 12%; 0 — 23 — 0% — 17%		Rumpert and Zietz, 1984

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Mercury (Methylmercury) (cont'd)	- Triturus viridescens	25 per group	adults	Initiation of regeneration of amputated hindlimbs was delayed in newts raised in 0.01-0.07 ppm. No delay in initiation observed in newts raised in 0.1-1.0 ppm. Rate of regeneration markedly increased in all concentrations. All newts raised in 0.3 and 1.0 ppm died after 17 and 8 days, respectively.		Chang et al., 1976
Natural (DDT + methyl parathion)	- Rana tigerina		tadpoles	Effects on feeding stage (F) and limb bud stage (L):	Temp. 30-38°C. Reduced length at metamorphosis at 0.0001% for feeding stage tadpoles.	Mohanty-Mohapatra and Dutta, 1981

Conc. %	% metamorphosis: F	% metamorphosis: L	Mean time for metamorphosis (days): F	Mean time for metamorphosis (days): L
0.0003	-	60	-	36.6
0.0002	-	80	-	37.5
0.0001	20	100	55.0	33.6
0.00009	20	no data	59.0	no data
0.00008	40	"	60.0	"
0.00007	40	"	59.5	"
0.00006	40	"	60.0	"
0.00005	60	"	64.6	"
0.00004	80	"	64.5	"
0.00003	100	"	64.6	"
0.00002	100	"	66.0	"
0.00001	100	"	67.2	"
Control	100	100	35.6	21.0

	tadpoles	Conc. % / % metamorphosis / Mean time for metamorphosis: 0.00005 — 20 — 62.5; 0.00004 — 50 — 60.2; 0.00003 — 80 — 62.5; 0.00002 — 90 — 59.2; 0.00001 — 100 — 57.6; Control — 100 — 36.9	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Methoxychlor (OC)	- Rana pipiens		abdominal skin	2×10^{-4} M produced no significant effect on the short circuit current or resistance.				Wade et al., 1979
	- Rana pipiens		adults 45 g, 8.9 cm long	Methoxychlor (ppm)	pH of solution at 25° C	No. of frogs dead at 30 days	Frogs placed in 200 cc of test solution. Frogs which died had exhibited neuromuscular changes, excessive thrashing and abnormal reactivity to stimulation. Survivors were free from disorders.	Kaplan and Overpeck, 1966
		20		0.80	6.31	5		
		10		0.40	6.10	0		
		10		0.40	5.95	0		
Methyldemeton (OP)	- Rana tigerina		eggs	Conc. %	% Metamorphosis	Mean time for metamorphosis (days)	Temp-30-36°C. Doses used were lower than suggested field applications. Reduced size at metamorphosis at 0.00003%.	Mohanty-Mojumdi and Dutta, 1981
				0.00003	20	42.0		
				0.00002	30	46.6		
				0.00001	50	47.2		
				Control	70	33.7		
			tadpoles feeding stage	Conc. %	% Metamorphosis	Mean time for metamorphosis (days)		
				0.00004	40	51.0		
				0.00003	60	51.6		
				0.00002	60	55.6		
				0.00001	80	56.5		
				Control	100	33.2		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Methyldemeton (cont'd)	Rana tigerina		tadpoles limb bud stage	Conc. %	% Metamorphosis	Mean time for metamorphosis (days)	Reduced size at metamorphosis at 0.0012%	Mohanty-Mojumdi and Dutta, 1981 (cont'd)
				0.0008	60	29.3		
				0.0007	60	24.6		
				0.0006	40	24.5		
				0.0005	80	22.5		
				0.0004	100	19.4		
				0.0003	100	18.6		
				0.0002	100	16.8		
				0.0001	100	18.8		
				0.00004	no data	no data		
				0.00003	no data	no data		
				0.00002	no data	no data		
				0.00001	no data	no data		
				Control	100	14.6		
Methyl Fluoroacetate	- Green frog		sciatic nerves	5 uM blocks nerve in 3-4 hr; 50 uM blocks within 1.5 hr. .025 and .001 M produced 80 and 25%, respectively, inhibition of respiration after 2 hr.				Boyarsky et al., 1949

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Methylhydrazine (Component of jet fuel)	- Xenopus laevis	50-110 per treatment	larvae	1 mg/L produced irreversible toxic effects and death within 48 hr. Normal metamorphosis occurred in larvae in continuous contact with 0.1 mg/L.		Greenhouse, 1976a
		50-100 per treatment	embryos	≤5 mg/L harmless if embryos transferred to uncontaminated water by completion of neurulation. Teratogenic at ≥10 mg/L.	Most common abnormality was tail kinks.	
	- Xenopus laevis		embryos	Concentration (mg/L) / % Malformed: 100 — control — 3 100 — 3 — 1 200 — 5 — 52 150 — 10 — 93 50 — 15 — 100	Kinked tail was most frequent malformation. Most larvae with kinked tails metamorphosed into normal frogs. Other malformations included excrescences, failure of elongation and edema.	Greenhouse, 1976b
				Susceptible during neurulation.		
Methyl isothiocyanate (F)	- Xenopus laevis	60 per concentration	embryos yolk plug stage	Conc. (ppb) / % Mortality at day 10 / % Abnormal at day 10: Control — 5 — 0 0.001 — 5 — 0 0.005 — 5 — 0 0.01 — 10 — 0 0.05 — 50 — 0 0.1 — 70 — 0 0.5 — 90 — 0 1.0 — 100 — 0		Birch and Prahlad, 1986
				Embryos exposed to 100-500 ppb did not survive after day 5; those exposed to 400 ppb did not complete neurulation.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Methyiparathion (MP) (see also metaxid)	- Bufo boreas	20	juveniles (X snout-vent length - 19.8 mm)	Exposure of hydrated toads to 25 ppb significantly lowered temperature tolerance.	Temp. 23-24°C; 24 hr exposure.	Johnson and Prins, 1976
	- Rana cyanophlictis	6 per treatment	tadpoles approx. 4 weeks old	Brain Glucose Level (mg/g ww) / Oxygen Consumption (ul/g/h): control — 4.76 ± 0.30 * — 283.81 ± 0.1 5 ppm — 2.4 ± 0.91 — 305.26 ± 65.42 p < 0.05 — p < 0.01 * values are mean ± S.D.	Exposure for 24 hr.	Yasmeen and Mayeenuunissa, 1985
	- Hyla regilla		tadpoles 3 weeks old	Thermal tolerance significantly lowered:	24 hr exposure.	Johnson, 1980
				Conc. (ppb) / Onset of Spasms (°C) — Mean / Range: Control — 37.3 — 36.9-37.8 25 — 35.7 — 31.7-37.8 50 — 34.3 — 29.1-36.8 100 — 33.6 — 30.7-36.3	Recommended dosage rate for mosquito control in California - 100 ppb.	
		54 27 36 36				
	- Rana cyanophlictis		tadpoles approx. 4 weeks old	Exposure for 24 hr to 2.5 ppm affected the qualitative nature of the brain lactate dehydrogenase isozymes.		Yasmeen and Mayeenuunissa, 1986
Mine Drainage (Iron, Zinc, Copper)	- Bufo boreas	200 - test, 200 - control	zygotes	All zygotes placed in mine drainage (pH 2.79, 260 mg/L Fe, 39 mg/L Zn, 3.7 mg/L Cu) died within 12 hours and before they reached the second cleavage stage.	Zygotes cultured in lake water (pH 7.1, negligible quantities of metals) developed normally through metamorphosis.	Porter and Hakanson, 1976

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Monosodium Methyl Arsenate (H) (MSMA)	- Lithobius forficatus Lumbricus terrestris	20 per conc.	adults	No deaths observed following exposure to 300-400 ppm for 96 hr. No deaths observed following exposure to 130-520 ppm for 96 hr.		Johnson, 1976
(Ansar 592 HC)	- Scaphiopus couchi	5 adults per group, 15 juveniles per group	adults 50-61 mm snout-vent length, juveniles 14-19 mm snout-vent length	100 ppm (47.74 ppm MSMA) was not toxic to juveniles. 1,000 ppm (477.4 ppm MSMA) was toxic to juveniles but not to adults. Cumulative % mortality for juveniles: 120 hr - 6.6%; 168 hr - 53.3%; 192 hr - 86.6% 10,000 ppm (4,774 ppm MSMA) was toxic to both adults and juveniles; adults were less sensitive. Cumulative % mortality for juveniles: 48 hr - 60.0%; 72 hr - 100.0% 100,000 ppm (47,740 ppm MSMA) was toxic to all adults and juveniles within 2 hr of exposure.	Possible reasons that juveniles were susceptible than adults: 1) surface area: volume ratio 2) skin permeability 3) metabolic rate 4) prior exposure	Judd, 1977
Nabam (F) (Ethylenebis dithiocarbamic acid disodium salt)	- Xenopus laevis	30-60 per treatment	embryos yolk-plug stage	1-3 ppm induced waviness of notochord and punctate melanophores.	Temp. 22 ± 1°C. Exposure - 1-10 days.	Bancroft and Prahlad, 1973

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Nabam (F) (Ethylenebis dithiocarbamic acid disodium salt) (cont'd)	- Xenopus laevis	60 per concentration	embryos yolk plug stage	Conc. (ppb)	No. surviving at day 10	% Abnormal at day 10	Abnormalities involved swollen and kinked notochords. Storage of dilute nabam solutions for 1 week greatly enhanced its teratogenic and lethal potential.	Birch and Prahlad, 1986
				Control	57	0		
				10	58	0		
				20	56	0		
				30	57	5		
				40	60	62		
				50	58	79		
				100	57	98		
				1000	56	96		
	- Xenopus laevis	30	embryos stage 10-11	Exposure to 4 ppm for 7 days caused alterations in pigmental retina, notochord and skin at ultrastructural level.				Prahlad et al., 1974
Nickel (Ni^{+2}) (H)	- Triturus pyrrhogaster		stomach mucous epithelial cells	Lower concentrations did not affect the electrical properties of the cells. 10^{-4} M decreased membrane potential to 82% of control value.				Kanno et al., 1978
	- Xenopus laevis		myelinated nerve fibres	Slowed down kinetics of K system, decreased permeability constant in K and Na systems, reversibly shifted Na activation curve in positive direction along potential axis.				Arhem, 1980
Niclosamide (molluscicide) (Bayluscide, Bayer 73)	Bufo regularis	100	adults 50 g	Force feeding 0.3 mg daily during non-breeding season induced formation of kidney tumours in 2 toads.			Other researchers induced lesions in 23% of animals treated during breeding season using same dose and methodology.	Sabry and El-Hefty, 1984
Nonachlor (OC)	- Rana pipiens		abdominal skin	Produced a decrease in the short circuit current and increased the resistance.				Webb et al., 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Nonylphenolethoxylate (surfactant)	- Rana esculenta		leg skin	5×10^{-5} M inhibited the isosmotic, active transport-coupled volume flow.		Calentano et al., 1979
HRDC 119 (PY)	- Rana temporaria		sartorius nerve-muscle	Multiple end-plate potentials appeared after exposure to >2 mg/L for 18-45 min. 1-100 mg/L had no effect on muscle resting potential or amplitude or time course of the initial evoked end-plate potential.		Evans, 1976
Octyl-phenyl- α-naphthylamine (component of jet and rocket fuel)	- Rana pipiens Xenopus laevis	200	embryos larvae	Up to 1 g/L not toxic to either species.		Greenhouse, 1976a
	- Rana pipiens Xenopus laevis	100 per treatment	embryos	200 and 1 mg/L had no deleterious effects on development of R. pipiens and X. laevis, respectively.		Greenhouse, 1976c
OMPA (OP) (Octamethyl Pyrophosphoramide)	- Frog		sartorius muscle	10 mM eliminated response to carbamylcholine almost completely. Amount of inhibition was strongly dependent upon concentration of OP and temperature.		Oakin et al., 1978
	- Rana pipiens	24 in each concentration	adults approx. 9 cm long	Following exposure to concentrations of 2500, 2800, 3100 ppm for 15 days: - Anemia and leucopenia produced, worsened with increasing concentration - Progressive neutropenia and lymphocytosis evident with increasing concentration - Posture drooping, activity decreased, flaccid paralysis.		Kaplan and Glaczenski, 1965
	- Frog		sartorius muscle	Reduced amplitude of response without significantly shifting the dose response curve along the abcissa.	Results suggest that OMPA may bind to at least 3 sites on receptor.	Guy et al., 1977

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Oxamyl (C)	- Rana temporaria		tadpoles	Exposure to 100 ppm oxamyl for 1 hour resulted in 90% of tadpoles with vertical curvature deformities.		Cooke, 1981
Paraquat	- Hyla chrysoscelis		embryos	Not toxic or teratogenic at 100 ppm.	Dermally exposed for 24 hr, no. survivors counted 120 hr post-exposure.	Fulton and Chambers, 1985
Paraquat (H)	- Rana pipiens	110 per dose group	embryos	Concentration (ppm): 0 / 0.1 / 0.5 / 2 / 10 No. alive on day 12 post-treatment: 83 / 76 / 6 / 0 / 0 Development proceeded normally until approximately 3 days post-hatch. Growth rate slowed in all groups; tail abnormalities noted at 0.5 ppm.	Temp. 21 ± 1°C. Paraquat rapidly disappears from water thus timing of application is critical.	Dial and Bauer, 1984
	- Rana pipiens	80 per dose group	embryos: early gastrula and 15 days of age	% alive 16 days post-treatment after exposure at gastrula stage (G) or 15 days of age (15): Paraquat (mg/L): 0.5 (G: 4.3; 15: 44.7); 2 (G: 0.0; 15: 9.0)	All figures significantly lower than controls. Retardation of growth, tail malformations, poor food development observed.	Dial and Dial, 1987
	- Rana esculenta	10 per treatment	lung and liver	Activities of antioxidant enzymes were decreased or not affected by LD_{50} injected into abdominal lymph sac at 4°C; LD_{100} resulted in significant increase in activity for most enzymes. Lipid peroxidation of liver decreased in response to paraquat at 4°C, increased at 28°C; in lung lipid peroxidation increased at both temperatures.	260 mg/kg = LD_{50} at 28°C; 360 mg/kg = LD_{100} at 28°C. Lower temperature increased survival.	Barahao et al., 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Parathion (OP)	- Acris crepitans	72 per treatment	adults	Mortality at 96 hr exposure was dose-related (0, 0.1, 1.0, 10 ppm).	pH - 7.6; $CaCo_3$ - 80 mg/L Temp - 23-24°C. 1 bird (Falco sparverius) died within 3 hr after consuming 5 frogs exposed to 10 ppm for 96 hr.	Fleming et al., 1982
	- Bufo arenarum		tadpoles	95% of tadpoles exposed to 5 ppm died. Bioconcentration factor: X 64	Continuous flow apparatus for 96 hr. Brain cholinesterase activity of mallards fed dosed tadpoles was 0.8% of that of controls. All ducks died within ½ hr.	Hall and Kolbe, 1980
	- Bufo arenarum	32 per treatment	adults brain, liver, blood	Toads exposed to 0.1 ml/m^2 fumigant for 45 days or 2 hr experienced a plasma cholinesterasic activity decrease of 86% the normal value. Activity quickly recovered when toads replaced into normal conditions.	Temp. 16 ± 2°C during hibernation, 22 ± 3°C during mating season.	Guzman and Guardia, 1978
	- Rana pipiens	24 in each concentration	adults approx. 9 cm long	Following exposure to concentrations of 5,10,15,20,25 ppm for 15 days: - Anemia and leucopenia which worsened with increasing concentration produced - Progressive neutropenia and lymphocytosis as concentration increased - Marked decrease in activity, decreased muscle tone, generalized edema		Kaplan and Glaczenski, 1965

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Parathion (cont'd)	- Bufo viridis	3 per compound	adults	Residual cholinesterase at time of death following injection of 2 LD50 into dorsal lymphatic sac:	LD50: parathion - 967 ppm. paraoxon - 188 ppm.	Edery and Schatzberg-Porath, 1960

	Parathion	Paraoxon
blood	57%	5%
mixed nervous tissue	43%	12%

		Metabolism of parathion incubated with liver slices for 90 min (values in mmol ± S.E.M.):		Paraoxon readily degraded thus levels given are difference between production and destruction.	Potter and O'Brien, 1964
	liver	No. replicates	Parathion remaining	Paraoxon recovered	
	Rana pipiens	6	52.2 ± 5.34	2.20 ± 0.47	
	Bufo sp.	6	37.6 ± 4.24	2.81 ± 0.45	
Permethrin (PY)	- Xenopus laevis		lateral-line nerve organ	5×10^{-6} M in vitro for 3 hr or 10^{-5} M in vivo for 5 hr induced short trains of nerve impulses.	Temp. 8-22°C. Number of impulses per train and train duration increased with cooling.	Vijverberg et al., 1982
			peripheral nerves	5×10^{-6} M caused repetitive activity within first 2 hr of exposure.		
pH (see also aluminum)	- Xenopus laevis	10-20	embryos stages 10-13	Total inhibition of hatching occurred at pH 3.9 in sulfuric acid and pH 4.3 in bog water. In both types, at low pH embryos underwent a tight coiling associated with shrinkage of the perivitelline space; when jelly layer removed at pH 4.3 the embryos developed normally.	Temp - about 25°C. Possible cause - block in functioning of hatching enzyme.	Dunson and Connell, 1982
	- Rana catesbeiana	26	adults 100-300 g	Brain AChE activity altered in pH-dependent fashion. Animals raised at pH 4.6 died within 7-10 days.	Brain AChE levels not affected by presence of Al^{+3}.	Marquis 1982

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	- Rana sylvatica		embryos 4 hr post-fertilization	Differences in acid tolerance among populations of embryos was not related to levels of acidity in ponds.	Temp. 15°C, medium changed daily.	Pierce and Harvey, 1987
			larvae stages 25-27	Tadpoles produced by adult from acidic ponds were more acid-tolerant.	Temp. 15°C.	
	- Ambystoma maculatum	30 per treatment	larvae	pH 3 - all died within 12 hr. pH 4 and 5 - significantly slower rate of development and growth than those reared at pH >5.		Ling et al., 1986
	Rana sylvatica		larvae	pH 3 - all died within 2 hr. No differences in development noted at any other treatment.		
	- Hyla crucifer		larvae 4 weeks old	96 hr survival was over 90% at pH 7.0 and about 70% at pH 5.0. No tadpoles survived 24 hr at pH 4.0.	Static bioassay.	Correll et al., 1987
	- Ambystoma jeffersonianum Rana sylvatica		larvae	Acute exposure to pH 2.5 - 4.0 depressed sodium influx and markedly accelerated sodium efflux - resulting net loss of 50% body Na was fatal. Chronic exposure caused 21-62% reduction in body Na level, K content did not change.		Freda and Dunson, 1985a
			embryos	A. jeffersonianum did not hatch below pH 4.50; R. sylvatica hatched at pH 4.25.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	- Rana temporaria	4 per treatment	eggs early cleavage	Mortality Rate (mean % ± SD):	Most hatchlings at pH 4.0 were deformed; at pH 4.5 or 5.0 few deformities noted.	Leuven et al., 1986

pH	Mortality (basic medium)
4.0	96.6 ± 2.8
4.5	16.0 ± 5.8
5.0	8.0 ± 0.5

Mean mortality increased significantly when Al concentration of medium raised from 7.4 to 92.5 umol/L.

Mortality figures for eggs of R. arvalis, R. esculenta, B. bufo similar.

				% Mortality		
			pH	Basic medium	Al medium
		larvae stage 29-33				Al medium: $[Al^{+3}]$ = 185 umol/L
	Rana arvalis		4.0	0.0	0.0	
			4.5	0.0	75.0	
			5.0	0.0	83.3	
	Rana esculenta		4.0	6.3	100.0	
			4.5	6.7	100.0	
			5.0	0.0	100.0	
	Bufo bufo		4.0	23.8	14.3	
			4.5	0.0	20.6	
			5.0	0.0	47.6	
	- Plethodon cinereus	30	adults	Animals given substrate choice were found 50% of time on pH 6-6.5, 31% on pH 4.5-5, 19% on pH 3-3.5.		Wyman and Hawksley - Lescault, 1987
		10 per pH		Animals held on substrates of pH 2-2.5 died within 1 week; those on pH 3-6 survived.	Temp. 8-10°C. 1 month exposure.	
		10 per pH		40% held on substrates of pH 3 died within 4 mo, all others survived. Growth reduced 40% and 45% in salamanders living on pH 3 and 4, respectively. O_2 consumption rate reduced by 32% in animals living on pH 3.	8 month exposure.	

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	-		adults	Selection of substrate pH:		Mushinsky and Brodie, 1975

Field pH	% selecting pH 5.5	% selecting pH 7.7
6.8-7.0	27.5	72.5*
4.5-5.5	43.2	56.8*
5.5	37.5	62.5*
-	40.0	60.0*
5.5	47.0	53.0
-	32.9	67.1*
-	50.0	50.0
-	53.3	46.7

SPECIES	N
Desmognathus f. fuscus	20
D. f. auriculatus	32
Ambystoma opacum	28
A. maculatum	16
Eurycea quadridigitatus	10
E. lucifuga	7
E. longicauda	4
Batrachoseps attenuatus	18

* = significant difference.

	- Assorted species/review		adults, larvae, embryos	Inter- and intraspecific variation and ontogenetic change in acid tolerance; mechanisms of tolerance reviewed.		Pierce, 1985
	- Bufo bufo	4 per test	tadpoles 30-35 mm	pH of solution / Survival time: 2.0 — 55 min; 2.8 — 125 min; 3.4 — 6.5 hr; 3.8 — 18 hr; 4.0 — over 24 hr; 4.2 — no apparent effect in 3 days	Temp. 18°C.	Jones, 1939
	- Rana clamitans	21-26	premetamorphic tadpoles, juveniles	Exposure to pH 4.0 (nominal) caused substantially increased transepithelial net ion loss and net acid uptake, and a slight inhibition of active ion transport. Disturbances disappeared by 7 hr exposure in tadpoles but persisted in juveniles.	Soft water (Ca^{+2} = 300 ueq/L). Prior acclimation to pH 5 did not reduce disturbances.	McDonald et al., 1984

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	-		embryos	Mean % of hatch or coil (±SE):		Karns, 1984

Bog (pH 4.2)		Fen (pH 7.3)	
Hatch	Coil	Hatch	Coil
11.0 ± 1.15 (1750)	77.2 ± 2.12	96.3 ± 0.61 (2121)	0.4 ± 0.21
0 (164)	59.1 ± 13.51	97.3 ± 0.29 (152)	0
0 (3038)	14.6 ± 2.76	93.0 ± 2.21 (1921)	0.2 ± 0.16
0 (435)	0.9 ± 0.66	98.5 ± 0.66 (987)	0
0 (285)	0	64.5 ± 10.38 (232)	0
0 (325)	0	71.5 ± 7.13 (419)	0.3 ± 0.30

SPECIES
Rana sylvatica
Ambystoma laterale
Bufo americanus
R. pipiens
Pseudacris triseriata
Hyla crucifer

Total number of eggs tested in parentheses.

Coil refers to embryos which develop but do not hatch.

Bog population of R. sylvatica more tolerant than marsh population of low pH, suggests adaptation to bog environment.

				Fertilization of R. sylvatica, B. americanus, R. pipiens eggs in bog water not significantly different than controls.		
	- Rana pipiens		tadpoles	Linear increase in survival time and thus pH tolerance during first 8 weeks of development. Tadpoles at pH 4.4 and 5.8 grow at rates of 5.9 and 16.0 mg/day, respectively.	24 day test period.	Freda and Dunson, 1985c

				"Lethal" pH (100% mortality)	"Critical" pH (50% mortality)		
	-		embryos			Sensitivity of X. laevis, Ambystoma jeffersonianum greater in late stages of development than during initial cleavage.	Rowe and Pough, 1982
	Xenopus laevis			3.0	3.5		
	Rana sylvatica			-	3.5		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS						REMARKS	REFERENCE
				pH	Ca (ppm)	% hatched	% of hatchlings deformed	% of embryos killed			
								Early	Late		
pH (cont'd)	-										Freda and Dunson, 1985b
	Ambystoma jeffersonianum	20 per treatment	embryos	5.0	1	100	0	0	0	Addition of Ca, Mg and >10 ppm Na prevented early mortality of embryos.	
					20	95	0	5	0		
					40	100	0	0	0		
					80	95	0	5	0		
				4.5	1	70	100	0	30	Increasing concentrations of ions caused curling defect.	
					20	100	5	0	0		
					40	100	0	0	0		
				4.0	1	0	0	100	0		
					20	0	0	0	100	A. jeffersonianum could usually hatch even though curled but R. sylvatica usually died.	
					40	20	0	5	75		
					80	40	50	5	55		
	Rana sylvatica	20-40	embryos	5.0	1	95	0	0	5		
					20	100	23	0	0		
					40	95	14	0	5		
				5.0	1	100	0	0	0		
					20	80	0	20	0		
					40	100	10	0	0		
					80	100	0	0	0		
				4.5	1	90	5	10	0		
					20	100	0	0	0		
					40	33	31	5	62		
					80	15	67	20	65		
				4.0	1	10	75	50	32		
					20	13	20	7	80		
					40	8	0	15	77		
					80	0	0	10	90		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS		REMARKS	REFERENCE
pH (cont'd)	- Rana pipiens, R. catesbeiana, R. clamitans		larvae	Acute exposure to pH 2.5 - 4.0 depressed Na influx and accelerated Na efflux. Increased external Ca slowed loss of Na. Initial body Na content was inversely correlated with acid tolerance.		Net loss of 50% body Na was fatal. Species differed in Na content under control conditions.	Freda and Dunson, 1984
	- Rana ridibunda		abdominal skin	Increase in influx and backflux of Cl and mannitol and in backflux of Na at pH 2.5 in short-circuited skin. Total conductance and short-circuit current increased at pH 2.5.		Control = pH 7.4.	Ferreira and Hill, 1982
	- Ambystoma opacum	6-12	embryos	Hatching most rapid between pH 6 and 7.		Temp. 22-24°C.	Potranke et al., 1982
	- Rana sylvatica			pH	Mean % hatching ± S.E.M.		Pierce et al., 1984
		662	embryos	7.2 - 7.6	71.7 ± 6.5		
		131		6.0	77.3 ± 9.1		
		169		5.0	88.6 ± 7.6		
		616		4.0	72.1 ± 6.0		
		375		3.0	0 ± 0		
		125		2.0	0 ± 0		
				pH	% survival		
		15	larvae (early feeding)	≥4.0	100.0		
				3.75	93.3		
				3.5	60.0		
				≤3.25	0		
		10	larvae (metamorphosis)	≥3.75	100.0		
				3.5	90.0		
				≤3.25	0		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	- Ambystoma texanum	100 per treatment	embryos	Hatching success:	Temp. - 17° C in pond water from collection sites, pH range 6.8 - 6.7.	Punzo, 1983

	Percent Survival pH							
Temp. °C	4	5	6	7	8	9	10	11
5	0	0	0	0	0	0	0	0
10	13	21	75	83	81	71	69	19
15	17	30	95	92	90	78	73	29
20	9	20	64	74	66	59	50	2
25	6	14	30	26	27	19	7	0
30	0	0	0	0	0	0	0	0

Highest oxygen consumption rates at pH 6.6; significant decrease at pH 4 and 11.

	- Rana pipiens		eggs	Spore motility decreased with decreasing pH below 6.5 in solutions acidified with H_2SO_4 or HNO_3; below 5.5 in solutions acidified with HCl. No eggs developed at $\leq$ pH 4.0. Fertilization and formation of healthy embryos decreased below pH 6.3. Lower limit for optimal fertilization and early development was pH 6.0.		Schlieter, 1981
	- Bufo boreas		larvae	pH 3.1 - all died within 24 hr. pH 4.0 - all metamorphosed.	Buffered solutions. Up to 59 days exposure.	Porter and Hakanson, 1976

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS					REMARKS	REFERENCE
pH (cont'd)	- Ambystoma macrodactylum		embryos at or prior to 4th cleavage division	12°C			21°C		At low temperature egg masses at pH $\leq$3.5 turned milky-opaque within 2 hr of exposure; at pH 4.5 embryos tightly coiled, those that hatched were deformed.	Dale et al., 1981
				pH	n	% hatch	n	% hatch		
				7.0	48	23	54	63		
					52	52	47	51		
				6.0	86	6	42	40		
					85	26	42	81		
				5.0	67	40	75	32		
					57	11	76	46		
				4.5	56	2	110	2		
					111	9	56	11		
				4.0	62	0	72	0		
					42	0	83	1		
				3.5	>50	0	>50	0		
					>50	0	>50	0		

	Hyla crucifer	35 per treatment	embryos mid-blastula stage	pH	% hatch	% hatch, later died	Temp. 20°C. All hatched larvae at pH 4.0 were deformed.	
				8.0	83	0		
				7.0	80	0		
				6.0	90	0		
				5.0	77	11		
				4.0	54	100		
				3.0	0	-		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	n	STAGE/TISSUE	LEVELS USED/EFFECTS					REMARKS	REFERENCE
				12°C			21°C			
				pH	n	% hatch	n	% hatch		
pH (cont'd)	Rana sylvatica		embryos at or prior to 4th cleavage division	7.0	451	92	423	92	At pH 4.0, those which hatched at low temperature died.	Dale et al., 1985 (cont'd)
					489	95	213	98		
				6.0	338	51	378	77		
					318	44	346	80		
				5.0	473	51	262	97		
					357	39	319	90		
				4.5	340	38	337	95		
					296	40	323	82		
				4.0	328	17	119	8		
					313	6	276	<1		
				3.5	>200	0	>200	0		
					>200	0	>200	0		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT	SPECIES	n	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
				pH	% survival after 30 days at 5°C	% survival after 14 days at 21°C		
pH (cont'd)	Rana clamitans	10 per replicate	larvae stage 25-29	6.0	40	100		Dale et al., 1985 (cont'd)
					30	100		
					60	100		
				5.0	80	100		
					80	100		
					80	100		
				4.5	60	100		
					90	100		
					60	100		
				4.0	50	100		
					90	100		
					80	100		
				3.5	30	100		
					60	100		
					60	100		
				3.3	-	90		
						0		
						0		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	n	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
pH (cont'd)	Rana palustris		embryos at or	pH	% hatch	% hatched, later died	Temp. 12°C.	Dale et al., 1985 (cont'd)
		43	before 32 cell	7.0	100	0		
		37	stage		100	0		
		41		6.0	100	0		
		53			100	0		
		49		5.0	41	50		
		22			34	50		
		40		4.5	5	50		
		44			5	50		
		39		4.3	0	-		
		48			0	-		
	Bufo americanus	46-51 per replicate	embryos early gastrula	pH		% hatched	Temp. 21°C.	
				6.0		100		
						100		
				4.5		100		
						100		
				4.3		100		
						82		
				4.0		84		
						94		
				3.8		0		
						0		
	Notophthalmus viridescens	10 per replicate	adults	pH	% survival after 30 days at 5°C	% survival after 14 days at 21°C		
				5.0	100	100		
					100	100		
					100	100		
				3.8	100	100		
					100	100		
					100	100		
				3.5	10	75		
					20	87		
					30	50		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	n	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	- Ambystoma maculatum	Approx. 150 per treatment	eggs at grey crescent stage	Initial perivitelline fluid pH's decreased significantly 7 to 96 hr after initial immersion of eggs in water altered to pH 4.0 and 5.0. Rate of H^+ flux into the fluid is greater in eggs in pH 4 than in those in pH 5 or 6.		Rath and Laven, 1987
		100 per treatment	larvae newly hatched	At pH 3.0, 100% mortality occurred after 1.25 hr larval immersion; at pH 3.5, 75% mortality occurred after 2.5 hr; at pH 4, 10% had mortality not occurred after 6 days.		
	- Rana temporaria	12 per treatment	tadpoles	Trends observed for pH 3.6 - 6.5: - maximum size attained was positively correlated with pH - time to foreleg emergence was negatively correlated with pH.	Tadpoles raised in given pH to metamorphosis.	Cummins, 1986
	- Ambystoma jeffersonianum Rana sylvatica	20 per pH	embryos	Mean % hatching:	In artificial soft water.	Freda and Dunson, 1986
				pH / A. jeffersonianum / R. sylvatica		
				5.00 / 100 / 95		
				4.75 / 100 /		
				4.50 / 25 * / 100		
				4.25 / 0 * / 60		
				4.00 / 0 / 0		
				* significant difference between the two species.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	Hyla andersonii Bufo woodhousei Rana clamitans	40 40 20 unless otherwise indicated	embryos	Percent hatching: B. woodhousei: pH 5.8 ASW 100; 4.25 ASW 100; 4.10 ASW 85, BW 45; 4.00 ASW 0*, BW 0; 3.75 ASW 0, BW 0 H. andersonii: ASW 100; 98; BW 93; 97**; BW 85; 25; BW 20 R. clamitans: ASW 100; 35; BW 10; 0 * n=80 ** n=60	Tests done in artificial soft water (ASW) or naturally acidic bog water (BW). Growth significantly reduced at lower pH's during first 10 days.	Freda and Dunson, 1984 (cont'd)
	- Rana sylvatica	20-30 per treatment	eggs	At pH 7.2 - 7.6 hatching success was 95.3%, success dropped significantly to 86.6% at pH 4 and to 41.9% at pH 3.75. Time required for hatching increased significantly as pH decreased.	At pH 4 only slight differences observed in embryo tolerance among progeny of males mated to same female; progeny from different females differed significantly in acid tolerance.	Pierce and Sikand, 1983
			larvae	All survived for 24 hours at pH 7.2 - 7.6; at pH 3.5 only 37% survived and average survival time was 18.4 ± 0.41 hr.		
	- Rana catesbeiana	30 per pH	tadpoles stages 25-30	Percent mortality increased from near 0% at pH 4.3 to 78% at pH 4.2; mortality was 100% at pH 3.9.	Exposure for 4 days.	Gosner and Rider, 1985
	Rana clamitans	30 per pH	tadpoles stages 25-30	Mortality (estimated from graph) was 10% at pH 4.5 and 90% at 4.1.		
	- Rana pipiens Xenopus laevis	10 9	adults/skins	Lowering of pH caused a decrease in short circuit current generated by the skins of both species.	Skin of X. laevis was more sensitive than that of R. pipiens to acid stress.	Frazer, 1981

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
pH (cont'd)	- Ambystoma maculatum A. jeffersonianum			A. maculatum - tolerated pHs from 4-10 - greatest hatching success at 7-9. A. jeffersonianum - tolerated pHs from 4-8 - greatest hatching success at 5-6.	pH optimum shifted upward with increasing temperature for A. jeffersonianum and downward for A. maculatum	Pough and Wilson, 1977
			embryos	Effects of acidity on development of [illegible] species:	Morphological and behavioural abnormalities due to shrinkage of the perivitelline membrane. Short term pH fluctuations are also toxic to developing embryos.	Gosner and Black, 1957
	- Acris gryllus Pseudacris nigrita Hyla andersonii H. crucifer H. versicolor Rana catesbeiana R. clamitans R. palustris R. pipiens R. sylvatica R. virgatipes			pH: 4.6, 4.1, 3.8, 4.2, 4.3, 4.3, 4.1, 4.3, 4.1, 3.9, 3.8 gpe: 4.1, 3.8, -, 3.8, -, -, -, 4.8, 3.7, -, - * pH levels at which 50% or more embryos develop into normal hatchlings. ** Observed lethal level.		
Phenol	- Xenopus laevis		embryos	No teratogenic effect. No effect on embryos at 10 ppm, but animals died within 4 days to 3 weeks of completing embryonal development.		Dumont, 1987
Phenols	- Assorted species/review			Cellular neurophysiological effects reviewed.		Koslo, 1982

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
N-Phenyl-α-naphthylamine (component of jet and rocket fuel)	- Rana pipiens Xenopus laevis	100-176 per treatment	embryos	All R. pipiens embryos exposed to 20 or 200 mg/L developed malformations and all were dead by heart beat stage; 94.8% of X. laevis exposed to 6 mg/L developed malformations while exposure to ≤5.2 mg/L produced no observable effects.			Estimated ED_{50} = 4-5 mg/L. Malformations included shortened trunk and intestinal tract and edema.	Greenhouse, 1976a,c
Phenyl Saligenin Cyclic Phosphate			embryos	Conc. (ppb)	Mortality (%)	Abnormal Embryos (%)	Exposure for 24 hr, no. survivors determined 120 hr after exposure. Edema was most common defect. 500 ppb induced earlier deformities in Gastrophryne carolinensis.	Fulton and Chambers, 1985
	Rana sphenocephala	120 160 210 140		0 100 500 1000	0 0 6 99	0 0 53 100		
	Hyla chrysoscelis	161 86 84 80 160 40		0 100 250 500 750 1000	2 7 10 32 52 92	0 0 0 10 58 100		
Phosdrin (DP)	- Rana pipiens	24 in each concentration	adults	Following exposure to concentrations of 4, 9, 12, 16, 20 ppm for 15 days: - Produced anemia and leucopenia which worsened with increasing concentration - Differential white cell count showed progressive lymphocytosis and neutropenia with successively higher concentrations - Decrease in activity, flaccid paralysis.				Kaplan and Glaczenski, 1965

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Pollution (Micropollutants in drinking water)	- Pleurodeles waltl		larvae approx. 40 mm	Larvae reared in untreated tap water had higher levels of micronucleated erythrocytes than those reared in filtered treated water.			Used to evaluate mutagenic activity in drinking water.	Jaylet et al., 1987
Textile Plant Effluent	- Rana cyanophlyctis	10	adults	25% mortality after 96 hr, 50% after 144 hr.			Frogs maintained in effluent.	Shrinivas et al., 1984
Potassium Chromate	- Rana hexadactyla		gastrocnemius muscle	Percent change in succinate dehydrogenase (SDH) and Mg^{+2} ATPase activity levels over control levels in muscle soaked in potassium chromate (A), well water contaminated by factory effluent (B) or factory effluent (C):			Exposure - 10 min. Contamination of effluent from a chromate and chemical factory examined.	Rajendrababu and Nandakumar, 1987
				Conc. (ug/ml)	% change SDH activity	% change Mg^{+2} ATPase activity		
				A 10	-8.30 ± 0.61	-7.12 ± 0.35		
				50	-24.65 ± 1.2	-9.0 ± 0.65		
				100	-32.28 ± 1.36	-17.11 ± 0.71		
				200	-47.17 ± 2.31	-36.31 ± 1.51		
				500	-59.34 ± 2.71	-46.43 ± 2.31		
				B 15%	-34.23 ± 2.71	-43.62 ± 2.12		
				C 15%	-50.00 ± 2.90	-27.64 ± 1.36		
Potassium Dichromate	- Xenopus laevis	30 per treatment	tadpoles stage 38	Only those kept in concentrations up to 10 ppm developed into tadpoles.			Exposure for 90 days.	Dumpert, 1987
		60 per treatment	eggs	In 5 and 7.5 ppm, 13 ± 5% and 30 ± 15%, respectively, of embryos died; weaker pigmentation in those at 2.5 ppm.				

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Promotryne (H) (Cesagard 50)	- Rana temporaria	500 per treatment	tadpoles just after hatching or after completion of operculum	All younger tadpoles treated with 0.05% suspension for 3 days died within a few days. All older tadpoles exposed to 0.01 or 0.05% suspension died after 24-72 hr.	Static tests.	Jordan et al., 1977
Pyrazophos (F) (Afugan)	- Rana temporaria		tadpoles	1 mg/L - lethal dose. .1 mg/L - stimulation of growth, total inhibition of metamorphosis. - marked change of transaminase enzymatic activity.		Paulov, 1981
Pyrethroids	- Xenopus laevis		myelinated nerve fibres	Closing of the activation gate in a fraction of the sodium channels that open during depolarization delayed, resulting in a prolonged sodium tail current after repolarization of membrane.	Rate of dissociation decreased gradually between 25° - 0°C.	Vijverberg and Ruigt, 1981
	- Assorted species/ review		nervous system	Principle effect is to induce repetitive activity, particularly in sensory nervous system.		Vijverberg and van den Bercken, 1982
Selenium (sodium selenite)	- Xenopus laevis	50-100 per treatment	tadpoles	Tadpoles surviving continuous exposure to 2, 5, 10 ppm had cellular damage, including disorganization and degeneration, in epithelial and muscle cells. Damage more extensive at higher doses.		Browne and Dumont, 1980
	- Xenopus laevis		embryos tadpoles	Concentrations of 2 ppm and above resulted in severe abnormalities and increased mortality. Toxicity increased with increasing concentration up to 20 ppm.		Browne and Dumont, 1979

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT*	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Sodium Diethyldithiocarbamate (F) (NaDEDC)	- Micropsetta aerata		embryos	5 mg/L - highly embryotoxic, growth and development retardation, death in "about 24 hr". 3 mg/L - survivors were "highly retarded" and malformed. After 72 hr - general oedema, abnormal bending of body axis, kinky notochord. 2 and 1 mg/L same effects, less severe. At 0.5 mg/L tadpoles were or less normal - tail slightly shortened and broader.	NaDEDC is potent teratogen. Mainly abnormalities in notochord.	Chuto and Mulherkar, 1980a
Sodium Dodecylbenzenesulfonic Acid	- Xenopus laevis	60 per treatment	eggs	No effect at 0.1,1,3 and 10 ppm. At 30 ppm, eggs did not develop past 2 cell stage.		Dumont, 1987
Sodium Fluoroacetate	- Green frog		cranial nerves	No action potential changes produced by up to 100 mM. 0.01 and 0.10 M produced 3 and 40%, respectively, inhibition of respiration after 3 hr.		Beyerman et al., 1949
Styrene	- Rana pipiens R. temporaria	7	adults	Extensive ultrastructural alterations in olfactory epithelium following exposure to 665 ± 30 ppm for 60 min: - increased secretion from sustentacular cells - oedema fusion of cilia. Reduction in EOG.		Ehlow et al., 1984
Temephos (OP)	- Hyla regilla		tadpoles 3 weeks old	Thermal tolerance significantly lowered:	24 hr exposure.	Johnson, 1980

Conc. (ppb)	Onset of Spasms (°C)	
	Mean	Range
Control (56)	37.9	36.9-37.8
25 (39)	34.2	30.6-37.1
50 (37)	33.9	30.3-36.8

Recommended damage rate for mosquito control in California - 50 ppm.

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
TEPP (OP) (tetraethyl pyrophosphate)	- Rana pipiens	24 in each concentration	adults 9 cm	Following exposure to concentrations of 10, 20, 40, 60, 80 ppm for 15 days: - With increasing concentrations, anemia and leucopenia became progressively more marked - Differential white cell count showed neutropenia and lymphocytosis - Red blood cells distorted in shape - Visceral organs desiccated, spasticity in hindlimbs.		Kaplan and Glazenovich, 1965
	- Bufo viridis	3	adults	Residual cholinesterase at time of death following injection of 2 LD50 into dorsal lymphatic sac:	LD50 - 540 ppm.	Edery and Schatzberg-Porath, 1960
			blood	15%		
			mixed nervous tissue	23%		
2,3,7,8 Tetrachlorodibenzo-p-dioxin (TCDD)	- Rana catesbeiana	15 per group	tadpoles	Dose (ug/kg) / % survival on day 50 post-injection (i.p.): 0 — 80 25 — 87 50 — 73 100 — 93 250 — 80 1000 — 80	All surviving tadpoles successfully completed metamorphosis with no morphological abnormalities.	Beatty et al., 1976
		5 per group	adults 150-250 g	No mortality during 35 day observation at same doses as above. Some lessened food intake in group injected with 500 ug/kg in early phase. No histopathologic lesions were found at any dose level.		

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Thallium (M)	- Frogs		embryos	In M/500,000 tadpoles killed on emergence.		Dilling and Healey, 1926
Thorium (M)	- Frogs		embryos	Development to tadpole stage in M/100,000 was 50%.		Dilling and Healey, 1926
TMBE (tert.-butyl methylether) / anti-detonation preparation)	- Rana temporaria		tadpoles	≤2000 ppm had no lethal effect. 100 ppm in water led to increased weight, stimulated course of metamorphosis.		Paulov, 1987c
Toluene	- Triturus cristatus		haematopoietic system	Exposure via inhalation 5 times/week at 5 ppm or 10 ppm caused anerocytic hypocromic anemia which was more severe at higher concentrations.		Garavini and Soren, 1979
	- Rana pipiens R. temporaria	17	adults	Structural alterations in olfactory mucosa following exposure to 585 ppm for 1 hr: - increased secretion from sustentacular cells. Reduction in COC in frogs exposed to 2000 ppm for 1 hr; no reduction in those exposed to 585 ppm.		Ekblom et al., 1984
Toluene Diisocyanate	- Rana pipiens		erythrocytes	Inhibited isoproterenol- and fluoride ion - stimulated adenylate cyclase activity in a dose dependent manner.	Suggests non-specific inhibition of activity.	McKay and Brooks, 1983

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS			REMARKS	REFERENCE
Toxaphene (OC)	- Rana pipiens		adults 65 g, 8.9 cm	Toxaphene (ppm)	pH of solution at 25° C	No. of frogs dead at 30 days	Frogs placed in 200 cc of test solution.	Kaplan and Overpeck, 1964
		20		0.60	5.83	5	Neuromuscular changes were produced; excessive thrashing and abnormal reactivity to stimulation were observed.	
		10		0.45	5.76	0		
		10		0.30	5.75	0		
Tributyltin (oxide and floride)	- Rana temporaria	10 per treatment	embryos post-gastrula	Survival not affected at 0.3 or 3 ppb; mortality was 40% and 50% at 30 ppb TBTO and TBTF, respectively. Weights declined only at 30 ppb.			Exposure for 5 days.	Laughlin and Linden, 1982
Trichlorphon (OP)	- Rana temporaria	30 per group	blood	Erythrocyte count, haemoglobin level and haematocrit value were decreased following administration of 50, 100, 300 or 3 x 100 mg/kg via dorsal lymphatic sac. 50 and 100 mg/kg decreased leucocyte count; 300 mg/kg caused no change; 3 x 100 mg/kg caused leucocytosis.				Szubertowska, 1979
	- Rana temporaria	30 per treatment	adults	3 hr following injection of 50, 100, 300 or 3 x 100 mg/kg b.w. into dorsal lymphatic sac: - Significant reduction in no. erythrocytes and erythroblasts - Reduction in haemoglobin level and haematocrit value - Changes in no. and % age composition of leucocytes and increased thrombocyte content.				Grzywac - Kalkowska and Szubertowska, 1984
Tri-o-tolyl Phosphate	- Hyla chrysoscelis		embryos	Not toxic or teratogenic at 10 ppm.			Dermally exposed for 24 hr, no. survivors counted 120 hr post-exposure.	Fulton and Chambers, 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Trisodium Nitrilotriacetate (NA_3NTA)	- Leopard frog		tadpoles	No significant mortality occurred in buffered solutions of 600 mg/L or less.	With pH 9.7 or greater, animals died within an hour irrespective of NTA concentration.	Flaningam, 1971
	Salamander sp.		larvae	Survived up to 350 mg/L over 96 hr in buffered solutions.	Complete mortality in unbuffered solutions with pH >9.6.	
Trithion (OP)	- Rana pipiens	24 per concentration	adults	Following exposure to concentrations of 120, 140, 160, 180, 200 ppm for 15 days: - Progressive anemia occurred with increasing concentration - White cell count dropped progressively - Weak response, lymphocytosis - Flaccid paralysis, excessive skin shedding.		Kaplan and Glasscock, 1965
Uranyl Ion (UO_2^{+2})	- Triturus pyrrhogaster		stomach mucous epithelial cells	Almost no effect on the electrical properties of the cell at 10^{-6} M to 10^{-4} M.		Kanno et al., 1978
5-cis-Verbenol	- Rana temporaria		tadpoles	≥15 ppm was lethal.		Poulov et al., 1985

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Zinc (N)	- Gasterophrysa carolinensis	100	eggs	Percent mortality and teratogenesis at hatching (H) and 4 days posthatching (PH) of eggs exposed to sediment-bound zinc:	Hardness - 200 ppm as $CaCO_3$. pH - 7.5 - 8.0. Percent mortality expressed as frequency in experimental population/ controls. * Percentages of survivors bearing gross congenital deformities at hatching.	Birge et al., 1977
				Zn added to sediment (ppm) / Concentration in sediment (ppm) / % mortality: H / PH 0.1 / 104.6 / 6 (3) / 14 1.0 / 112.6 / 3 (0) / 5 10.0 / 124.5 / 7 (1) / 14 100.0 / 222.7 / 7 (2) / 8		
	- Frogs		tadpoles	Few survived long in N/20,000; those which survived in N/50,000 for >3 months were stunted and had no limb buds.		Dilling and Healey, 1926
	- Triturus pyrrhogaster		stomach mucous epithelial cells	10^{-4} N Zn^{+2} decreased membrane potential to 50% of the control value. Effect increased with increasing concentration from 10^{-7} to 10^{-4} N.	Action antagonized by cysteine and acetylpenicillamine.	Kanno et al., 1970
	- Bufo borcas		larvae	39 mg/L Zn^{+2} - all died within 24 hr. 0.1 mg/L - all metamorphosed.	Up to 61 days exposure.	Porter and Hakanson, 1976
	- Xenopus laevis		cylindrical nerve fibre	3.4 mM slowed down kinetics of K system: decreased the permeability constant, increased the time constant, shifted the K activation curve along the potential axis.		Arhem, 1980

Table 2 - Laboratory Studies (cont'd)

CONTAMINANT[a]	SPECIES	N	STAGE/TISSUE	LEVELS USED/EFFECTS	REMARKS	REFERENCE
Zinc Sulphate (see also cadmium chloride)	- Xenopus laevis	20 per test	tadpoles stage 54-58	Mortality (%) of pre-treated (P) and non pre-treated (C) tadpoles:	Pretreatment with 5.0 mg/l Zinc sulphate or 2.5 mg/l cadmium chloride for 96 hr.	Woodall et al., 1980
				Pre-treat./ Exposure Metal / Cd conc. (mg/l) / Time (hr): 15 / 45 / 75 Zn/Zn: 10 P: 0 / 0 / 0; C: 37 / 80 / 80 15 P: 0 / 0 / 0; C: 14 / 33 / 65 20 P: 4 / 15 / 15; C: 44 / 50 / 50 Cd/Zn: 10 P: 0 / 0 / 0; C: 0 / 10 / 10 15 P: 0 / 0 / 0; C: 0 / 15 / 10 20 P: 4 / 5 / 5; C: 12 / 24 / 40		

[a] OC - organochlorine insecticide; OP - organophosphate insecticide; C - carbamate; IGR - insect growth regulator; PY - pyrethroid; A - acaricide; H - herbicide; F - fungicide; M - metal

TABLE 3
FIELD STUDIES

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Acephate (OP) (Orthene)	- Larose Forest, Ontario Mixed wood stands Rana sylvatica, Hyla crucifer, Ambystoma laterale	0.56 kg active ingredient applied by fixed-wing aircraft flying just above canopy; Micronair AV-3000 delivery system.	Caged R. sylvatica tadpoles not affected. No mortality noted in natural populations of 3 species.	- T in ponds after spray - 0.3 L/ha. - T around ponds - 0.2 L/ha.	Buckner and Macleod, 1975
Aldrin (OC)	- 0.025 ha pond in California Rana catesbeiana	Sprays prepared from emulsifiable concentrate formulations and applied at rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) / % 24 hour or cumulative mortality days after treatment 1 / 2 / 5 / 6 / 7 0.11: 10 / 30 / 30 / - / - 0.56: 80 / 100 / * / 0 / - * 10 fresh tadpoles added.	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6° - 35°C. One pond per treatment: N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963
	- Field trials in California Rana catesbeiana	.11 kg/ha. .56 kg/ha.	Deemed 'toxic' to tadpoles.		Mulla, 1962
Aluminum	- Temporary woodland ponds in coastal Maryland Ambystoma maculatum		Embryonic survival negatively correlated with aluminum concentration (range 0.11-0.51 ppm) in water.		Albers and Prouty, 1987
Ambithion (OP) (malathion + fenitrothion)	- Indian rice fields Frog species unspecified	Sprayed by helicopter from height of 2-3 m over fields at 1.0 kg/ha between 7:30 and 9:00 a.m.	90% mortality.	Population estimates made 48 hrs. after spraying.	Thirumurthi et al., 1973

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Aminocarb (C) (Matacil)	- Laurentians, Quebec Balsam fir, black spruce, hazel, cherry, aspen, maple. Grasses: rubus, honeysuckle, wildlily, blueberry Rana pipiens, R. sylvatica, Bufo americana	175 g active ingredient in 2.24 L/ha. 150 ha sprayed June 28, 1979 by fixed-wing aircraft.	R. pipiens and R. sylvatica - no significant change in activity over short term. B. americana - activity decreased during 2 months post-spray possibly related to reduced prey availability.	Data collected July 12 - Aug. 31, 1978 and June 1 - Aug. 31, 1979. 2 sites - 25 km apart.	Bracher and Hider, 1982
	- Richibucto, New Brunswick Rana clamitans asserted other species	Aerially applied at 0.11 kg/ha.	Mortality in caged R. sylvatica tadpoles; no other toxic effects noted.		Rick and Price, 1974
	- Unspecified	0.06 kg/ha.	Adult and larval amphibians were apparently unaffected.		Pearce and Price, 1977
Arco (Larvicide)	- Butte County, California, 2 test ponds - 3.4 x 38.5 m with 4.3 m between them Rana catesbeiana	22.4 L/ha applied using a hand sprayer on June 19, August 3, 24, and September 8, 1971.	No dead frogs were observed after August 24 and September 8 treatments.	Water depth at centre 0.6 m; water temperature 20.5-25.5°C. R. catesbeiana migrated into the ponds prior to the 1rd treatment.	Hagen et al., 1973
Azinphos-methyl (OP) (Guthion)	- 0.025 ha pond Location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
	- Field tests Rana catesbeiana	.45 kg/ha.	"Safe" to tadpoles.		Mulla, 1962

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Bayer 22408 (OP)	- 0.025 ha pond Location unknown *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.56 and 2.24 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
Bayer 29952 (OP)	- 0.025 ha pond Location unknown *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	Application Rate — % 24 hr mortality .11 — 0 .45 — 5	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
Bayer 34042 (OP)	- 0.025 ha pond Location unknown *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1963
Bayer 37289 (OP)	- 0.025 ha pond Location unknown *Bufo boreas* *Scaphiopus hammondi*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
	- Field tests in California *Bufo boreas* *Scaphiopus hammondi*	.11 kg/ha. .45 kg/ha.	Deemed 'safe' at these application rates for tadpoles of both species.		Mulla, 1962
Bayer 38920 (OP)	- 0.025 ha pond Location unspecified *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.56 and 2.24 kg/ha.	Application Rate — % 24 hr Mortality 0.56 — 100 2.24 — 100	Water depth 20.32 - 30.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1963
	- Field tests in California *Rana catesbeiana*	0.56 kg/ha.	Caused 100% mortality to tadpoles.		Mulla, 1962

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Bayer 44831 (OP)	- 0.025 ha pond Location unspecified *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.22 and 0.90 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
Carbofuran (C) (Furadan 3)	- Rice fields, Texas *Acris crepitans blanchardi*	.56 kg/ha carbofuran applied from fixed-wing aircraft 5 to 8 weeks after rice planted. 6 applications.	Frogs found paralyzed or exhibited abnormal behaviour 15 minutes after treatment. Only 1 death recorded.	Study dates - May 2 and July 7, 1970 and 1973-75. Searches conducted 15 mins. - 11 days post-treatment. Rice was 30-50 cm high and in 20-30 cm of water.	Flickinger et al., 1980
Carbophenothion (OP) (Trithion)	- 0.025 ha pond Location unknown *Rana catesbeiana*	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.45 and 1.79 kg/ha.	100% mortality after 24 hours at both rates.	Water depth 20.32 - 30.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1963
	- Field test in California *Rana catesbeiana*	.45 kg/ha.	100% mortality of tadpoles.		Mulla, 1962
Chlordane (OC)	- 0.025 ha pond in California *Rana catesbeiana*	Sprays prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) — % 24 hour or cumulative mortality days after treatment 1 / 2 / 5 / 6 / 7 0.11 — 0 / 0 / 0 / 0 / - 0.56 — 0 / 30 / 30 / 30 / -	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Chlordane (OC) (cont'd)	- Field trials in California Rana catesbeiana	.11 kg/ha. .56 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
Cobalt 60	- Yosemite, California mountain meadow Bufo canorus	Tablets containing 20-30 uc used as radio-tagging device.	3 toads survived 1 year.		Karlstrom, 1957
	B. boreas halophilus	Subcutaneous tag of 40 uc.	Survived.		
DDT (OC)	- Golf course greens Bufo boreas	Hand sprayed: 1.12 kg/ha. 2.24 kg/ha.	95-98% control of juvenile toads after 1 treatment.	Combination DDT and toxaphene.	Mulla, 1962
	- Stream in Algonquin Provincial Park, Ontario Bullfrogs, Green Frogs, Mink Frogs, Wood Frog	Aerially applied on July 21: 6.7 kg/ha.	14 of 22 caged frogs killed within 5 days of spray.	DDT - oil solution.	Logier, 1949
	- Forest in northeastern Ontario Wood Frogs	Up to 4.5 kg/ha.	"Very slight" effects on amphibians.	Formulations of both oil-water emulsions and standard oil solutions used.	Speirs, 1949
	- 64 coastal plain ponds within 20 mile radius of Savannah, Georgia Frog species unespecified	.11-.45 kg/ha DDT applied by rotary duster. 0.6-.22 kg/ha DDT applied by air pressure hand sprayer.	A few frogs were killed by treatment of 0.11 DDT as dust or emulsion. DDT emulsions more toxic than solutions and dusts.	Observations at 24 and/or 48 hours. Weekly sprays.	Terzwell, 1950

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
DDT (cont'd)	- Great Britain Rana temporaria a) Small ditch - 1 m wide Max. depth - 0.4 m b) Larger ditch - 5-6 m wide Max. depth - 1.0 m c) Pool - 11 x 16 m Max. depth - 1.3 m d) 2 marshy pools (20 x 20 m; 4 x 30 m) separated by a bank Max. depth - 0.5 m and 0.1 m respectively e) Gravel pit - 25-30 x 140 m Max. depth - 2.0 m	0.4-0.5 kg/ha applied on water surface in a 2 m wide band around water edge by hand operator.	Site/Cage / Behaviour of exposed tadpoles: Day 1 / Day 3 / Day 14 a)/ 1) Frantic* / Resigned / No survivors 2) Frantic / Some frantic, Some resigned / No survivors b)/ 1) Some normal, Some frantic / Few normal, Most frantic, Few resigned / Four normal, One moribund 2) Frantic / Most frantic, Few resigned / Thirteen normal, Three moribund c)/ 1) Frantic / Frantic / Normal 2) Frantic / Frantic / Normal * Frantic behaviour is characterized by incoordinated hyperactive activity.	Distance of caged tadpoles to bank: a) cage 1 central; cage 2 central b) cage 1 1 m; cage 2 1 m c) cage 1 2 m; cage 2 1 m d) cage 1 4 m; cage 2 1 m e) cage 1 5 m; cage 2 7 m All survivors on day 3 had abnormal snouts at site a. No behavioural or morphological abnormalities noted at site d or e.	Cooke, 1971a

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
DDT (cont'd)	- Hubbard County, Minnesota Two small woodland pools Rana sylvatica	Aerial application.	2 and 33 hours after spray - frogs were as numerous and activity was normal. 2½ days after spray - dead frogs found on each pool. On June 1 and July 2 no live frogs were found.	May 21 date of spraying. Tent caterpillar larvae dropped into ponds, covering the surface. All but two frog stomach samples (n=34) contained caterpillars; not known if mortality caused by contact or accumulation of contaminated prey.	Fashingbauer, 1957
	- Four open ponds in Florida Rana pipiens R. grylio Hyla c. cinerea	Aerial application: 1 - 5% DDT at .34 kg/ha; 2 - 5% DDT at .67 kg/ha; 3 - 20% DDT at .34 kg/ha; 4 - 20% DDT at .67 kg/ha.	2 of 3 frogs from #4 returned to lab for observation (R. pipiens, R. grylio) showed lack of coordination symptoms. The third (H. cinerea) showed no symptoms. R. grylio died after 4 days; R. pipiens recovered in 10 days.	Canopy cover dispersed the spray.	Herald, 1969
	- 0.025 ha pond in California Rana catesbeiana	Sprays prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) / % 24 hour or cumulative mortality days after treatment 1 / 2 / 5 / 6 / 7 0.11: 0 / 0 / 0 / - / - 1.121: 30 / 80 / * / 0 / - * 10 fresh tadpoles added.	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6° - 35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Dichlobenil (H)	- 5 m diameter pond in Great Britain Rana temporaria Bufo bufo Triturus vulgaris	1 mg/L granules.	No deaths or changes in activity or development rate within 32 days. T. vulgaris continued to breed.	Water depth 1-2 m in winter; 1 m in summer. Frogs and toads were caged; newts were free living.	Cooke, 1977
Dieldrin (OC)	- 0.025 ha pond Rana catesbeiana	Sprays prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) / % 24 hours or cumulative mortality days after treatment 1 / 2 / 5 / 6 / 7 0.11: 100* / 30 / 30 / 30 / - 0.56: 100* / 100 / * / 30 / 30 * 10 fresh tadpoles added.	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6° - 35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963
	- Pond in California Rana catesbeiana	.11 - .56 kg/ha.	Complete tadpole kill.		Mulla, 1962
Diquat (H)	- 5 m diameter pond in Great Britain Rana temporaria Bufo bufo Triturus vulgaris	1 mg/L sprayed.	No deaths or changes in activity or development rate within 32 days. T. vulgaris continued to breed.	Water depth 1-2 m in winter; 1 m in summer. Frogs and toads were caged; newts were free-living.	Cooke, 1977
Endosulfan (OC)	- Savanna woodland, Zimbabwe Chiromantis xerampelina	Ultra-low volume aerial applications from a fixed-wing aircraft at 14 g a.i/ha.	No deaths 19 days post-treatment.	N = 14 treatment, 8 control; animals kept in cages in pond.	Cockbill, 1979

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians						Water Temperature, Residue Levels, Miscellaneous	Reference
Endrin (OC)	- 0.025 ha pond in California Rana catesbeiana	Sprays prepared from emulsified concentrate formulations at a rate of 89.9 L/ha and applied from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha)	% 24 hours or cumulative mortality days after treatment					Water depth 28.32 - 10.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-15°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Wallis, 1963
				1	2	5	6	7		
			0.11	10	90	90*	0	0		
			0.56	100*	90	100*	90	20		
			* 10 fresh tadpoles added.							
	- Field tests in California Rana catesbeiana	.11 - .56 kg/ha.	Complete kill of tadpoles at these application rates.							Wallis, 1962
	- 2 year old artificially evacuated pool (10 x 10 m) with clusters of filamentous algae; no inlet Rana temporaria R. esculenta	0.5 L of 20% Endrin poured into pool. 0.047 mg/L measured 10 m from point of contamination after 6 hours.	Adult R. esculenta (wild population) tossed about because of strong cramps and paralysis of limbs and died. Caged R. temporaria tadpoles (without developed limbs) alive only at most distant test stations after 72 hours. 7 weeks after treatment, R. esculenta repopulated pond.						Average water depth 0.44 m. Temperature 26°C, sky 2/3 cloud cover, sunny breaks.	Mehlquenath and Fenhave, 1979
Ethyl Guthion (OP)	- 0.025 ha pond Location unknown Rana catesbeiana	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.22 and 0.90 kg/ha.	No mortality after 24 hours.						Water depth 28.32 - 10.48 cm. 10 tadpoles caged in pond.	Wallis et al., 1963

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Fenitrothion (OP)	- Fundy National Park, New Brunswick Rana clamitans R. sylvatica unsorted other species	Aerially applied at 0.14 kg/ha in 2 sprayings 9-17 days apart.	No mortality observed in caged R. sylvatica tadpoles, or in caged or wild adults or tadpoles of other species.		Rich and Morse, 1974
	- Pond in New Brunswick Rana clamitans Bufo americanus	Aerially sprayed with .14 kg/ha on June 1 and 9.	No mortality observed.		Peakin and Forster, 1969
	- Indian rice fields Frog species unspecified	Sprayed by helicopter from height of 2-3 m over fields at 0.5 kg/ha between 7:10 and 9:00 a.m.	100% mortality.	Population estimates made 48 hours after spraying.	Thirumurthi et al., 1973
Fenthion (OP)	- Indian rice fields Frogs species unspecified	Sprayed by helicopter from height of 2-3 m over fields at 0.5 kg/ha between 7:30 and 9:00 a.m.	100% mortality.	Population estimates made 48 hours after spraying.	Thirumurthi et al., 1973
FLIT MLO (Larvicide)	- Butte County, California 2 test ponds - 3.4 x 38.5 m with 4.3 m between them Rana catesbeiana	2.24 L/ha applied using a hand sprayer on June 15, August 3, 24, and September 8, 1971.	No dead frogs were observed after August 24 and September 8 treatments.	Water temperature 20.5-25.5°C. Water depth at centre 0.6 m. R. catesbeiana migrated into the ponds prior to the 1st treatment.	Hagen et al., 1971
Fuel Oil N°2 (Larvicide)	- Butte County, California 2 tests ponds - 3.4 x 38.5 m with 4.3 m between them Rana catesbeiana	112.1 L/ha applied using a hand sprayer on June 15, August 3, 24, and September 8, 1971.	No dead frogs were observed after August 24 and September 8 treatments.	Water temperature 20.5-25.5°C. Water depth at centre 0.6 m. R. catesbeiana migrated into the ponds prior to the 1st treatment.	Hagen et al., 1971

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
G-27345 (OP)	- 0.025 ha pond Location unknown Rana catesbeiana	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.22 and 0.90 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 10.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1961
G-28029 (OP)	- 0.025 ha pond Location unknown Rana catesbeiana	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.45 and 1.79 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 10.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1961
G-30493 (OP)	- 0.025 ha pond location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.22 and 0.90 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 10.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1961
	- Field tests in California Bufo boreas Scaphiopus hammondi	0.22 kg/ha. 0.90 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
G-30494 (OP)	- 0.025 ha pond Location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	Application Rate: .11 — % 24 hour mortality: 0 Application Rate: .45 — % 24 hour mortality: 1	Water depth 20.32 - 10.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1961
	- Field tests in California Bufo boreas Scaphiopus hammondi	.11 kg/ha. .45 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
OC-3582 (OP)	- 0.025 ha pond Location unknown Rana catesbeiana	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.45 and 1.79 kg/ha.	Application Rate: .45 — % 24 hour mortality: 0 Application Rate: 1.79 — % 24 hour mortality: 100	Water depth 20.32 - 10.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1961

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
OC-3582 (cont'd)	- Field tests in California Rana catesbeiana	.45 kg/ha. 1.8 kg/ha.	No mortality in tadpoles. 100% mortality in tadpoles.		Mulla, 1962
Heptachlor (EC)	- 0.025 ha pond in California Rana catesbeiana	Sprays prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) — % 24 hour or cumulative mortality days after treatment: 1, 2, 5, 6, 7 0.11: 0, 0, 0, 0, - 0.56: 10, 80, 60*, 0, 0 * 10 fresh tadpoles added.	Water depth 20.32 - 10.48 cm. pH 7.5 - 8.0. tests conducted from June to October 1962. Maximum daily water temp. 26.0°-35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1961
	- Field tests in California Rana catesbeiana	.11 kg/ha. .56 kg/ha.	Deemed 'toxic' to tadpoles at these application rates.		Mulla, 1962
Kepone (OC)	- 0.025 ha pond in California Rana catesbeiana	Spray prepared from emulsifiable concentrate formulations at a rate of 89.9 L/ha and applied from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) — % 24 hour or cumulative mortality days after treatment: 1, 2, 5, 6, 7 0.11: 0, 0, 0, -, - 0.56: 0, 0, 0, -, -	Water depth 20.32 - 10.48 cm. pH 7.5 - 8.0. tests conducted from June to October 1962. Maximum daily water temp. 26.0°-35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1961

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Kepone (cont'd)	- Field studies in California Rana catesbeiana	.11 kg/ha. .56 kg/ha.	Deemed 'safe' to tadpoles at these two application rates.		Mulla, 1962
Leptophos (OP)	- Indian rice fields Frog species unspecified	Sprayed by helicopter from height of 2-3 m over fields at 1.0 kg/ha between 7:30 to 9:00 a.m.	100% mortality.	Population estimates made 48 hours after spraying.	Thirumurthi et al., 1973
Lindane (OC)	- 0.023 ha pond in California Rana catesbeiana	Spray prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.49 L) sprayers.	Application Rate (Active Ingredient) (kg/ha) / % 24 hour or cumulative mortality days after treatment 1 / 2 / 5 / 6 / 7 0.11: 0 / 0 / 0 / 10 / - 0.56: 10 / 10 / 10 / 10 / -	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963
	- Field tests in California Rana catesbeiana	.11 kg/ha. .56 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
Malathion (OP)	- Oak - poplar forest, North Carolina Plethodon glutinosus	5.6 kg/ha applied with back pack sprayer during 10 weekly applications (27 May to 23 August).	No brain ChE inhibition, changes in abundance, effects on lipid storage patterns noted in adults or juveniles.	ChE inhibition noted at this concentration in lab study.	Baker, 1981

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Metals	- Ironworks Katowice region, Poland Rana ridibunda	Emissions from metallurgic work.	Frogs collected from an area close to the factory had lower metabolic rates than those from farther away.		Pytasz et al., 1982
Methyl Parathion (OP)	- 0.023 ha pond Location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	No mortality after 24 hours.	Water depth 20.32-30.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1963
	- Field tests in California Bufo boreas Scaphiopus hammondi	.11 kg/ha. .45 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
Mexacarbate (C) (Zectran)	- Acadian Forest Experiment Station, New Brunswick Rana clamitans, R. sylvatica Assorted other species	Aerially applied at 0.087 kg/ha.	Mortality in R. sylvatica tadpoles; no other toxic effects noted.		Dick and Price, 1974
Naled (OP)	- 0.023 ha pond Location unknown Rana catesbeiana	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.56 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 10 tadpoles caged in pond.	Mulla et al., 1963

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Dimevi (C)	- Ditches beside fields in Great Britain Cage 1 - between barley and potato fields Cage 3 - between barley and sugar beet fields Cage 7 - between sugar beet and potato fields Cage 17 - control pond Rana temporaria	Unspecified.	Cage 1: Development and growth slow, with many deformities, mortality. Cage 3: Development rapid, growth satisfactory, survival good. Late in experiment tadpoles developed 'lateral kinks'. Cage 7: High mortality during first month, growth and development normal. Deformities after 6 weeks. Cage 17: Survival good, development and growth average. 'Lateral' kinks noted late in experiment.	Caged tadpoles maintained near potato fields showed more deformities. No other fields were treated with dimevi.	Cooke, 1981
Parathion (EP)	- 0.089 ha pond Location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.11 and 0.45 kg/ha.	No mortality after 24 hours.	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Halle et al., 1961
	- 2 wetland habitats in forest areas of California: Habitat 1 - duck pond (4.05 ha) with watergrass, smart weed, rushes. Existing vegetation burned in July before pond flooded. Habitat 2 - borrow pit (18.1 ha) filled with underground seepage water. Vegetation included cattails, watergrass, hornwort. Rana catesbeiana	Applied by aircraft: Habitat 1 - sprayed once at a rate of 1.12 kg a.i./ha from height of 7.62 m. Habitat 2 - sprayed 6 times at weekly intervals at a rate of .11 kg/ha from height of 12.2 m.	Habitat 1 - abundance or survival not seriously effected. Habitat 2 - no apparent effect on populations.	Habitat 1 - 25.4 cm in depth. Habitat 2 - 1.83 m in depth. Average amount of parathion which reached water surface: Habitat 1 - .95 kg/ha; Habitat 2 - 0.38-0.88 kg/ha.	Halle et al., 1966

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Parathion (cont'd)	- Field tests in California Bufo boreas Scaphiopus hammondi	.11 kg/ha. .45 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Halle, 1962
Pesticides (organochlorine)	- Rice field discharge canal and relatively clean pond in the North Caucasus, U.S.S.R. Rana ridibunda	Wide range of pesticides used on fields.	Haemoglobin content increased, leucocytosis observed and number of stab-nuclear neutrophils and monocytes in the leucocytic formula of the blood increased in frogs from rice field.	8-9 times greater organochlorine levels in muscles of frogs from rice fields.	Zhukova, 1967
	- Water sources polluted with pesticides Bufo melanostictus		Higher sister chromatid exchange frequency noted in field specimens than controls exposed identically to BrdU solution.	Attributed to high concentration of mutagens/clastogens in tissues of field specimens.	Chakrabarti et al., 1986
pH	-Natural breeding ponds Ambystoma maculatum		Hatching success at pH 4.6 - 0.0%. Hatching success at pH 6.9 - 77.1%.	Eggs interchanged between acidic and neutral ponds had intermediate hatching success. Tolerance to acid is built up upon exposure.	Nicholson et al., 1977
	-Sandy coastal dunes and inland heaths in Great Britain Bufo calamita		B. calamita avoided acid heathland ponds (pH > 5.0).	Fluctuating temperature, light; no rainfall. Spawn taken and reared in lab: no tadpoles survived to metamorphosis below pH 4.75 and success was minimal below 6.0.	Beebee and Griffin, 1977

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (cont'd)	-Southeast England agricultural land and uncultivated heathland Triturus vulgaris, T. helveticus, T. cristatus, Rana temporaria, Bufo bufo, B. calamita		pH 4-5 / 5-6 / 6-7 / 7-8 T. vulgaris 0* / 0 / 5 / 1 T. helveticus 2 / 2 / 7 / 1 T. cristatus 0 / 0 / 2 / 0 R. temporaria 0 / 1 / 1 / 6 B. bufo 0 / 0 / 4 / 1 B. calamita 0 / 1 / 1 / 0 TOTAL PONDS 13 / 4 / 12 / 7	Survey of amphibian breeding sites. * No. of ponds in each category containing the various species.	Beebee, 1981
	-Stream draining a construction site in Great Smoky Mountains National Park, North Carolina Leurognathus marmoratus		None of the caged salamanders placed upstream (X pH = 7.0) died; most of those placed downstream (X pH = 4.6) died.	Water temperature downstream - 9-10°C.	Huckabee et al., 1975
	-Ephemeral meltwater ponds in central Ontario Ambystoma maculatum		Percent hatching success was correlated with pH - 88% at pH 6.15, 80% at pH 4.51. Survival of larvae in pond of pH 4.97 was 66%; in enclosures where $CaCO_3$ added to elevate pH to 5.30, survival was 64%.	Results of transfer experiments suggest that larvae from population breeding in acidic ponds were more tolerant of increased acidity than those from less acidic ponds.	Clark, 1984b
	-Temporary woodland ponds in coastal Maryland Ambystoma maculatum		Egg mass abundance and survival of embryos not correlated with pond pH. Survival of eggs transferred among ponds of pH 3.66 - 5.10 reduced only at pH 3.66.	Pond longevity, water temperature and perhaps oxygen content seem more important to salamander reproduction than acid precipitation.	Albers and Proudy, 1987

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (cont'd)	-Bog pools at a heathland site in south-east England Triturus helveticus		Bred in all pools; were rarest in rain-fed ones.	Eutrophicated ponds had pH 5.0 - 6.5; rain-fed ponds had pH 3.8 - 4.3.	Beebee, 1987
	Bufo calamita		Bred only in eutrophicated ponds, as did 4 other unspecified amphibian species.		
	-Ephemeral meltwater ponds in central Ontario Bufo americanus, Rana catesbeiana, Hyla crucifer, R. sylvatica, R. clamitans		B. americanus, H. crucifer and R. catesbeiana did not occur in ponds with pH > 5.0. R. clamitans and and R. sylvatica occured in all ponds. R. sylvatica densities increased in more acidic ponds; egg mass density was reduced as pH decreased.	Surveys done in ponds with pH 3.66 - 6.63. Results suggest chronic sublethal effects may be affecting populations at pH's greater than those causing acute toxicity.	Clark, 1984a
	-Forest and fen sites in Britain Triturus helveticus T. vulgaris		T. vulgaris rarely encountered in water with pH < 4. T. helveticus rarely encountered in water with pH < 4.	Breeding site characteristics studied.	Cooke and Frazer, 1976
	-Temporary forest ponds in central Pennsylvania and sphagnus bogs, seeps and ponds in New Jersey Ambystoma jeffersonianum Rana sylvatica Bufo woodhousei Hyla andersoni		A. jeffersonianum and B. woodhousei not observed in ponds with mean pH < 4.42 and 4.25, respectively; presence of R. sylvatica and H. andersoni unrelated to pond pH. In embryo transfer experiments, mortality of A. jeffersonianum embryos increased as pH decreased; hatching of R. sylvatica was not related to pH.		Franz and Dunson, 1984

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (cont'd)	-Poorly buffered water bodies in the Netherlands Bufo bufo B. calamita Rana arvalis R. esculenta R. temporaria Triturus alpestris T. cristatus T. helveticus T. vulgaris		Relation between water pH and % systems with amphibians: pH 5 (n=37): 88.6, 13.3, 21.9, 76.6, 48.8, 9.1, 8.1, 21.6, 18.9 Δ pH 5 (n=31): 97.5, 3.0, 62.5, 76.9, 55.0, 9.7, 6.5, 32.3, 6.5 pH 6 (n=28): 11.8, 0.0, 35.3, 61.3, 17.7, 0.0, 0.0, 10.7, 0.0 At many spawning sites of R. arvalis, R. esculenta, R. temporaria and B. bufo no tadpoles found in pH > 5.0.	In acidic waters, egg masses of R. arvalis, R. esculenta, R. temporaria, B. bufo, B. calamita became heavily infested with fungi.	Leuven et al., 19-
	- Laurentians, Québec Rana sylvatica	Snowmelt water.	Egg mass density was negatively correlated with acidity. Hatching success was inversely correlated with pH. Occurrence of mould on eggs increased in low pH.	Egg mass density also correlated with total organic carbon.	Casson and Planas,

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (cont'd)	-Hardwood-hemlock forests of Delaware Co., New York Plethodon cinereus		8.8% of quadrats with soil pH > 3.7 contained a salamander; 50.0% of quadrats with pH < 3.8 contained a salamander. Juveniles never found on soil with pH > 3.7. Seasonal density correlated with % quadrats with pH > 3.7.	Soil temperature and moisture did not affect density or distribution.	Hoven and Hawksley-Lescault, 1981
	-Temporary forest ponds in central Pennsylvania Rana sylvatica		Tadpoles from pond with pH 4.5-4.9 had lower body sodium, chloride and water concentrations than those from nearby pond with pH 5.7a - 6.3?. Tadpoles from either pond placed in low pH had higher sodium efflux than when placed in high pH.		Frada and Duncan, 1981c
pH (acid precipitation)	-5 ponds in Tompkins Co., New York Ambystoma maculatum	Acid precipitation.	Pond: A, B, C, D, E pH: 7, 6, 5.5, 5.0, 4.5 Maximum mortality %: 0.66, 0.91, 43.7, 63, 63 Mortality low during early embryonic development. Abnormalities observed at low pH.	Altitude 460 - 625 m. pH 6.5-7.0 at time eggs laid; by hatching pH had increased by 0.25 to 0.5 units in each pond. Water temperature rose from 8-23°C during development.	Pough, 1976
	- Lake with sphagnum bottom, Sweden Rana temporaria	Lake acidified by acid precipitation.	No embryos survived to become adults.	pH 4.0-4.5.	Hagstrom, 1977

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (acid precipitation) (cont'd)	- Lake Tranevatten, Sweden Triturus vulgaris Bufo bufo, Rana temporaria	acid precipitation.	T. vulgaris increased in acidified lakes; R. temporaria and B. bufo decreased.	Suggests that newts increased as a result of extinction of predatory fish populations.	Hagstrom, 1981
	- Ponds in Ithaca, NY Ambystoma maculatum	pH range of 4.5-7.0 in ponds.	pH / Mortality of embryos 7 — 0.6% 6 — 0.9% 5.5 — 43.7% 5.0 — 65% 4.5 — 65%	Eggs from ponds with pH 4.5, 5.0 failed to retract yolk plug; eggs in pH 5.5 developed cheat swellings and stunted gills after gastrulation.	Tome and Pough, 1982
			Reciprocal egg transplants indicated no difference in acid tolerance dependent on pond of origin.	Some of this data appeared in Pough 1976.	
	- Connecticut Valley, Massachusetts Ambystoma maculatum A. jeffersonianum	Rainfall: pH - 4.16 in 1976. pH - 4.07 in 1977.	No significant correlation between pond pH and % embryonic mortality. A. maculatum 2.1-10.2% mortality. A. jeffersonianum 12.4-40.1% mortality.	pH of 6 ponds: 1976 $\bar{x}$ = 5.42. 1977 $\bar{x}$ = 5.10.	Cook, 1983

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
pH (bogwater)	- Bear Meadows Bog, Pennsylvania sphagnum bog	Embryos were transferred to a naturally acidic stream draining the bog.	Mortality of transferred embryos:	Maintained in lab at 25°C before exposure. Lab tests using bog water support findings in the field. Observed lethality at pH's of 4.0, thus possibly some other toxic agent is responsible.	Saber and Dunson, 1978
			pH range / Temp. range (°C) / N animals (Stage) / % mortality 3.80-3.82 / 14.5-19 / 20 / 100 3.80-3.82 / 14.5-19 / 20 / 100 4.00-4.01 / 18 / 30 / 100 14-19 / 10(26) / 100 3.72-3.99 / 14-19 / 6(32-36) / 16.7 3.78-3.99 / 14-19 / 5(26-30) / 0 3.60-4.0 / 14-19 / 6(26-30) / 83.3		
	Rana catesbeiana R. clamitans Bufo sp.			Several locally common species were absent from bog.	
Phosphamidon (OP)	- Indian rice fields Frog species unspecified	Sprayed by helicopter from height of 2-3 m over fields at 0.25 kg/ha between 7:30 and 9:00 a.m.	75 % mortality.	Population estimates made 48 hrs. after spraying.	Thirumurthi et al., 1973
Pollution (unspecified)	- Minsk and Berezinsky Reserve, U.S.S.R. Rana temporaria	"anthropogenic" pollution.	No differences in chromosome aberration frequencies in frogs in two regions.	Regions subjected to different degrees of anthropogenic impact.	Eliseyeva et al., 1985
	- South Sakhalin, U.S.S.R Rana chensinensis	Site A - polluted with sewage effluent of paper factory. Site B & C - contaminated by municipal gutters.	Site / No. with Limb Abnormalities / No. with Chondrodysplasia Lesions A — 448 (42%) — 136 (11.5%) B — 1424 (39%) — 202 (5.5%) C — 500 (31%) — 0	Developmental anomalies and dysplasia also found in R. amurensis, Bufo bufo gargarizans	Mikgirnov et al., 1984

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Pollution (industrial waste products)	- Shanghai, China Bufo bufo gargarizans	Waste water treatment plant and factory oxidation pond.	Sister chromatid exchange frequencies of samples were markedly higher than frequencies of controls.		Wan et al., 1984
	- Rana temporaria		Level of genetic damage in frogs from polluted regions was higher than in those from protected areas.		Krasnowski et al., 1986
Pollution (sewage)	- Playa Lake, Texas Ambystoma tigrinum	Polluted with sewage and asphalt wastes.	Hepatic microsomal aryl hydrocarbon hydroxylase levels were elevated. Bladder contents were mutagenic using Ames test.		Baubee et al., 1978
Ronnel (OP,A)	- 0.023 ha pond Location unknown Bufo boreas Scaphiopus hammondi	Applied as aqueous spray at rate of 89.9 L/ha. Active ingredient = 0.22 and 0.90 kg/ha.	Application Rate: .22 — % 24 hr mortality: 10 Application Rate: .90 — % 24 hr mortality: 0	Water depth 20.32 - 30.48 cm. 20 tadpoles caged in pond.	Mulla et al., 1963
	- Field tests in California Bufo boreas Scaphiopus hammondi	.22 kg/ha. .90 kg/ha.	Deemed 'safe' to tadpoles at these application rates.		Mulla, 1962
Rotenone	- Shallow .05 ha ponds in Georgia Rana pipiens	2 and 5 uL/L of Pro-Noxfish (0.05 and 0.125 uL/L rotenone) applied on Aug. 24 into propeller wash of boat.	Partial mortality of resident population of larval frogs in 5 uL/L treatment.	Water temp. 31-32°C, pH 8.6-9.6 at time of treatment.	Burress, 1982

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians	Water Temperature, Residue Levels, Miscellaneous	Reference
Thiodan I (OC)	- 0.023 ha pond in California Rana catesbeiana	Sprays prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 L/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) — % 24 hour or cumulative mortality days after treatment: 1: 0.11 = 60*, 0.56 = 100*; 2: 0.11 = 20, 0.56 = 80; 5: 0.11 = 0, 0.56 = 0; 6: 0.11 = -, 0.56 = 0; 7: 0.11 = -, 0.56 = - * 10 fresh tadpoles added.	Water depth 20.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-35°C. One pond per treatment; N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963
	- Pond Rana catesbeiana	.11 kg/ha applied by hand sprayer.	Complete kill of tadpoles.		Mulla, 1962
	- Residential area Bufo boreas Scaphiopus hammondi	1.12 kg/ha.	98% control of juvenile toads in 24 hrs.		Mulla, 1962
Thiodan II (OC)	- 0.023 ha pond in California Rana catesbeiana	Spray prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 kg/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active ingredient) (kg/ha) — % 24 hour or cumulative mortality days after treatment: 1: 0.11 = 10, 0.56 = 100*; 2: 0.11 = 10, 0.56 = 30; 5: 0.11 = 10, 0.56 = 30; 6: 0.11 = -, 0.56 = -; 7: 0.11 = -, 0.56 = - * 10 fresh tadpoles added.	Water depth 20.32 - 30.48 cm pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-35°C. One pond per treatment, N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963

Table 3 - Field Studies (cont'd)

Contaminant[a]	Location/Habitat Species	Rate and Method of Application	Reported Effects on Amphibians						Water Temperature, Residue Levels, Miscellaneous	Reference
Toxaphene (OC)	- 0.025 ha pond in California *Rana catesbeiana*	Spray prepared from emulsifiable concentrate formulations and applied at a rate of 89.9 kg/ha from 3 gallon (12.45 L) sprayers.	Application Rate (Active Ingredient) (kg/ha)	% 24 hour or cumulative mortality days after treatment					Water depth 28.32 - 30.48 cm. pH 7.5 - 8.0. Tests conducted from June to October 1962. Maximum daily water temp. 26.6°-35°C. One pond per treatment, N = 20 caged tadpoles (age unspecified) per pond.	Mulla, 1963
				1	2	5	6	7		
			0.11	0	0	0	-	-		
			0.56	100*	100	0	0	-		
			* 10 fresh tadpoles added.							
	- Golf course greens *Bufo boreas* *Scaphiopus hammondi*	1.12 kg/ha. 2.24 kg/ha.	95-98% control of juvenile toads.						In mixture with DDT.	Mulla, 1962
	- Field tests in California *Rana catesbeiana*	.11 kg/ha. .56 kg/ha.	100% kill of tadpoles.							Mulla, 1962

[a] OC-organochlorine; OP-organophosphate; C-carbamate; H-herbicide; A-acaricide.

TABLE 4
RESIDUES

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Acephate (OP)	- Rana clamitans/lab		9-10 per pool	Tadpoles	Approximately 0.60 after 1 day (identical to concentration in water).	Exposed to 1 ppm.	Lyons et al., 1976
	Rana sylvatica/ Larose Forest, Ontario	June 1975		Tadpoles	0.27 after 1 hour; 0.12 at day 1 (similar to water concentration).	Aerially sprayed at rate of 560 g a.i./ha.	
Acridine	- Xenopus laevis/lab		5 per 200 ml test solution	Larvae	Maximum concentration in larvae reached 85 ppm wet weight in 60-90 mins. Elimination, when larvae placed in clean water, was rapid-acridine was not detectable after 2 hrs.	Larvae exposed to 5.0 mg/L. Acridine animal/ Acridine water = 170.	Hayes et al., 1981
Aldrin (OC)	- Bufo americanus/Missouri corn fields	1963-67		Whole body	Sample: Wet (g) 10.00; Dry (g) 2.17; Lipid (g) 0.0930 Residues: Aldrin 0.83; Dieldrin 1.37; Range A + D 1.25-1.58; DDT DDE DDD 0.13 Sample: Wet (g) 10.01; Dry (g) 1.75; Lipid (g) 0.2192 Residues: Aldrin -; Dieldrin 4.60; Range A + D 2.31-8.30; DDT DDE DDD 0.07	2 fields treated with 1.121 kg/ha for 15 and 16 of the past 17 years. Young toads (4 pooled samples of 10 tadpoles each) in August contained three times the residues of adults specimens collected in June.	Korschgen, 1970
			2 40	Adults Young			
	- Rana pipiens/lab			Ventral abdominal skin	Decreasing rate of accumulation by skin vs. time.	Pithed frogs were suspended in solution of 11 ppb.	Kaiser and Dunham, 1972-73

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Azamerark (C)	- Québec	1984		Larvae	Days Post-Spray / Stage / Residue: 0.25 / 25-31 / .109 - .128 1.25 / 26-31 / .023 - .025 3.0 / 28-35 / .022 - .026 9.0 / 28-36 / .007 14.0 / 31-40 / < .005 0.25 / 31-32 / .022 12.0 / 29-37 / < .005	Aerially treated with 87.5 g a.i./ha. No metabolites detected.	Mawarbachi et al., 1987
	Bufo americanus		3 3 2 3 3				
	Rana sylvatica		3 1				
Atrazine (H)	- Rana catesbeiana/ponds in Kansas	1973-74		Tadpoles	Residues (ppm) of atrazine during 1973 and 1976:	Application with 0.1 ppm both summers.	Klaassen and Kadoum, 1979
					1973 Days post treatment: 0 / 1 / 22 / 55 / 120 ND[a] / ND / - / - / -	ND - less than 0.4 ppb.	
					1976 Days post treatment: 0 / 2 / 23 / 91 / 85 0.289 / 0.278 / 0.309 / 0.255		
		1973			Residues (ppm) of atrazine and carbofuran at given intervals post-treatment:		
					Pre-treatment: Atr. ND; Carb. ND 3 days: Atr. 0.276; Carb. ND 21 days: Atr. 0.289; Carb. ND 77 days: Atr. 0.207; Carb. ND	Application with 0.1 ppm atrazine and 0.025 ppm carbofuran. ND - less than 0.4 ppb.	

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Atrazine (cont'd)		1973			Residue (ppm) of atrazine and carbofuran at given intervals post-treatment: Pre-treatment: Atr ND, Carb ND; 2 days: Atr 0.277, Carb ND; 23 days: Atr -, Carb -; 54 days: Atr -, Carb -; 91 days: Atr -, Carb -	Application with 0.3 ppm atrazine and 0.050 ppm carbofuran. *ND-less than 0.4 ppm. No bio-magnification of atrazine was observed.	Klaassen and Kadoum, 1979 (cont'd)
Benzene Hexachloride (BC) (BHC)	- Rana catesbeiana R. sphenocephala R. C. clamitans/ Oxbow lakes in Louisiana	1980	5 5 5		Not detected in all three species.	Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 140 cm.	Mathhammer et al., 1984
Benzo(a)pyrene	- Pleurodeles waltl/lab			Larvae	Ratio of BaP in larvae to that in surrounding water after 12 hours was approximately 200; not dependent on dose; maximal levels attained after 12 hours. Contaminated larvae placed in uncontaminated water lost 99% contaminant after 100 hrs.	Dose levels between $0.075 - 0.3 \times 10^{-3}$ ppm tested.	Grinfeld et al., 1986
Cadmium (II)	- Rana temporaria/lab			Liver, kidney, skin	Subcutaneous injections of 0.12-0.24 mg/100 g/day for 10 days resulted in significant accumulation in liver and kidney. Within 10 days of exposure to >0.0075 $CdCl_2$ in the aquatic environment, significant amounts Cd found in skin; small amounts found in liver and kidney.		Vasil'eva et al., 1987

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Cadmium (cont'd)	- lab Rana japonica R. nigromaculata		3-5 per dose	Liver Kidney Liver Kidney	Concentrations (mean ± SD): Cd: 33.5 ± 21.0*; 65.7 ± 27.1*; Cu: 175 ± 24; 9.04 ± 2.99 Cd: 53.9 ± 33.6*; 96.8 ± 22.5*; Cu: 48.2 ± 19.9*; 10.8 ± 3.5 * Significantly different than controls.	0.225 Cd/kg body wt. injected i.m. 11 times during 15 days; frogs killed 2 days after last injection. No effect on concentrations of 9 other elements.	Suzuki et al., 1986
	- Plethodon cinereus, P. glutinosus, Desmognathus ochrophaeus, D. wrighti, Rana sylvatica/oak forest 10 km upwind from 2 zinc smelters in eastern Pennsylvania	June - Oct., 1979	23	Whole body	Mean ± SE (ppm dry weight) for all species: 1.4 ± 0.1.		Beyer et al., 1985
	- Unspecified species/ New Lead Belt, Southeastern Missouri	Sept. 1972	8 pools 1 pool	Tadpoles	Area: Strother Creek; Control Residues (ppm dry wt.): 1.6-3.0; 1.1	Dam across Strother Creek provides the principal tailings pond for mine and mill effluents. Samples taken from below dam.	Gale et al., 1973

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Cadmium (cont'd)	- Rana catesbeiana/ lead mining district, southeastern Missouri	1981-82	15	Whole body	Residues (geometric mean, range in parentheses):	Site A - upstream from lead belt, sites B and C - downstream from tailings ponds.	Niethammer et al., 1985
					Site A: not detected; Site B: 0.31* (nd-0.89); Site C: 0.26* (nd-0.64)		
					* Significantly higher than Site A.		
	- Patuxent Wildlife Research Centre					Collected from uncontaminated areas. Some of Bufo spp. were captured on or near highways.	Hall and Mulhern, 1984
	Rana catesbeiana		5 pools	Whole body	0.16-0.24		
	R. clamitans		2 pools	Whole body	0.10-0.19		
	Bufo spp.		6	Whole body	0.15-4.0		
			2	Liver	0.08-0.11		
			1 pool	Kidney	1.9		
			12	Carcass	0.19-7.3		
	Rana spp.		11	Whole body	0.10-0.36		
	- Xenopus laevis/lab				Cd^{+2} content (ug/kg wet wt.) following 100 day exposure:	Exposed to 29 ug/L cadmium chloride in a dynamic test system.	Canton and Slooff, 1982a
					Control: 0.09 ± .01; Cd-exposed: 3.77 ± .25		
			2 pools of 2	Whole body			
			4	Liver	0.26 ± .16; 4.02 ± 3.09		
			4	Kidney	0.76 ± .21; 8.22 ± 1.86		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Cadmium (cont'd)	- Bufo regularis/lab		260	Adult females 25 ± 3.3 g	Concentrations (estimated from graph, ug/g) following single dose i.m. injection of 6.2 mg Cd^{+2}/kg in thigh:		Hilmy et al., 1984c
					24 hr / 48 hr / 72 hr / 96 hr	Injection of EDTA reduced Cd level in all organs except kidneys.	
				Liver	90 / 127 / 93 / 101		
				Heart	13 / 16 / 21 / 16		
				Kidney	29 / 51 / 67 / 90		
				Lung	5 / 18 / 10 / 11		
				Spleen	1 / 3 / 1 / 1		
				Blood	17 / 4 / 2 / 2		
				Muscle	72 / 36 / 14 / 14		
	- Rana perezi/ Donana Natl. Park, Spain		10 composite samples	"A fillet with bones, skin"	Geometric mean: 0.10.		Hernandez et al., 1987
	- Rana perezi/ Donana Natl. Park, Spain	1984 1985 1986	7	Muscle	0.08 0.07 0.19		Rico et al., 1987
	- Triturus vulgaris Rana esculenta R. temporaria Palaeochen fuscus Bufo viridis/Poland	June 1975		Whole body	Location / Residues (ppm dry weight) range in brackets	Revealed some evidence of food chain accumulation.	Ozenski and Karolewski, 1979
			8		Protected zone near a zinc mill: 7.9 (1.5-16.6)		
			1		Pine forest near a zinc mill: 10.7		
			11		Control area: 0.7 (0.3-1.6)		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Cadmium (cont'd)	- Rana pipiens/lab		100 per treatment	Larvae	Cd Concentration Sediment[a] (mg/kg) / Water[a] (ug/L) / Tissue[b] (ug/g) 1.04 ± 0.6 / 1.0 ± 0.6 / - 2.28 ± 0.14 / 1.1 ± 0.8 / 0.08 11.48 ± 0.21 / 2.1 ± 4.4 / 0.34 96.8 ± 2.4 / 4.4 ± 1.8 / 3.08 1074 ± 14 / 76.5 ± 17.1 / 12.55	Exposed to Cd-enriched sediment as embryos to 4 days post-hatch. [a] mean ± SD for 3 replicate treatments. [b] composite from 3 replicate treatments.	Francis et al., 1984
Carbaryl (C) Carbofuran (see atrazine)	- Rana pipiens/lab			Skin	Half time rate of dermal penetration (mins.): 6.4.	1 mg/kg total insecticide applied to a 1 cm^2 area on back immediately behind head.	Shah et al., 1983
Cesium	- Hyla cinerea/ contaminated floodplain of Savannah River, South Carolina	June - Aug, 1972	141	Whole body	Mean ± S.E. body burden (pCi/g dry wt.): 204.2 ± 6.6.	Approximately 261 Ci radiocesium released from reactors upstream between 1960-1970.	Dapson and Kaplan, 1975
	/lab		33	Whole body	Biological half-life in unfed frogs averaged 98.1 days.	Frogs held at 20°-30°.	

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Chlordanes (OC)	- Tarichia torosa/ British Columbia		N/Days post-treatment		Techn. Chlordane / α-Chlordane / γ-Chlordane / γ-Chlordane / Cis-nonachlor / Trans-nonachlor	Lake in B.C. sprayed with technical chlordane to a level of 0.010 ppm.	Albright et al., 1980
			0/14	Whole body	8.906 / 1.289 / 1.132 / .857 / NA / 0.959	Metabolized and eliminated chlordane more effectively than cutthroat trout.	
				Liver	26.321 / 3.566 / 2.883 / 1.992 / NA / 3.210		
				Stomach	2.233 / .301 / .232 / .161 / NA / 0.238		
			8/279	Whole body	3.425 / .452 / .231 / ND / 0.310 / 1.063		
				Liver	6.234 / .590 / .285 / ND / 0.373 / 1.406		
				Stomach	.756 / .112 / .041 / ND / 0.054 / 0.222		
			6/451	Whole body	2.116 / .327 / .144 / ND / 0.153 / 0.668		
				Liver	3.106 / .392 / .172 / ND / 0.196 / 0.925		
				Stomach	.322 / .039 / .013 / ND / 0.013 / 0.063		
			8/1,036	Whole body	.176 / ND / ND / ND / 0.010 / 0.090		
				Liver	.289 / ND / ND / ND / 0.016 / 0.160		
				Stomach	Trace / ND / ND / ND / ND / Trace		
	- naturum levels/ Nonuse River drainage, North Carolina		50	Whole body	Geometric Mean of residues: Cis-chlordane / Trans-nonachlor / Cis-nonachlor / Oxy-chlordane 0.02 / 0.04 / ND* / ND	* not detected at 0.01 ppm level of sensitivity.	Hall et al., 1985
Chlorophenols	- Rana catesbeiana Conagagique Creek, Ontario	Nov. 13 1980		Tadpoles	2,6-DCP* / 2,4,6-TCP / 2,4,5,-TCP / PCP 84,000 / 10,000 / 31,000 / 19,000 2,4-DCP; 3,4-DCP; 2,3,4,6-TTCP not detectable.	*DCP - dichlorophenol; TCP - trichlorophenol; PCP - pentachlorophenol. Creek receives domestic and industrial sewage effluent.	Metcalfe et al., 1984

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Chromium (M)	- Rana esculenta River Foce, Italy	April-Sept. 1974	150	Tadpoles	Residues (ppm dry wt): 0.42-9.22.		Baudo, 1976
					Premetamorphosed Frogs: Premet. Frogs = 0.3; males = 10-22; females = 10-28 Adults: males = 0.90 (0.49); females = 0.85 (0.30) Liver: 0.93 (0.83); 2.57 (1.01); 0.50 (0.13); 0.41 (0.12); 1.34 (0.97); -; - Heart & Lungs: 0.90 (0.49); 1.41 (0.85); 2.42 (1.74); 0.46 (0.30); 2.67 (2.01); 2.67 (2.01); - Kidneys: 0.85 (0.30); 0.87 (0.30); 2.82 (2.15); 0.33 (0.22); 1.27 (1.07); 1.27 (1.07); 0.25 (0.12) Muscles Skin Gonads Eggs	Standard deviations in brackets.	
Copper (M) (see also cadmium)	- Rana esculenta/ South Moravia, Czechoslovakia	1982-1984		Whole body	Mean copper content (ppm dry wt.): Brno: 10.5 ± 1.1 (10)*; 11.8 ± 0.8 (8); - Musov: 10.7 ± 0.7 (12); 12.5 ± 0.6 (12); 14.8 ± 1.2 (10) Lednice: 6.2 ± 0.6 (13); 10.6 ± 1.1 (8); -	Brno: Suburban gardens, fields, lakes. Musov: large water reservoir. Lednice: abandoned sand pit.	Pavel and Kucera, 1986
					* Number of samples.		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Copper (cont'd)	- Rana perezi/Donana Natl. Park, Spain		10 Composite samples	"A fillet with bones, skin"	Geometric mean: 1.16.		Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1984 1985 1986	7 6 2	Muscle	0.80 0.28 2.61		Rico et al., 1987
	- Rana esculenta/ River Foce, Italy	April-Sept. 1974	150	Tadpoles	Residues (ppm dry wt): 7.63-32.40.		Baudo, 1976
					Premetamorphosed Frogs: Premet. Frogs = 50.86 (22.52); 10.22 (5.22); 4.53 (2.07); 5.01 (2.64); 9.18 (1.15); -; - Adults: males = 63.08 (24.87); 10.49 (2.97); 10.71 (7.49); 2.82 (1.91); 5.95 (2.08); 3.71 (1.30); - females = 61.44 (50.54); 7.20 (3.18); 6.69 (3.79); 2.53 (1.47); 6.14 (3.00); 10.20 (3.88); 9.54 (0.90) Liver Heart & Lungs Kidneys Muscles Skin Gonads Eggs	Standard deviations in brackets.	
	- Unspecified species/ New Lead Belt, southeastern Missouri	Sept. 1972	8 pools 1 pool	Tadpoles	Area: Strother Creek; Control Residues (ppm dry wt.): 17-44; 8	Dam across Strother Creek provides the principal tailings pond for mine and mill effluents. Samples taken from below dam.	Gale et al., 1973

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	n	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Copper (cont'd)	- Unspecified species/ New Lead Belt southeastern Missouri		1 pool	Tadpoles		Intestine included contents.	Jennett et al., 1979
			1 pool	Liver & Heart	91 ppm.		
			2 pools	Eviscerated	15-20 ppm.		
			2 pools	Intestine	234-260 ppm.		
			1 pool	Whole body	169 ppm.		
	- Patuxent Wildlife Research Centre						Hall and Holmes, 1974
	Rana catesbeiana		3 pools	Tadpoles	1.4-3.2		
	R. clamitans		2 pools	Tadpoles	0.93-1.2		
	Bufo spp.		4	Whole body	2.1-5.0		
	Rana spp.		11	Whole body	1.2-3.5		
	- Rana temporaria/ northern Finland	1971-1972	106	Liver	Mean ± SE (ppm dry wt.):		Pesonen and Kossela, 1976
				Male	154.9 ± 19.3 (entering wintering) - 903.2 ± 53.8 (emerging).		
				Female	314.0 ± 69.6 (entering wintering) to 885.1 ± 85.6 (spawning).		
	- Bufo marinus, Hyla sp., Leptodactylus sp.		21	Liver	10-1640 ppm dry wt.		Berk, 1976
	- Bufo marinus/Dominican Republic		6	Liver	367-2091 ppm dry wt.		Goldfischer et al., 1970

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	n	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Copper (cont'd)	- Plethodon cinereus, P. glutinosus, Bufo americanus, B. woodhousei, Rana pipiens/mm forest 10 km upwind from 2 zinc smelters in eastern Pennsylvania	June-Oct. 1979	23	Whole body	Mean ± SE (ppm dry weight) for all species: 3.8 ± 0.6.		Beyer et al., 1985
	- Xenopus laevis/lab		10 per dose		Concentration (mean ± SD):	1 mg Cu/kg body wt. injected i.m. daily for 5 days; frogs killed 1 day after last injection.	Tvinnereim et al., 1983
					Control / Cu		
				Liver	10.8 ± 40.2 / 41.5 ± 24.9		
				Kidney	9.15 ± 1.73 / 25.7 ± 6.0*		
					* Significantly accumulated.		
Cypermethrin (PY)	- Rana temporaria/lab		60	Brain	Mean ± SE associated with acute toxic signs: 0.08 ± 0.03 ppm.	Value is lower than that for trout, mouse, quail.	Edwards and Millburn, 1985
2,4-D (H)	- Rana temporaria/ lab			Tadpoles	Not detected (0.1 ug detection limit).	Treated at 10 ppm for 48 hours.	Cooke, 1972
DDD (DC) (see also aldrin, DDT)	- Rana catesbeiana/ Clear Lake, California	May 1958	9	Visceral fat	5 ppm.	DDD sprayed periodically 1949-1958.	Hunt and Bischoff, 1960

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
					Lake Providence		Lake Bruin			
					Residues (geometric mean)	% lipid	Residues (geometric mean)	% lipid		
DDE (DC) (see also aldrin, DDT)	- Oxbow lakes in Louisiana	1980							Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm. Biomagnification through the food chain was evident. Frogs had generally low levels due to increased mobilization and excretion, thus not useful as an indicator.	Wiethammer et al. 1984
	Rana catesbeiana R. sphenocephala R. s. clamitans		5 3 3		0.25 0.64 0.05	1.7 3.3 1.1	0.03 0.04 0.04	0.6 1.1 0.6		
	- Rana perezi/Donana Natl. Park, Spain		10 Composite samples	"A fillet with bones, skin"	Geometric mean: 0.02.					Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1985 1986	4 2	Muscle	0.02 0.19					Hino et al., 1987
	- Iowa			Viscera	Sex	Weight (g)	Residues (ppb)		Collections from agricultural areas.	Purdue et al., 1979
	Bufo americanus	1976	6		F	9.83 (9.10-10.27)	Not detected (ND)		Numbers in parentheses show the range of values.	
			17		M	3.85 (2.02-6.23)	79 (ND-176)			
	Rana pipiens	1976	6		F	1.58 (0.19-4.29)	ND		Values low due to method of analysis used.	
			3		M	0.25 (0.04-0.71)	ND			

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]					LEVELS USED/REMARKS	REFERENCE
DDE (cont'd)	- Rana boylei/Sierra Nevada Mountains, California	1970		Fat bodies	Occurrence of p,p'-DDE in fat bodies of frogs:					Concentrations probably represent locally persistent residues of 1953 and 1954 spraying.	Cory et al., 1970
					Location	Average Altitude (m)	Distance from spray area (km)	N	Residues ± S.E.	Concentrations highest in southern and central area due to wind drift.	
					Northern Yosemite	1993.9	90.0	19	1.32 ± 0.16		
					Sonora	1017.9	136.7	35	5.30 ± 0.93		
					Central: west face	1127.3	56.7	12	3.46 ± 0.25		
					west face	2953.0	128.3	48	1.19 ± 0.27		
					east of crest	2961.8	153.3	9	0.97 ± 0.19		
					southern	1988.9	90.0	19	2.87 ± 0.26		
DDMU (DC)	- Oxbow lakes in Louisiana Rana catesbeiana R. sphenocephala R. s. clamitans	1980	5 3 3		Not detected in all 3 species.					Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm.	Wiethammer et al., 1984

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
DDT (DC) (see also aldrin)	- Colorado	1960			Date Found	Residues DDT	DDD	Grazed rangeland with a few small stockponds sprayed with 9.36 kg/ha DDT on June 15, 1960.	Finley and Pillmore, 1963
	Leopard Frog				June 19	0.5			
					June 20	6			
					"	6			
					"	1	1		
					June 26	0.8	0.5		
					June 29	2	0.7		
	Garden toad				June 28	8		No DDD has ever been sprayed in Colorado thus residues must have been converted by animal tissues.	
	- Oxbow lakes in Louisiana	1980			Lake Providence		Lake Bruin	Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm.	Hoffhammer et al., 1986
					Residues (geometric mean)	% lipid	Residues (geometric mean)	% lipid	
	Rana catesbeiana		5		0.03	1.7	ND	0.4	Biomagnification through the food chain was evident.
	R. pipiens		3		ND*	3.3	ND	1.1	
	R. c. clamitans		3		ND	1.1	ND	0.6	
	- Necturus lewisi Neuse River drainage North Carolina		50	Whole body	* Not detected.			* ND - lower limit of reportable residues = 0.01 ppm	Hall et al., 1983
					Compound	Geometric Mean of Residues			
					DDT	ND*			
					DDE	0.04			
					DDD	0.04			

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]					LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Plethodon cinereus/ forest, Northern Maine		N = 10-14 pooled	Whole body	Location	DDE	DDD	DDT	TOTAL	Beaver Brook: sprayed at rate of 1.12 kg/ha in 1958, 1960, 1963.	Ramaswam, 1974
		July 1970			Beaver Br.	-	-	-	.440		
					Sterling Br.	-	-	-	.150	Sterling Brook: sprayed at rate of 1.12 kg/ha in 1960, 1963, 1964.	
					Greenlaw Mt.	-	-	-	.020		
					Weeks Br.	-	-	-	.080	Other 2 areas untreated buffer of 1.5 km.	
		June 1972			Beaver Br.	.035	tr	.035	.070		
					Sterling Br.	.090	.015	.055	.120		
					Greenlaw Mt.	tr	tr	.020	.020		
					Weeks Br.	tr	0	.025	.025		
		August 1972			Beaver Br.	.045	tr	.060	.120		
					Sterling Br.	.140	.025	.150	.270		
					Greenlaw Mt.	.010	0	.045	.055		
					Weeks Br.	tr	0	.020	.020		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Rana temporaria/lab			Tadpoles	Exposure (days)	Residues		Residues accumulated during chronic dosing did not cause mortality or behavioural abnormalities at body levels of 2-5 ppm. After acute exposure, tissue levels of 2 ppm can cause hyperactivity in tadpoles and death in small frogs.	Cooke, 1971b
						pp'-DDE	pp'-DDT		
					Control (39)	0.004	0.02		
					0.0001 ppm DDT (33)	not detected	0.02		
					(39)	0.08	0.57		
					0.001 ppm DDT (15)	not detected	2.0		
					(33)	0.25	4.3		
					(39)	0.21	3.3		
	- Rana temporaria/lab		6	Liver Muscle	16.9 ± 4.0 ppm. 1.4 ± 0.3 ppm.			Exposure to 9 subcutaneous injections of 12 mg/kg.	Cooke, 1974
	- Rana perezi/Donana Natl. Park, Spain		10 Composite Samples	"A fillet with bones, skin"	Geometric mean: 0.01.				Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1984 1985 1986	7 4 2	Muscle	0.05 0.04 0.55				Rico et al., 1987

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Rana pipiens/lab			Ventral abdominal skin	Decreasing rate of accumulation by skin vs. time.			Pithed frogs were suspended in solution of 10 ppb.	Kaiser and Dunham, 1972-73
	- Rana temporaria/lab		6 large, 6 small	Tadpoles: large - 689 ± 44 mg small - 363 ± 11 mg	Residues (mean ± S.E.): large: 2.5 ± 0.2 small: 7.5 ± 0.6			Tadpoles exposed to 0.1 ppm for 2 days in static system.	Cooke, 1979
	- Rana temporaria/lab				Stage	Nominal conc. DDT (ppm)	Residues (% of DDT originally present in test medium)	Exposure for 1 hour. No DDE found in samples.	Cooke, 1970
					I-V	0.01	16.0%		
						0.01	11.0%		
						1.0	2.1%		
						10.0	1.1%		
					IX-XVI	1.0	3.2%		
						10.0	1.7%		
	- Rana temporaria/lab		pool of 10 per group	Tadpoles younger than stage IV	Treatment time (h)	Mean content of one tadpole (ug) pp'DDT	pp'DDE	Tadpoles exposed to 0.05 ppm.	Cooke, 1971
					5	0.10	<0.001		
						0.61	<0.001		
					19	0.75	0.01		
						1.20	<0.001		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	Triturum cristatus				Tadpoles used: Mean Residue DDT / DDE Control: 0.36 / <0.002 5 h exposure: 0.23 / 0.62 19 h exposure: 0.65 / 1.38	Newts fed on R. temporaria tadpoles exposed as outlined on previous page.	Cooke, 1971 (cont'd)
	- Rana sylvatica/lab		100 per treatment	Liver, fat, head, lung, heart, gut, rest of body	Only trace residues (<0.01 ppm) found in head, lung, heart, gut and rest of body. High concentrations found in liver and fat. Mean residues in 10 frogs (treated with .001 ppm as tadpoles) sacrificed 14 days after metamorphosis: 37.4 ppm in fat, 9.7 ppm in liver.	Pre-metamorphic, stage 23 tadpoles were exposed to 0.001 or 0.003 ppm DDT for 7 days, then placed in fresh water. DDT content determined in young frogs after metamorphosis. Newly transformed frogs lost DDT more slowly while living a terrestrial life than did tadpoles.	Licht, 1976a
	- Rana temporaria/lab			Liver, fat body, gall bladder, kidney, stomach, brain, skeletal muscle, ovary, spleen, bone, spinal chord, oviduct, heart	Tissue DDT concentration was correlated with fat concentration of tissues: strong accumulation in fat body, liver, gall bladder, ovary, spleen and bones; minimal accumulation in spinal chord; no activity in kidney, oviduct, heart or skeletal muscle.	Mixed 5 uCi force fed to frogs using gelatin capsules. DDT pre-treatment increased elimination rate.	Mazzi et al., 1979

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Rana sylvatica/lab		50 per treatment, except 30 for stage 20 embryos		Temp. (°C) / Embryonic Stage: 13 / 16 / 19 / 20	Treated for 24 hrs. in 0.025 ppm.	Licht, 1985
				Embryos	9: 0.188 (.214-.153) / 0.150 (.168-.141) / 0.145 (.151-.135) / 0.956 (1.080-.832) 15: 0.150 (.159-.140) / 0.142 (.153-.133) / 0.147 (.153-.141) / 0.817 (.864-.770) 21: 0.147 (.179-.155) / 0.156 (.171-.145) / 0.160 (.173-.147) / 1.57 (.171-1.37)	95% confidence limits in parentheses. Jelly capsules around eggs restrict DDT uptake.	
				Jelly	9: 0.015 (.016-.14) / 0.017 (.019-.013) / 0.018 (.020-0.15) / - 15: 0.015 (.016-.014) / 0.019 (.017-.013) / 0.019 (0.022-.016) / - 21: 0.015 (.016-.015) / 0.013 (.016-.014) / 0.017 (.019-.015) / -		
	- Rana pipiens/lab			Skin	Half time rate of dermal penetration (mins): 783.	1 ug/kg applied to 1 cm^2 area on back immediately behind head.	Shah et al., 1981

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Rana temporaria/lab		10-30	Tadpoles at external gill stage	19.4 ppm detected in tadpoles hatched from frogs spawn exposed to DDT. No DDT detected in tadpoles hatched from spawn treated later in development. DDE not detected in any samples.				Spawn immersed for 1 hour in 0.5 ppm DDT. Detection level 0.01 ug/sample.	Cooke, 1972
				Tadpoles:	Nominal Conc. (ppm)	Treatment time (h)	DDT (ppm)	DDE (ppm)	Tadpoles exposed for 24 or 48 hours.	
			10 per group	External gills	0.5	24	75.5	ND	ND = not detected (<0.01 ug/sample).	
				Internal gills, no hind limbs	0	24	ND	ND		
						48	ND	ND		
					.005	24	2.6	ND		
						48	3.9	ND		
					.05	24	13.1	ND		
						48	19.6	ND		
					.5	24	93.2	ND		
				Hind limb paddles or hind legs	0	24	0.23	ND		
						48	ND	ND		
					.0008	24	0.17	0.01		
						48	0.29	0.03		
					.02	24	2.6	0.12		
						48	5.6	0.19		
					.5	24	36.6	0.67		
				Large hind legs	0	24	ND	ND		
					.5	24	20.2	ND		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)				Tadpoles:	Conc. (ppm)	Treatment time (h)	DDT (ppm)	DDE (ppm)	Tadpoles exposed for 24 or 48 hours.	Cooke, 1972 (cont'd)
	Bufo bufo		10 per group	External gills	0	24	ND	ND		
						48	ND	ND	ND = not detected (<0.01 ug/sample).	
					.005	24	ND	ND		
						48	ND	ND		
					.05	24	10.0	ND		
						48	ND	ND		
					.5	24	82.0	ND		
						48	104	ND		
				Internal gills, no hind limbs	0	24	ND	ND		
						48	ND	ND		
					.005	24	6.9	ND		
						48	13.5	0.40		
					.05	24	65.2	1.0		
						48	139	ND		
					.5	24	326	ND		
						48	478	ND		
				Hind limb paddle or hind legs	0	24	ND	ND		
						48	ND	ND		
					.0008	24	0.09	ND		
						48	0.54	ND		
					.02	24	3.5	0.24		
						48	7.4	0.22		
					.5	24	73.2	3.8		
						48	134	5.1		
	Triturus vulgaris		5 per group	Tadpoles with 4 legs and external gills	0	24	ND	ND		
						48	ND	ND		
					.005	24	1.5	ND		
						48	3.0	0.13		
					.5	24	86.3	2.7		
						48	196	2.2		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N		TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
						Years of DDT spray treatment	DDT metabolites (ppm)			
			1966	1967			1966	1967		
DDT (cont'd)	- Plethodon cinereus/ forests in Maine	1966-67			Whole body					Dimond et al., 1968
				22		1967		0.528	DDE comprised 40 - 60% of total residue; DDD comprised 5-10% of total.	
				17				0.640		
				6				0.160		
				7				0.648		
				5				2.057		
			7			1966	0.237			
			2				0.104		Seasonal variation in levels suggested, with higher levels in spring. Possible explanations include differing age structure of population, seasonal differences in food habits, elimination of DDT in females' eggs.	
				19		1963		0.101		
			6	22		1961	0.178	0.027		
			7	6			0.110	0.036		
				8				0.058		
				7				0.178		
				27				0.041		
			6	23		1960	0.246	0.055		
			7	29			0.190	0.036		
				22				0.037		
			16	30		1958	0.044	0.024		
			5	12			0.079	0.011		
			6			63-60-58	0.157	0.076		
			23	33		64-63-60	0.242	0.106		
				12				0.240		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
					Time after Application	Residues (mean ± SE, ppm)			
DDT (cont'd)	- Marsh at southwestern edge of Lake Erie	July 1966		Whole body				Treated by helicopter with 0.22 kg/ha technical DDT (chlorine - 36 ring-labelled).	Hooks, 1968
	Rana pipiens		3		8 hr	0.2 ± 0.0			
			6		1 day	1.3 ± 0.5			
			6		1 week	1.2 ± 0.5			
			3		1 month	1.2 ± 1.1			
			1		2 months	0.3			
			6		9 months	0.3 ± 0.1			
			1		12 months	0.2			
					Months after Application	Weight (g)	Residues (ppm)	*ND = not detected.	
	Rana catesbeiana			Liver	2	61	1.8		
					12	240	0.1		
					13	290	0.3		
				Kidney	2	61	1.8		
					12	240	0.9		
					13	290	*ND		
				Ovary	2	61	-		
					12	240	-		
					13	290	0.6		
				Testes	2	61	1.5		
					12	240	ND		
					13	290	-		
				Muscle	2	61	0.2		
					12	240	ND		
					13	290	0.3		
				Fat bodies	2	61	1.7		
					12	240	0.6		
					13	290	0.6		
				Heart, Brain, Eye, Lung, Blood, Femur, Pancreas, Spleen	No detectable residues.				
	Rana pipiens				No residues detected in tissues.				

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
DDT (cont'd)	- Rana arvalis/lab		5	Liver of tadpoles	Residues (ppm) of DDT in liver following treatment for seven days in different concentrations of DDT:	Figures given are the mean of five animals.	Licht, 1976b
					TREATMENT: 21°C — 0.001 ppm / 0.003 ppm; 15°C — 0.001 ppm / 0.003 ppm	95% confidence limits in brackets.	
					Hours after treatment: 24 — 9.73 (23.8 to -4.37) / 35.84 (61.51 to 10.17) / 18.49 (35.13 to 5.84) / 27.04 (52.27 to -0.18)	Tissue levels reached their maximum in 24 hours.	
					48 — 7.03 (20.06 to 5.30) / 5.65 (7.96 to 3.34) / 38.85 (64.05 to 13.65) / 21.20 (37.20 to 5.18)		
					72 — 14.84 (29.23 to 0.49) / 10.40 (14.80 to 6.00) / 20.96 (42.84 to -0.91) / 18.26 (28.38 to 8.14)		
					96 — 2.96 (4.86 to 1.06) / 6.20 (9.53 to 2.88) / 16.51 (34.85 to -1.81) / 16.73 (27.92 to 5.54)		
					168 — 2.67 (9.01 to 0.34) / 5.27 (11.38 to -0.84) / 9.43 (16.15 to 2.71) / 9.71 (16.28 to 3.13)		
DEHP (di-2-ethylhexyl-phthalate)	- Rana arvalis/lab		100 eggs per treatment	Tadpoles	DEHP accumulated in tadpoles at concentrations ranging from 0.28-246.80 ppm fresh weight.	Exposures for 60 days; DEHP mixed into sediment so levels of 10-800 ppm were obtained.	Larsson and Thurén, 1987

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Dieldrin (DC) (see also aldrin)	- Oxbow lakes in Louisiana	1980			Not detected in all three species.	Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall 130 cm.	Miethammer et al., 1984
	Rana catesbeiana		5				
	R. sphenocephala		3				
	R. c. clamitans		3				
	- Rana sp./Louisiana	1978		Legs	.010 ppm (mean is exceeded by standard error).		Wood et al., 1981
				Whole body	Not detected (detection limit .05 ppm).		
					Sex — Weight (g) — Residues (ppb)	Collections from agricultural areas.	Punzo et al., 1979
	- Iowa			Viscera			
	Bufo americanus	1974	6		F: 9.65 (9.10 - 10.27) — 8 (ND* -16)	* Not detected.	
			17		M: 3.89 (2.02 - 6.23) — 10 (ND - 33)	Numbers in parentheses show the range of values.	
	Rana pipiens	1974	6		F: 1.50 (0.19 - 4.29) — ND		
			3		M: 0.25 (0.04 - 0.71) — ND		
	- Rana pipiens/lab			Skin	Half time rate of dermal penetration (mins.): 1766.	1mg/kg applied to 1 cm^2 area on back immediately behind head.	Shah et al., 1983
	- Rana pipiens/lab			Ventral abdominal skin	Decreasing rate of accumulation by skin vs. time.	Pithed frogs were suspended in solution of 110 ppb.	Kaiser and Dunham, 1972-73

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
Dieldrin (cont'd)	- lab		10 per group	Tadpoles:	Nominal Conc. (ppm)	Treatment Time (h)	Mean Dieldrin (ppm)	Tadpoles exposed for 24 or 48 hours.	Cooke, 1972
	Rana temporaria			Stages IV-XIV	0	24	ND		
						48	ND		
					.0008	24	0.17		
						48	0.31		
					.02	24	1.8		
						48	6.1		
					.5	24	31.3		
						48	62.9		
	Bufo bufo			Stages VI-XVI	0	24	ND		
						48	ND		
					.02	24	3.6		
						48	5.6		
					.5	24	88.5		
						48	138		
	- nocturna levis/ Neuse River drainage, North Carolina		50	Whole body	Geometric Mean of residues: 0.02.				Hall et al., 1985
Endrin (OC)	- Oxbow lakes in Louisiana	1980			Not detected in all three species.			Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm.	Riethmuller et al., 1986
	Rana catesbeiana		5						
	R. sphenocephala		3						
	R. c. clamitans		3						

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
Dursban 4e Hew (anthropoofing chemical)	- Rana temporaria/lab			Tadpoles:	Treatment	Dursban residue		Tadpoles placed in solutions; residues determined after 35-36 days of exposure.	Osborn and French, 1981
			4 pools	Stage 31	Control	not detected			
			1 pool	Stage 26-29	1.0 mg/L	24.1			
			4 pools	Stage 31	0.1 mg/L	7.71 ± 1.60			
			4 pools	Stage 31	0.01 mg/L	0.53 ± 0.11			
			1 pool = 5 indivs.						
Fenitrothion (OP)	- Bufo americanus/ Quebec	1984		Larvae	Days Post-Spray	Stage	Residue	Aerially treated with 210 g a.i./ha. No metabolites detected.	Mesarbachi et al., 1987
			2		Pre	26-32	<.005		
			2		1.5	27-34	.045-.123		
			2		3.0	27-35	<.005-.021		
			1		19.0	33-44	<.005		
	- Rana clamitans/lab		10-15 per pool	Larvae	5.30 after 1 hour, pooled at 11.50 at day 1.			1 ppm test medium formulated, but chemical concentration was .523 ppm, indicating poor mixing.	Lyons et al., 1976
	Rana sylvatica/Lorne Forest, Ontario	June 1975	5-16 per pool	Tadpoles	0.61 after 1 hour (185 times the concentration in water at same time); 0.04 at day 2 (18 times water sample concentration).			Aerially sprayed at rate of 280 g a.i./ha.	

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Fenthion (OP)	- Pseudacris triseriata/ hay meadows near Laramie, Wyoming	June	175	Whole body	No residues detected in frogs collected 1-3 days after treatment. Lower limit of detection was 0.01 ppm.	Sprayed at rate of 52 g a.i./ha as either oil-based formulation (2.3 L ha. 2 diesel oil/ha) or water-based (emulsifier and 2.3 L water/ha). Concluded frogs not a likely source of secondary poisoning for predatory vertebrates.	Powell et al., 1982
	- Rana catesbeiana/lab			Tadpoles	Significant bioconcentration: average magnification 62 times. Maximum level found in pooled samples was 320 ppm. No fenoxon (metabolite) detected in tissues at sensitivity limit of 0.5 ppm.	Exposed to .01-1 ppm for 96 hr in continuous flow apparatus.	Hall and Kolbe, 1980
Fenvalerate (PY)	- Bufo fowleri, Rana utricularia, R. clamitans, Hyla crucifer/in and around cotton fields near Garland, Arkansas	July 23-26, 1979	1 or 2 (pooled)	Carcass with skin and g.i. tract removed	0.02 (ne2) found in B. fowleri. No detectable residues (0.01 ppm detection limit) found in other species.	Aerial application had occurred 5 days prior to collection at rate of 0.112 kg ai/ha. Applications had also been made in previous years.	Barnett et al., 1983

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
g-HCH (OC)	- Rana perezi/Donana Natl. Park, Spain		10 composite samples	"A fillet with bones, skin"	Geometric mean: 0.01.				Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1984	2	Muscle	0.01				Rico et al., 1987
Heptachlor (OC)	- Rana pipiens/lab			Ventral abdominal skin	Decreasing rate of accumulation by skin vs. time.			Pithed frogs were suspended in solution of 6 ppb.	Kaiser and Chatham, 1972-73
Heptachlor Epoxide (OC)	- Location not specified	1962			Residues (ppm) from animals found dead one month after spray:			Area sprayed with 2.24 kg/ha.	Hewitt et al., 1962
	Bullfrog		2		13.5				
	Leopard Frog		2		13.8				
	Green Frog		6		1.5				
	Toads		2		19.4				
	- Iowa			Viscera	Sex	Weight (range in brackets) (g)	Residues (ppm)	Collections from agricultural areas. Values low due to method of analysis used.	Punzo et al., 1979
	Bufo americanus	1974	4		F	9.45 (9.10-10.27)	Not detected (ND)	DDT was used as a general purpose insecticide in this region in past years and aldrin and heptachlor were used for control of soil-insect pests.	
			17		M	3.85 (2.02-6.23)	ND		
	Rana pipiens	1974	4		F	1.50 (0.19-4.29)	ND		
			3		M	0.25 (0.06-0.71)	ND		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Heptachlor epoxide (cont'd)	- Taricha torosa/ British Columbia	1980	N/Days post treatment		Residues	Lake in B.C. sprayed with technical chlordane to a level of 0.010 ppm.	Albright et al., 1980
			4/14	Whole body	0.343	No application rate given.	
				Liver	0.962		
				Stomach	0.072		
			8/279	Whole body	ND (not detectable)		
				Liver	ND		
				Stomach	ND		
			6/651	Whole body	ND		
				Liver	ND		
				Stomach	ND		
			8/1,036	Whole Body	ND		
				Liver	ND		
				Stomach	ND		
Iron (N)	- Rana esculenta/South Moravia, Czechoslovakia	1982-1984		Whole body	Mean iron content (ppm dry wt.):	Brno: Suburban gardens, fields, lakes.	Pavel and Kucera 1986
					Locality / Years: 1982 / 1983 / 1984	Musov: large water reservoir.	
					Brno: 125.3 ± 7.7 (10)* / 119.4 ± 13.6 (8) / -	Lednice: abandoned sand pit.	
					Musov: 132.8 ± 13.2 (12) / 168.0 ± 25.1 (11) / 197.8 ± 83.0 (10)		
					Lednice: 74.7 ± 6.2 (13) / 136.6 ± 12.6 (7) / -		
					* number of samples.		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Lead (N)	- Triturus vulgaris Rana esculenta R. temporaria Pelobates fuscus Bufo viridis/Poland	June 1975	0 1 11	Whole body	Location / Residues (range in brackets): Protected zone near a zinc mill: 124.8 (46.7-282.9) Pine forest near a zinc mill: 461.8 Control area: 0.0 (0.0-0.0)	Some evidence of food chain accumulation.	Dmowski and Kozlowski, 1979
	- Xenopus laevis/lab		3 groups of 6 toads each	Bone Skin Muscle Kidney Liver	Group A / Group B / Group C: 23.97 ± 6.09 / 7.95 ± 0.97 / 5.00 ± 0.20 2.98 ± 0.30 / 1.58 ± 0.25 / 0.81 ± 0.17 1.14 ± 0.08 / 1.01 ± 0.20 / 5.9 ± 0.04 15.07 ± 1.82 / 10.43 ± 3.21 / 3.48 ± 0.58 7.80 ± 1.78 / 5.96 ± 1.58 / 1.13 ± 0.08	Group A - intake of 11.99 ± 1.11 ug/day/toad avail. Pb for 4 wks; 13.63 ± 2.57 avail. Pb for 4 wks. Group B - intake of 12.89 ± 1.19 ug/day/toad for 4 wks. Group C - intake of 0.76 ± 0.10 ug/day/toad for 4 wks; 1.13 ± 0.14 for 4 wks.	Ireland, 1977
	- Highway drainage ditches in Maryland and Virginia	July-Dec. 1982			Geometric mean (ppm dry wt.) / 95% Confidence Intervals	Lead concentrations positively correlated with average daily traffic volumes.	Birdsall et al., 1984
	Rana catesbeiana			Tadpoles	14 / 2.5-72	Concentrations in sediment were usually higher (4-5 times) than those in tadpoles.	
	R. pipiens			Tadpoles	14 / 7.3-25		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Lead (cont'd)	- Rana perezi/Donana Natl. Park, Spain		10 Composite samples	"A fillet with bones, skin"	Geometric mean: 0.83.		Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1986 1984 1985	2 7 4	Muscle	1.19 0.48 1.82		Rico et al., 1987
	- Plethodon cinereus, P. glutinosus, Bufo americanus, R. sylvatica/ oak forest 10 km upwind from 2 zinc smelters in eastern Pennsylvania	June-Oct., 1979	23	Whole body	Mean ± SE (ppm dry weight) for all species: 12 ± 1.3.		Beyer et al., 1985
	- Unspecified species/ New Lead Belt, southeastern Missouri	Sept. 1972	8 pools 1 pool	Tadpoles	Area: Strother Creek — Residues (ppm dry wt.): 36-1990 Area: Control — Residues (ppm dry wt.): 28	Dam across Strother Creek provides the principal tailings pond for mine and mill effluents. Samples taken from below dam.	Gale et al., 1973
	- Unspecified species/ New Lead Belt, southeastern Missouri		1 pool 2 pools 2 pools 1 pool	Tadpoles: Liver & Heart Eviscerated Intestine Whole Body	22 ppm. 93-213 ppm. 4419-7329 ppm. 4139 ppm.	Intestine included contents.	Jennett et al., 1979

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Lead (cont'd)	- Patuxent Wildlife Research Centre						Hall and Mulhern, 1984
	Rana catesbeiana		5 pools	Tadpoles	2.5-3.2		
	R. clamitans		2 pools	Tadpoles	1.4-1.5		
	Rana spp.		11	Whole body	0.88-3.2		
	Bufo spp.		3 pools	Carcass	21		
			1 pool	Kidney	4.9		
			2	Liver	2.1-2.6		
	- Watershed in east-central Illinois			Whole body	Mean residues (ppm dry wt):	Collected from areas 5 and 50 m from interstate highway.	Rolfe et al., 1977
	Acris crepitans		14		2.7		
	Bufo americana		31		3.0-3.5		
	- Rana pipiens			Liver	R. pipiens: 1.0 - 6.2	R. sylvatica: 14.8 - 31.3	Schroeder and Tipton, 1968
	R. sylvatica/ sparsely inhabited areas of Vermont			Kidney	1.3 - 8.2	10.2	
	- Rana catesbeiana/ lead mining district, southeastern Missouri	1981-82	14-15	Whole body	Residues (geometric mean, range in parentheses): Site A: 0.97 (.11-4.10); Site B: 13.9* (3.50-31.0); Site C: 14.0* (2.90-41.0). * significantly higher than Site A.	Site A - upstream from lead belt, Sites B and C - downstream from tailings ponds.	Niethammer et al., 1985
	- Xenopus laevis/lab		12	Dorsal skin	Residues (ppm dry wt) in frogs pretreated for 2 months on either dark or light background: Dark- adapted: 187.9 ± 13; Light-adapted: 114.8 ± 17.6	Subjected to 50 ppm lead nitrate for 21 days, medium changed daily. Pb concentrated within melanophores.	Ireland et al., 1979

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Lead (cont'd)	- Rana esculenta/ River Toce, Italy	April-Sept. 1974	150	Tadpoles	Residues (ppm dry wt): 0.58-1.47.		Baudo, 1976
					Nonmetamorphosed Frogs: Nonmet. Frogs = 43; males = 10-22; females = 10-28	Adults: males; females	Standard deviations in brackets.
				Liver	0.94 (0.65)	1.16 (0.48)	7.56 (10.57)
				Heart & Lungs	1.42 (0.41)	0.40 (0.33)	2.73 (2.83)
				Kidneys	0.94 (0.68)	2.34 (2.21)	4.64 (3.37)
				Muscles	1.70 (1.40)	0.68 (0.52)	3.69 (5.49)
				Skin	0.96 (0.47)	3.66 (4.00)	3.18 (2.39)
				Gonads	-	0.56 (0.29)	4.16 (4.96)
				Eggs	-	-	0.89 (0.33)
Lindane (OC)	- Rana pipiens/lab		6		Frogs were exposed to vaporized lindane for 18, 23 or 38 days:		Frogs had higher residues than chickens, rats, mice, fish or plants. Failure to eat may have lowered their metabolism and thus lowered their excretion of lindane. Whitacre and Ware, 1967
					Days following exposure: 18; 23; 38		
					Brain residues (ppm): 3.7; 6.3; 55.5		
					Liver residues (ppm): 6.5; 26.0; 53.0		
					Fat residues (ppm): 10.0; 7.5; None		
	- Rana pipiens/lab			Ventral abdominal skin	Decreasing rate of accumulation by skin vs. time.	Pithed frogs were suspended in solution of 10 ppm.	Kaiser and Dunham, 1972-73

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Magnesium (Mg)	- Patuxent Wildlife Research Centre			Tadpoles		Collected from uncontaminated areas.	Hall and Mulhern, 1984
	Rana catesbeiana		5 pools		58-160		
	R. clamitans		2 pools		14-29		
	- Rana temporaria/ Northern Finland		106	Liver Male Female	Mean ± SE (ppm dry wt): 298.5 ± 30.7 (wintering) -604.2 ± 49.0 (feeding) 335.8 ± 69.1 (pre-emerging) -749.0 ± 61.8 (feeding).	Winterings: Nov. - March Feedings: June - August Pre-emerging: April.	Paasonen and Koskela, 1978
Manganese (Mn)	- Rana esculenta/South Moravia, Czechoslovakia	1982-1984		Whole body	Mean Manganese content (ppm dry wt):	Brno: Suburban gardens, fields, lakes. Musov: large water reservoir. Lednice: abandoned sand pit.	Pavel and Kucera, 1986
					Locality: Years: 1982; 1983; 1984		
					Brno: 24.1 ± 2.8 (10)*; 25.8 ± 5.9 (8); -		
					Musov: 24.7 ± 2.0 (12); 35.9 ± 4.2 (12); 51.9 ± 6.3 (10)		
					Lednice: 15.4 ± 1.4 (13); 23.4 ± 4.2 (8); -		
					* Number of samples.		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
Manganese (M) (cont'd)	- Rana esculenta/ River Toce, Italy	April-Sept. 1974	150	Tadpoles	Residues (ppm dry wt): 23-958.				Bauso, 1976
					Neometamorphosed Frogs	Adults		Standard deviations in brackets.	
						males	females		
			Neomet. Frogs = 43; males = 10-22; females= 10-28	Liver	15.4 (4.2)	28.2 (4.5)	22.1 (7.8)		
				Heart & Lungs	8.0 (3.3)	8.4 (2.8)	6.7 (3.7)		
				Kidneys	8.4 (6.8)	21.3 (14.8)	5.1 (3.9)		
				Muscles	5.3 (1.8)	6.5 (2.2)	5.2 (2.2)		
				Skin	14.6 (8.8)	67.9 (19.8)	30.3 (8.2)		
				Gonads	-	5.8 (4.1)	53.8 (23.1)		
				Eggs	-	-	8.1 (1.5)		
	- Unspecified species/ New Lead Belt, southeastern Missouri	Sept. 1972		Tadpoles	Area	Residues (ppm dry wt.)		Dam across Strother Creek provides the principal tailings pond for mine and mill effluents. Samples taken from below dam.	Gale et al., 1973
			8 pools		Strother Creek	500-5650			
			1 pool		Control	710			
	- Patuxent Wildlife Research Centre			Tadpoles					Hall and Mulhern, 1984
	Rana catesbeiana		5 pools		14-42				
	R. clamitans		2 pools		1.1				

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
Mercury (M)	- Lake St. Clair, Ontario	1970			Sex	Carcass	Liver		Dustman et al., 1972
	Rana pipiens		1		F	0.18	0.61		
			1		M	<0.10	1.1		
			1		M	<0.10	0.51		
	R. catesbeiana		1		F	<0.10	0.28		
	- Rana tigrina/lab			Liver, Kidney	^{203}Hg accumulated in both high and low MW fractions of kidney 2,4,7 days after administration. In liver, ^{203}Hg appeared in high MW fractions at 2 days, in both high and low at 4,7 days.			Single i.p. dose of 90 uCi/100 g body weight given.	Mehra et al., 1980
	- Paljava, Yugoslavia	1975			Hg	MeHg (ppm fresh weight)		Samples collected from uncontaminated area.	Byrne et al., 1975
	Bufo bufo male (m)			Liver	1.51	0.37			
	B. bufo female (f)				0.94	0.27			
	Rana dalmatina (m)				0.67	0.83			
	Bombina variegata (m)				2.07	0.35			
	R. arvalis (f)				1.96	1.1			
	B. bufo (m)			Kidney	1.28	-			
	B. bufo (f)				0.60	0.12			
	R. dalmatina (m)				1.01	-			
	B. variegata (m)				0.93	0.08			
	R. arvalis (f)				1.63	0.40			

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	n	TISSUE	RESIDUE LEVEL[b]		LEVELS USED/REMARKS	REFERENCE
Mercury (cont'd)	Paljevo, Yugoslavia (cont'd)	1975			Hg	MeHg (ppm fresh weight)		Byrne et al., 1975 (cont'd)
	B. bufo (m) B. bufo (f) R. dalmatina (m) R. vertepoda (m) R. arvalis (f)			Lungs	0.17 0.16 0.25 0.11 0.67	0.02 0.13 - 0.04 0.25		
	B. bufo (m) B. bufo (f) R. dalmatina (m) R. vertepoda (m) R. arvalis (f)			Muscle	0.17 0.16 0.50 0.17 0.40	0.15 0.19 0.33 0.26 0.45		
	Idrija, Yugoslavia	1975						
	Bufo bufo male (m) B. bufo female (f) B. bufo (f) Rana temporaria (f)			Liver	23.4 22.2 25.5 25.9	6.1 3.8 5.5 5.1	Samples collected from contaminated areas outside a mercury mine. Liver is prime accumulating area.	
	B. bufo (m) B. bufo (f) B. bufo (f) R. temporaria (f)			Kidney	24.8 21.0 25.3 16.2	2.1 1.8 2.6 3.0		
	B. bufo (m) B. bufo (f) B. bufo (f) R. temporaria (f)			Lungs	1.60 1.11 1.70 1.14	1.15 0.77 1.05 1.8		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	n	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
Mercury (cont'd)	Idrija, Yugoslavia (cont'd)	1975			Hg	MeHg (ppm fresh weight)				Byrne et al., 1975 (cont'd)
	B. bufo (m) B. bufo (f) B. bufo (f) R. temporaria (f)			Muscle	2.85 1.39 2.76 5.66	2.76 1.55 2.66 5.66				
	B. bufo (m) B. bufo (f) B. bufo (f) R. temporaria (f)			Eggs	2.15 2.30 1.2.	0.60 1.40 1.5				
	- Finland Rana temporaria				Mercury content (ppm fresh weight) given as mean ± S.E.:				Seems that frogs, though representing a high trophic level, are not an important pathway for transfer of mercury along food chains. * Number of samples.	Tejhvuo et al., 1984
					Tavisa	Parvoo	Helsinki	Hammarbyvo		
				Liver	.06 ± .01 (9)*	.13 ± .04 (12)	.19 ± .04 (6)	.05 (1)		
				Kidney	.00 ± .03 (7)	.05 ± .01 (10)	.03 (1)	-		
				Lung	.07 ± .01 (9)	.07 ± .02 (12)	.10 ± .03 (6)	.04 (1)		
				Muscle	.03 ± .00 (9)	.04 ± .02 (12)	.07 ± .03 (6)	-		
				Egg	.04 ± .01 (3)	-	-	-		
				Whole body	-	-	-	.01 (1) .01 (10)		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
Mercury (cont'd)	Finland (cont'd)				Mercury content (ppm fresh weight) given as mean ±S.E.(n)				[a] Number of samples.	Tarkiwa et al., 1984 (cont'd)
					Tavina	Porvoo	Helsinki	Hammarbyra		
	Bufo bufo			Liver	.11 ± .02 (4)[a]	-	-	-		
				Kidney	.08 ± .01 (3)	-	-	-		
				Lung	.04 ± .01 (4)	-	-	-		
				Muscle	.04 ± .01 (4)					
	- Rana sp./Aiken, South Carolina	Aug. 1973 to Jan. 1974		Tadpoles					Low level mercuric ion concentrations of 1.0 and 5.0 ug/l maintained in streams. Tadpoles eat algae which concentrates Hg.	Cox et al., 1975
			N of analyses		Total Mercury	Methyl Hg	% Methyl Hg			
			3		1.08	0.01	0.48			
			3		4.36	0.03	0.69			
			1		6.41	0.00	0.00			
	- Rana perezi/Donana Natl. Park, Spain		10 Composite samples	"A fillet with bones, skin"	Geometric mean: 0.20.					Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1984 1985 1986	7 6 3	Muscle	0.08 0.09 0.13					Rice et al., 1987

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]			LEVELS USED/REMARKS	REFERENCE
Mercury (cont'd)	- Patuxent Wildlife Research Centre								Hall and Mulhern, 1984
	Rana catesbeiana		5 pools	Tadpoles	0.05-0.10				
	R. clamitans		2 pools	Tadpoles	0.06-0.10				
	Rana spp.		11	Whole body	<0.01-0.14				
	Bufo spp.		4	Whole body	0.04-0.14				
	- Savannah River floodplains and tributaries, Georgia	1972-73		Adults	Skeletal Muscle		Liver	Savannah River contaminated sites contaminated fish at levels thought to be hazardous to human health.	Clemmie, 1974
	Rana pipiens				0.07-0.10		0.25		
	R. catesbeiana				0.09-0.24		0.09-0.44		
	Hyla cinerea				0.03		-		
	Bufo terrestris				<0.0.14		-		
Methoxychlor (DC)	- Bufo americanus/Lab			Whole body	Days of exposure	Dosage Group (ppm)	Residues geometric mean (95% confidence limits)	No changes in organ weights, feeding, behaviour or survival.	Hall and Swineford, 1979
			4		1	0.024 in food	0.011 (0.003-0.045)	Residues not correlated with exposures.	
			4		4	"	0.008 (0.002-0.024)		
			5		1	0.125 in food	0.033 (0.012-0.088)		
			5		1	0.069 ± 0.036 in water	0.145 (0.061-0.321)		
			5		4	"	0.244 (0.124-0.482)		
			4		14	"	0.124 (0.060-0.327)		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Monocrotophos (C)	- Rana clamitans/ pond in mixed boreal forest, northeastern New Brunswick	June 1985		Tadpoles Whole body	None of the samples of caged or wild tadpoles contained detectable levels (>10 ppb, fresh wt.).	Two aerial applications of 70 g a.i./ha on June 2, 8.	Sundaram et al., 1986
Mirex (OC)	- Acrie crytiua/1.2 ha fish pond, Mississippi	Treatment on Oct. 23, 1970	6 or more individuals in each pooled sample	Whole body	Mirex residues (ppm) at given intervals after treatment:	Application rate (aerial) 1.4 kg/ha.	Collins et al., 1972
					Pre-spray: ND; 3 days: 0.03; 30 days: 2.26; 6 months: 2.68; 12 months: 0.02; 16 months: 0.05	Pond was part of a larger drainage system.	
	- Rana crytiua Winston Co., Mississippi	Treatment in May and June, 1972	1	Tadpole	Residue in sample collected less than 6 months after application - 0.12.	Application rate (aerial) 1.4 kg/ha.	Hooper and de la Cruz, 1973
	- Oxbow lakes in Louisiana Rana catesbeiana R. sphenocephala R. c. clamitans	1980	5 5 5		Not detected in all three species.	Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm.	Matthumner et al., 1980

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION		DATE	N	TISSUE	RESIDUE LEVEL[b]						LEVELS USED/REMARKS	REFERENCE
Mirex (cont'd)	- Southwest Georgia					Application Rate (g/ha)	Pre-treatment	Residues (ppm) Post Treatment: 7 days	1 mo	3 mos	6 mos	12 mos	Figures in brackets represent number of samples in the pool.	Hajek et al., 1975
	Hyla versicolor	Turner Co.	1971			2.10	0	0	0.94 (5)				D= discarded cross-contaminated samples. ND= no residues detected at 0.01 ppm level.	
						4.20	0		0.18 (1)	0.39 (1)				
		North Co.	1971-72			1.12	ND (3)		0.24 (5)		0.02 (2)		Maximum levels detected 1-3 months after spray; gradually declined to lower levels over 1 year.	
						4.20					ND (1)			
	Gastrophryne carolinensis	Tift Co.	1971			1.12	0			0.47 (2)				
						1.68	0			3.46 (3)				
		Turner Co.	1971			2.10	0	0	2.02 (9)	0.41 (16)				
						4.20			0.33 (1)	1.06 (3)				
		North Co.	1971-72			1.12	ND (16)					0.17 (6)		
						4.20	0.12 (5)					0.18 (1)		
	Pseudacris nigrita	North Co.	1971-72			1.12					0.10 (1)			
	Acris crytiua	North Co.	1971-72			1.12			9.27 (2)		0.14 (1)			
						4.20			1.69 (9)					
	R. sphenocephala	North Co.	1971-72			1.12						ND (5)		
						4.20						0.08(2)		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]							LEVELS USED/REMARKS	REFERENCE
					Application Rate (g/ha	Pre-treatment	Residues (ppm) 7 days	1 mo	3 mos	6 mos	12 mos		
Mirex (cont'd)	Southwest Georgia											D = discarded cross-contaminated samples. ND = not detected at 0.01 ppm level.	Wojcik et al., 1975 (cont'd)
	Apis mellifera Taft Co.	1971			1.68				0.08 (1)				
	Turner Co.	1971			2.10 4.20		D	 0.24 (1)	1.08 (1)				
	North Co.	1971-72			1.12 4.20	 ND (1)		 0.34 (1)	0.56 (1)				
	B. columbarum Taft Co.	1971			1.12 1.68			0.05 (1)	 0.15 (4)				
	Turner Co.	1971			2.10	D		0.43 (1)					
	North Co.	1971-72			1.12 4.20	ND (4)		0.08 (12) 0.25			0.03 (4) 0.09 (1)		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]							LEVELS USED/REMARKS	REFERENCE
Mirex (cont'd)	- 8,000 ha near Jacksonville, Florida	1972-1974		Whole body	Residues (ppm fresh weight) at given interval post-treatment:							Application of 0.15 bait by fixed-wing aircraft at an altitude of 45.7 m at rate of 1.12 kg/ha of mirex.	Wheeler et al., 1977
					pre-treatment	1 mo.	3 mo.	6 mo.	9 mo.	12 mo.	24 mo.		
	Bufo terrestris				(6) ND (1) 0.03	-	-	(1) ND	(1)0.09				
	B. quercicus				(7) ND (6) 0.42	(6) 0.79	(11) 0.11	(1) 1.81 (1) 0.20	(1) 0.15	(3) 0.04	(1) ND		
	Rana heckscheri				(7) ND	(3)0.28 (12) 0.02	(5) ND	(2) ND	(1) ND (12) ND (4)0.26	(6) ND			
	R. pipiens				(1) ND	(1) ND (1)0.04	(1) ND (1)0.13		(3) 0.01				
	R. sphenocephala				(1) ND	(6) 0.04	(3) ND (1)0.02		(7) ND		(6) ND		
	R. areolata					(6) 0.14							
	Rana sp. tadpoles					(7) ND	(12) ND	(10) ND					
	Gastrophryne carolinensis				(38) ND	(3) 0.52	(3) 0.09	(3) 0.17	(1) 0.01	(1) 0.04	(10) ND		
	Scaphiopus holbrooki				(1) ND	(1) ND		(1)0.02 (1)0.02		(3) 0.05	(1) ND		
	Mole rumors					(1) ND	(1) ND		(1) ND				
	N. fumarolis					(2) 0.08		(1) ND					
Nematicides (EC) (see chlordane)													

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Organochlorine compounds (DDT, DDD, DDE, g-Hexachlorocyclohexane)	- Rana ridibunda/ clean pond and rice field discharge canal, North Caucasus, U.S.S.R.		280	Muscle	Residues (ppm): Pond: immature .00196, .00193; mature .00174, .00177. Rice Field: immature .15970, .16010; mature .15610, .15700. Female: Pond: immature .00195, .00191; mature .00168, .00171. Rice Field: immature .15930, .15920; mature .15530, .15960.	Residues in water - 0.00006 in pond, 0.00330 in rice field.	Zhukova, 1987
Oxychlordane (OC)	- Taricha torosa/ British Columbia	1980	N/days post treatment		4 residues	Lake in B.C. sprayed with technical chlordane to a level of 0.010 ppm. No application rate given.	Albright et al., 1980
			4/14	Whole body	ND - not detectable		
				Liver	ND		
				Stomach	ND		
			8/279	Whole body	0.115		
				Liver	0.134		
				Stomach	0.027		
			6/451	Whole body	0.060		
				Liver	0.097		
				Stomach	0.010		
			8/1,036	Whole body	0.011		
				Liver	0.011		
				Stomach	ND		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Parathion (EP)	- Acris crepitans/lab		1 pool of 12 per treatment	Whole body	Group: Control; 0.1 ppm; 1.0 ppm; 10 ppm. Parathion Concn.: <0.05; <0.05; 0.08; 4.6	Frogs held individually for 96 hours in static systems.	Fleming et al., 1982
	- Rana pipiens/lab			Skin	Half time rate of dermal penetration (elim.): 19%.	1 mg/kg applied to 1 cm^2 area on back immediately behind head.	Shah et al., 1981
	- Rana catesbeiana/lab			Tadpoles	Significant bioconcentration; average amplification - 64 times. Maximum level found in pooled samples was 96 ppm. No paraoxon (metabolite) detected in tissues at sensitivity limit of 0.5 ppm.	Exposed to .01-.1 ppm for 96 hr in continuous flow apparatus.	Hall and Kolbe, 1980
PCBs (polychlorinated biphenyls)	- Oxbow lakes in Louisiana: Rana catesbeiana; R. sphenocephala; R. G. clamitans	1980	5 3 3		Not detected in all three species.	Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 130 cm.	Hathaway et al., 1986
	- Rana sp./Louisiana	1978		Legs; Whole body	.017 ppm (mean exceeded by standard error). Not detected (0.10 ppm detection limit).	Site was an area of low insecticide use.	Dowd et al., 1985
	- Rana perezi/Donana Natl. Park, Spain	10 Composite samples		"A fillet with bones, skin".	Geometric means 0.06.		Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	7 4 2		Muscle	0.05 0.49 1.08		Rico et al., 1987

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
PCBs (cont'd)	- Nocturva lewisi/ House River Drainage, North Carolina		50	Whole body	Geometric Mean of residues: 0.39.	Analysed for PCB 1254.	Hall et al., 1985
Permethrin (PY)	- Rana pipiens/lab			Skin	Half time rate of dermal penetration (area): 420.	1 mg/kg applied to 1 cm^2 area on back immediately behind head.	Shah et al., 1983
Phenols	- Rana temporaria/lab				Distribution of phenols (g/g) in various tissues at given times after injection via lymph sac at 5 mg/kg (PH = phenol, NP = 3-nitrophenol, DP = 3,5-dinitrophenol, AP = 4-aminophenol):	Average weight of frogs used was 28 ± 8 g. Temperature 20° C.	Nagel and Urich, 1981

	0.5 h				2 h				6 h				15 h			
	PH	NP	DP	AP	PH	NP	DP	AP	PH	NP	DP	AP	PH	NP	DP	AP
Blood	11.6	8.9	16.5	11.3	5.8	6.4	13.3	18.5	2.1	5.6	15.9	7.1	1.3	0.8	2.7	4.4
Gut	6.9	2.3	7.2	7.1	3.2	3.5	7.2	7.0	1.5	4.0	7.2	8.7	1.2	1.6	7.8	3.5
Brain	3.7	3.9	9.4	5.9	3.0	1.5	3.5	3.5	1.1	1.4	1.9	3.5	0.5	0.1	0.5	1.2
Skin	6.3	16.3	8.8	5.5	3.1	3.9	5.4	6.2	1.4	1.9	3.8	3.8	1.5	0.6	0.9	1.9
Heart	5.2	6.3	9.6	6.7	3.6	3.8	5.9	3.9	1.2	2.6	6.6	3.2	0.8	0.3	1.1	1.5
Liver	6.3	5.3	8.3	12.9	2.7	6.6	8.9	6.7	1.4	3.9	7.5	5.6	0.9	1.2	2.2	2.0
Lung	5.4	7.1	10.7	10.2	2.6	4.2	7.9	5.6	1.7	3.3	5.7	6.0	0.9	0.7	1.9	3.8
Stomach	4.8	2.7	8.3	6.4	2.3	3.3	7.1	6.8	1.3	2.8	5.3	4.3	0.5	0.6	1.0	1.5
Spleen	8.0	2.6	9.7	6.9	4.7	4.6	10.3	5.2	1.5	3.4	7.2	6.9	1.1	0.6	2.1	2.5
Kidney	8.9	16.0	17.9	10.7	12.4	15.1	13.6	6.3	7.8	18.4	9.2	5.8	2.6	8.1	2.9	3.1
Pancreas	6.2	4.3	10.5	7.2	5.0	6.9	10.4	6.5	1.5	2.5	7.5	6.5	1.1	0.7	1.7	2.0
Remainder	2.9	6.2	6.6	6.0	1.8	2.9	6.3	3.5	1.1	2.0	3.1	3.6	0.6	0.3	0.8	1.1
Gall bladder	6.1	6.8	16.3	15.5	2.6	11.9	113	29.4	7.3	73.8	287	69.8	10.0	48.4	126	14.0
Contents of guts	1.8	8.3	3.8	3.6	3.6	8.9	8.8	9.7	1.4	12.7	7.4	20.9	7.7	10.4	84.3	30.0

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Selenium (N)	- Xenopus laevis/lab			Embryos at gastrula stage	Uptake rapid and linear until 36 h of exposure. Linear decrease in Se over first 24 h of clearing; rate of depuration similar to rate of uptake.	Gastrulae placed in 5 ppm ^{75}Se for up to 48 h. After 24 h, some transferred to untreated water	Browne and Dumont, 1979
				Tadpoles	80% decrease in Se content within 30 min; Se level remained stable for rest 3 days.	Embryos exposed for 48 h; hatched tadpoles transferred to untreated water.	
Strontium (N)	- Taricha granulosa/lab			Whole body	Biological half-lives - 136 and 131 days for chronic and acute treatment, respectively. Significant concentrations found in blood, liver, skin, muscle and bone; after 25 days, most ^{85}Sr in bone.	Retention of ^{85}Sr following chronic and acute (single i.p. injection) administration at 10°C.	Willis and Valett, 1971
	Rana pipiens			Whole body	Biological half-life of 222 days.	Retention of ^{85}Sr following i.p. injection at 10°C.	
	- Taricha granulosa/lab				Slower loss component had biological half-lives of 130 and 88 days at 10° and 20°C, respectively.	Retention of ^{85}Sr following chronic or acute treatment at 10° or 20°C.	Willis et al., 1976
	Rana pipiens				Half-lives longer than those found for newts at the 2 temperatures.	Retention following i.p. injection.	
	- Rana temporaria/ lab		100 per tanks	Tadpoles 5-7 weeks old	100 tadpoles absorbed approximately 18% (0.02 ug) of the strontium-90 and about 72% of the yttrium-90 in 175 hr.	Treated with 0.125 ug strontium-90 in 500 ml water, giving 2.5×10^5 disintegrations/min strontium-90 and yttrium-90.	Lucas and Pickering, 1958

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]						LEVELS USED/REMARKS	REFERENCE
2,3,7,8-TCDD (2,3,7,8-tetrachlorodibenzo-p-dioxin)	- Toad Sp./Italy	1980	1	Whole body	0.0002 ppm found in toad 2 yrs. after exposure.						Explosion at a chemical plant released 2,3,7-8 TCDD.	Fanelli et al., 1980
	- Rana catesbeiana/ Central Arkansas	1984-85	10	Liver	Levels in females (n=7) ranged from 1.2 - 48.0 ppb, wet wt. Levels in males (n=3) ranged from 0.64 - 11.0 ppb, wet wt.							Kerfoocher et al., 1984b
	- Rana catesbeiana/ Central Arkansas	1984-85	6		Residues (ppt ± 95% confidence limits):						Results suggest frogs tolerate substantial body burden of 2,3,7,8-TCDD.	Kerfoocher et al., 1984a
				Sex	F	F	F	F	M	M		
				Muscle	218 ± 60	277 ± 60 229 ± 61	87 ± 26	183 ± 17	437 ± 202	566 ± 73		
				Skin	366 ± 66	423 ± 105 661 ± 134	217 ± 34	437 ± 188	1710 ± 834 1598 ± 426	- -		
				Oviduct	988 ± 488 1188 ± 156 *	696 ± 57	148 ± 22	2050 ± 197	-	-		
				Liver	2398 ± 279	3488 ± 566	1250 ± 176	1268 ± 122	1130 ± 2288	9800 ± 983		
				Ovaries	7468 ± 1468	10488 ± 5778	2838 ± 998	2788 ± 348	-	-		
				Fat	-	-	-	68888 ± 7128	> 14888	-		

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]				LEVELS USED/REMARKS	REFERENCE
2,3,7,8 - TCDD (cont'd)	- Southern toad/ 3.8 km^2 area on Eglin Air Force Base, Florida	1973-79		Whole body	Residue (ppt): 1,368.				Approx. 2.8 kg applied as contaminant in aerial spraying of 2,4,5-T.	Young and Cockerham, 1982
TDE (DC)	- Oxbow lakes in Louisiana Rana catesbeiana R. septentrionalis R. p. clamitans	1980	5 3 3		Not detected in all three species.				Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 150 cm.	Pfathenauer et al., 1984
Toxaphene (DC)	- Oxbow lakes in Louisiana	1980			Lake Providence		Lake Bruin		Lakes in fertile alluvial cropland. Subjected to heavy pesticide applications. Annual rainfall - 150 cm.	Pfathenauer et al., 1984
					Residues (geometric mean)	% lipid	Residues (geometric mean)	% lipid		
	Rana catesbeiana R. septentrionalis R. p. clamitans		5 3 3		0.09 0.21 ND*	1.7 3.3 1.1	ND ND ND	0.4 1.1 0.4		
					* Not detected.					

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Zinc (M)	- Triturus vulgaris Rana esculenta R. temporaria Pelobates fuscus Bufo viridis /Poland	June 1975	8 1 11	Whole body	Location: Protected zone near a zinc mill — Residues (ppm dry weight) range in brackets: 190.8 (154.0-205.6) Pine forest near a zinc mill: 533.5 Control area: 282.2 (103.7-301.4)	Some evidence of food chain accumulation.	Dmowski and Karolewski, 1979
	-Rana perezi/Donana Natl. Park, Spain		10 composite samples	"A fillet with bones, skin"	Geometric mean: 18.80.		Hernandez et al., 1987
	- Rana perezi/Donana Natl. Park, Spain	1984 1985 1986	7 4 2	Muscle	9.46 7.65 31.87		Rico et al., 1987
	- Rana esculenta/ South Moravia, Czechoslovakia	1982-84		Whole body	Mean zinc content (ppm dry wt.):		Pavel and Kucera, 1986

Locality	Years: 1982	1983	1984	
Brno	181.7 ± 12.8 (10)*	157.4 ± 11.8 (8)		Brno: suburban gardens, fields, lakes.
Musov	151.8 ± 6.0 (12)	173.0 ± 8.1 (12)	142.9 ± 9.1 (10)	Musov: large water reservoir.
Lednice	128.1 ± 9.1 (13)	177.8 ± 10.4 (8)	-	Lednice: abandoned sand pit.

* Number of samples.

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Zinc (cont'd)	- Triturus cristatus/lab		10 expti. 20 ctrl.s 8 samples analyzed/ organ		Concentrations of Zn (ug/mg dry wt):	Newts left in tank with zinc-plated bottom.	Taban et al., 1982

Tissue	Controls		Poisoned	
Anterior brain	.1531	.0505	.3254	.0785
	.0579	.1619	.1502	.1958
Posterior brain	.4326	.1012	.5482	.1020
	.0714	.2581	.9000	.2848
Liver	.1135	.0434	.3333	.0829
	.0793	.0952	.1161	.2389
Intestine	.4646	.1190	.5373	.1681
	.2500	.3710	.4762	.4564
Kidney	.2920	.1523	3.1405	.2647
	.2469	.1608	.5072	.4742
Pancreas	3.3918	.7059	2.6961	3.5593
	3.4186	.9231	3.2368	1.9192
Skin	.6579	.6818	1.5714	.3968
	.8296	1.5975	.7042	1.2575
Cheese	5.1282	.2961	2.1533	.6872
	1.0938	1.8142	.9067	1.1644

Concentrations in controls were .26 mg/L in water of control tank - .20 to .26 mg/L in water of experimental tank approx. .3 to 2.8 mg/L. Zinc-rich, unusual cells present in primordium hippocampi of poisoned newts.

	- Tanytarsus gregarius/lab			Whole body	Equilibrium body burden attained by 30 days. Two distinct loss components evident - slower component had half-life of 3 1/3 years and accounted for over 90% of initial body burden. Half-life following injection at room temp. was 1½ years. Highest concentrations in skin, muscle, blood, liver.	Uptake of ^{65}Zn from water and loss from body after transfer to clean water at 10°C studied. Excretion was directly related to ambient temp.	Willis and Valeti, 1978

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Zinc (cont'd)	- Unspecified species/ New Lead Belt, southeastern Missouri	Sept. 1982	8 pools 1 pool	Tadpoles	Area: Strother Creek; Control Residues (ppm dry wt.): 160-265; 42	Dam across Strother Creek provides the principal tailings pond for mine and mill effluents. Samples taken from below dam.	Gale et al., 1973
	- Unspecified species/ New Lead Belt, southeastern Missouri		1 pool 2 pools 1 pool 2 pools	Tadpoles: Whole body Eviscerated Liver & Heart Intestine	2808 ppm. 240-256 ppm. 47 ppm. 6926-4696 ppm.	Intestine included contents.	Jennett et al., 1979
	- Patuxent Wildlife Research Centre Rana catesbeiana R. clamitans Bufo spp. Rana spp.		5 pools 2 pools 6 pool of 10 pool of 10 11	Tadpoles Tadpoles Whole body Carcass Kidney Whole body	9.7-15 3.7-6.8 23-94 84 39 6.2-31		Hall and Mulhern, 1984
	- Rana temporaria/ Northern Finland	1971-1972	106	Liver male female	Mean ± SE (ppm dry wt.): 45.0 ± 3.0 (pre-emerging) - 90.9 ± 7.1 (feeding) 61.0 ± 5.1 (pre-emerging) - 88.8 ± 2.9 (spawning)	Pre-emerging: April; feeding: June - August; spawning: May.	Paasivuo and Koskela, 1974

Table 4 - Residues (cont'd)

CONTAMINANT[a]	SPECIES/LOCATION	DATE	N	TISSUE	RESIDUE LEVEL[b]	LEVELS USED/REMARKS	REFERENCE
Zinc (cont'd)	- Xenopus laevis/lab		10 per dose	Liver Kidney	Concentrations (mean ± SD): Control: 15.8 ± 1.5; 17.3 ± 1.5; Zn: 39.8 ± 5.5*; 53.6 ± 6.3* * Significantly accumulated.	5 mg Zn/kg body wt. injected i.m. daily for 5 days; frogs killed 1 day after last injection.	Suzuki et al., 1983
	- Plethodon cinereus, P. glutinosus, Bufo americanus, B. woodhousei, Rana sylvatica/ oak forest 10 km upwind from 2 zinc smelters in eastern Pennsylvania	June - Oct., 1979	23	Whole body	Mean ± SE (ppm dry weight) for all species: 88 ± 8.7.		Beyer et al., 1985
	- Rana catesbeiana/ lead mining district, southeastern Missouri	1981-1982	15	Whole body	Residues (geometric mean, range in parentheses): Site A: 28.9 (15.5-28.4); Site B: 42.7* (29.8-64.5); Site C: 29.9* (28.5-37.5) * Significantly higher than site A.	Site A - upstream from lead belt, Sites B & C - downstream from tailings ponds.	Niethammer et al., 1985

[a] OC = organochlorine insecticide, OP = organophosphate insecticide, C = carbamate insecticide, PY = pyrethroid, H = herbicide, M = metal.

[b] Residues given in ppm wet weight unless otherwise indicated.